

Polysaccharides as wound healing agent: a mini review¹Rohini J., ²Wan Ezumi M.F. and ^{1,*}Rabeta M.S.¹Food Technology Divisions, School of Industrial Technology, Universiti Sains Malaysia, 11800 Minden, Penang, Malaysia²Biomedicine Programme, School of Health Sciences, Health Campus, Universiti Sains Malaysia, 16150 Kubang Kerian, Kelantan, Malaysia**Article history:**

Received: 3 August 2020

Received in revised form: 10 September 2020

Accepted: 30 September 2020

Available Online: 13

February 2021

Keywords:

Polysaccharides,

Wound healing,

Bioactive compound

DOI:[https://doi.org/10.26656/fr.2017.5\(2\).421](https://doi.org/10.26656/fr.2017.5(2).421)**Abstract**

Medicinal eukaryotes, such as plants and fungi, have prompted researchers to conduct extensive studies on their medicinal values for drug discovery. Current trends focus on bioactive compounds of medicinal plants to produce inventions in the medical and health fields. Among many bioactive compounds, polysaccharides attract attention because they are non-toxic and have no side effects. Polysaccharides have been widely used in food and pharmaceutical industries as a secondary ingredient for several decades. This paper reviewed the applications of polysaccharides as wound healing agents. Wounds can affect the patient's well-being, self-image, working capacity and independence. Research studies on different sources of polysaccharides by *in vitro* and *in vivo* model have been investigated. Based on the scientific evidence related to polysaccharides, this work will be a baseline study for future investigations in different fields. All literature was accessed through available electronic databases.

1. Introduction

Carbohydrate polymers which consist of many units of monosaccharides are known as polysaccharides (Lin *et al.*, 2009). Those long chains of monosaccharide units are connected by glycosidic linkages (Xie, Jin, Morris *et al.*, 2016). Polysaccharides can be extracted from numerous sources, such as animals, plants, microorganisms, and seaweeds (Gomez-Zavaglia *et al.*, 2019).

Among many sources, organic products capture more attention because they are innately less toxic and have fewer side effects than synthetic products (Xu *et al.*, 2008). Nowadays, many researchers have changed the focus of medicinal plant studies to further explore bioactive compounds of the plants (Xie, Tang, Jin *et al.*, 2016). Among numerous bioactive compounds, the polysaccharide has been the most studied. This polymer is non-cytotoxic and has beneficial pharmacological properties. The changes on trend focus and interest gain on polysaccharide research can be proven with the number of articles published (Xie, Tang, Jin *et al.*, 2016).

Studies on polysaccharides and its antioxidant (Jeddou *et al.*, 2016), hypoglycemic (Li *et al.*, 2006), anticancer (Huang, 2013), wound healing (Khamlue *et*

al., 2012), and antihypertensive (Kolsi *et al.*, 2016) properties were conducted. Moreover, journals on the polysaccharides' characteristics, such as lipid profile (Chen *et al.* 2010) and radio-protective effects (Li and Zhou, 2007), were published.

Every 30-second leg is lost due to diabetes in some part of the world (International Diabetes Federation, 2017). It shows the failure of current treatment methods of wound healing. Religious and vegetarian lifestyle choices may prohibit certain consumer groups from taking animal-based wound healing products (Boyer, 2013). Effective wound care from plant-based is needed because some people do not like the peculiar taste of snakehead fish and sea cucumber when taken orally, which are the popular animal-based products with wound healing properties. Hence, this review aims at providing some updated information on polysaccharides as wound healing agent for supporting the data on the effectiveness of wound care from plant-based. All literatures were accessed through available electronic databases.

2. Polysaccharides in wound healing

Plant polysaccharides are made up of long carbohydrate chains with either linear or branched structures. These polysaccharides build up 80% of the

*Corresponding author.

Email: rabeta@usm.my

compounds and each compound has a specific function in the cell, which mostly contribute to biological functions. Polysaccharides, which appear to have biological functions, have glycuronic acids, primarily D-galacturonic and D-glucuronic acids, in their structure (Ovodov, 2010).

In recent times, a number of research on the correlation of polysaccharides with wound healing activity have been published. Valizadeh *et al.* (2015) stated that some monosaccharide units, arabinose and tannin compounds, shorten wound healing duration by coagulating surface proteins. Besides, pectin was found to ease the wound healing process by providing a favourable environment for angiogenesis and epithelialization. On the other hand, Carvalho *et al.* (2014) suggested that mucilage could be a growth factor that increases collagen production to promote wound healing in a dose-dependent manner. Thus, this mini review will tackle the role of polysaccharide in the wound healing process.

3. Overview of wound healing process

Repair of the skin or other soft tissues, which happens after different injuries, is known as wound healing (Nayak and Pereira, 2006). This process entails hemostasis, inflammation, reepithelization, and remodelling phases, which are interconnected and overlapping with one another (Demirci *et al.*, 2014).

The phase which occurs right after injury is known as hemostasis. During this phase, platelets rush to the site of injury, and a clot begins to form. This event subsequently triggers the release of growth factors and cytokines, which act as promoters of the inflammatory phase. Entries of inflammatory cells, such as neutrophils, monocytes, and macrophages, increase tissue debridement and influx of growth factors, which are needed for wound healing (Lakshmi *et al.*, 2011). Although the presence of neutrophils at the site of wound area benefits the process by clearing the microbial contamination and cellular debris, neutrophils also leave a negative effect by releasing protease and reactive oxygen species (ROS). Monocytes in blood, which identify macrophages when they enter the wound site, are responsible for cleaning mechanism and pave a way to end the inflammation state. During this period, macrophage undergoes differentiation while in the repairing state by inducing the needed cells, such as keratinocytes and fibroblasts, as well as angiogenesis.

The process is then transited to the next phase (Guo and DiPietro, 2010). Proliferative phase takes over; this phase is characterized by epithelization, the formation of

blood vessels, formation of granulation tissue, and deposition of collagen. Continuing the sequence of previous phases, cell detachment, cell migration, and cell differentiation occurs, which are all initiated by transforming growth factor β (TGF- β), vascular endothelial growth factor (VEGF), and multiple cytokines (Gothai *et al.*, 2016). Formation of new blood vessels, known as angiogenesis, also provides nourishment for newly formed cells and tissues. As a final phase of wound healing, fibroblasts replication and collagen production are accelerated to gain 80% of the tensile strength of the uninjured tissue, which is called as remodelling phase (Velnar *et al.*, 2009).

4. Wound healing polysaccharides sources

4.1 *Aloe barbadensis* (Aloe vera)

This plant, which belongs to family Liliaceae, originated from South Africa and is very popular at subtropical and tropical climate areas, including the South USA (Radha and Laxmipriya, 2015). Traditionally, Aloe vera was the preferred medication for wounds, burns, or infections. These medicinal purposes have attracted researchers to study more about Aloe vera properties (Sandhya and Gowri, 2015). Shi *et al.* (2017) have conducted the extraction and fractionation of Aloe vera leaf skin polysaccharides. The plant contained a huge amount of monosaccharide molecules, particularly acetylated glucomannan. Several studies were done on the wound healing activity of Aloe vera and proved that the possible effect was due to the presence of glucomannan polysaccharides (Hashemi *et al.*, 2015).

Although studies on Aloe vera alone proved to have effective wound healing properties, the latest attempt on combining Aloe vera with other plants was done to further examine its wound healing activity. The main aim of the study was not to isolate any particular compound but rather to study the effect of the formulations on fibroblast cell and main cell at all phases of the wound healing process (Negahdari *et al.*, 2017).

4.2 *Phellinus gilvus* (Mustard-yellow polypore)

Mushrooms have become a well-used natural source in the pharmaceutical industry and have been recognized as a functional food (Chang *et al.*, 2008). Among various genera of *Phellinus*, *P. gilvus* is in demand because it only requires a short duration for growth, which makes it more affordable and easier to produce than others (Park *et al.*, 2009). Bae *et al.* (2005) conducted a pharmacological study to determine the wound healing effect of *P. gilvus* polysaccharide in streptozocin-induced diabetic rat. The findings showed that the polysaccharides extract of the plant has a positive effect

in wound healing at different doses, which might be due to the antioxidant activity. Moreover, free radical damages the cells and prolongs the wound healing process. Chang *et al.* (2008) stated that *P. gilvus* increased scavenging activities more as compared with the standard compound, ascorbic acid. The wound-healing effect might be due to the anti-inflammatory properties of mushrooms. Inflammation is one of the stages in the wound healing process; thus, inhibiting the phase would fasten the healing activity (Kim *et al.*, 2011).

4.3 *Tremella fuciformis* (White jelly mushrooms)

Tremella fuciformis, commonly known as white jelly mushrooms, “snow fungus”, or “silver ear”, is a member of *Tremellaceae* family and is widely found in subtropical areas, especially in China (Wen *et al.*, 2016). This mushroom has unique physical characteristics: gelatinous, smooth, and elastic when wet but rigid and curved when dry. Since this species is edible, this mushroom has been traditionally used as a basic ingredient in food formulation, such as ice cream, porridge, soup, and drinks (Permana and Purnawan, 2015). Aside from the traditional uses, polysaccharide of this mushroom possesses many biological activities because the chemical structure of the polysaccharides comprises (1→3)- β -D-glucans and (1→3)- α -mannan backbone with small xylose- and glucuronic acid-containing side chains (Bin, 2010). Furthermore, Khamlue *et al.* (2012) have conducted wound healing study using porcine skin wound model ex-vivo. Two different types of mushrooms were used in the study, and both purified polysaccharide extracts had significantly promoted wound healing activities at different doses. The inconsistent results between different plants may be due to the different monosaccharide compositions in the plant polysaccharides. Collagen formation is an important factor in the remodeling phase.

According to Wen *et al.* (2016), *T. fuciformis* polysaccharide treated group enhanced collagen production up to 26%. Increased production of collagen will help the wound to heal. Moreover, the wound healing potentials may also due to the antioxidant activity of the mushroom polysaccharides. As mention earlier, free radicals damage cell epithelization, thereby prolonging the wound healing process. Bin (2010) studied the antioxidant properties of *T. fuciformis* polysaccharides and revealed that it can scavenge those free radicals at 78% in a 0.2 mg/mL solution.

4.4 *Opuntia ficus-indica* (Barbary fig)

Another widely used medicinal plant is *Opuntia ficus-indica* from the family of Cactaceae; it is a native plant

of South America and also available in tropical and subtropical regions (Zhong *et al.*, 2010). A number of researchers have compared the composition of the plants, and the compositions were different even though the species are the same. The results varied possibly because of the different methods of cultivation (Matsuhiro *et al.*, 2006). Therefore, the main component of the plant is similar to pectins, which are available from other species (Majdoub *et al.*, 2001). Moreover, a study on wound healing activities of two types of lyophilized polysaccharides extract was evaluated by Trombetta *et al.* (2006) using an animal model. The results obtained suggested that plant polysaccharides with a molecular weight greater than 10^4 Da had effective wound healing properties in rat model wounds by hastening the reepithelization and remodelling phases. Similarly, a wound healing study done by Galati *et al.* (2003) confirmed that the polysaccharide fractions from the plant significantly speeded up the healing process by reducing the inflamed cell and stimulating fibroblast, angiogenesis, and collagen production. Both findings complement each other. The isolated polysaccharides acted on the proliferation and remodelling phases.

4.5 *Althaea officinalis*

Marshmallow, scientifically known as *Althaea officinalis*, is a member of family Malvaceae and is commonly found in Europe (Deters *et al.*, 2010). Traditionally, this plant is used in treatments for respiratory disorders, such as asthma, bronchitis, and sore throat (Ghavi, 2015). Benbassat *et al.* (2013) stated that the roots of the plant contain 5% to 10% of water-soluble polysaccharides and other phenolic compounds. Additionally, Valizadeh *et al.* (2015) have taken the initiative to study the wound healing effects in rabbits. The outcome showed that the wounds which were treated with the different concentration of flower extract healed within 14 days. However, the control group took 21 days to heal completely. The flower polysaccharides positively influenced the wound healing activities on the rabbit model. This possible mechanism is due to the enhancement of epithelization process by the polysaccharides.

Furthermore, Kardošová and Machová (2006) found that *A. officinalis* possesses higher antioxidant activity (30%) than the standard compound, α -tocopherol. Inhibition of microbial growth shortens the inflammatory stage and fastens the healing process (Valiei *et al.*, 2011); hence, the antimicrobial activity of the flower extract plays a role in this positive mechanism.

4.6 *Ganoderma lucidum*

Ganoderma lucidum is a well-known medicinal

mushroom from the family Polyporaceae. The fruiting bodies of this species are known in different names in different countries. In China, this plant is called “Lingzhi”, whereas, in Japan, it is known as “Reishi” (Zhonghui *et al.*, 2014). This mushroom is rich in bioactive secondary metabolites with a vast range of chemical structures (Veljović *et al.*, 2017). Although it contains many bioactive compounds, polysaccharides of this fungi are available in high amount (Zhang and Lin, 2004). Additionally, Cheng *et al.* (2013) have studied the wound healing properties of the *G. lucidum* polysaccharides on diabetic rats.

Polysaccharides application in aqueous cream significantly shortened the wound healing process in streptozocin-induced rats, possibly due to the increased antioxidant effect of the extract, which inhibited the cell damages. Similarly, Jia *et al.* (2009) reported that polysaccharides from *G. lucidum* naturally had an antioxidant activity that inhibited the oxidative stress occurring on plasma and liver of streptozocin-induced diabetic rats. In addition, Barbieri *et al.* (2017) stated that the polysaccharides of this plant exhibited anti-inflammatory properties by activating the cytokines associated with an inflammation response. This characteristic is essential in the wound healing process; delayed inflammatory phase will result in the delay of the whole process of healing.

5. Proposed mechanism of polysaccharide in wound healing process

ROS were formed as a consequences of neutrophil release to the site of injury (Guo and DiPietro, 2010). An unstable atomic or molecular structure with imbalance electron arrangements is known as free radical; this molecule plays a negative role in various fields, such as food science, pharmacology, and physiology (Köksal *et al.*, 2017). During tissue metabolism, ROS are formed and maintained in low levels by the redox reaction. However, imbalance in redox reactions leads to oxidative stress that interrupts the physiology of cells and tissues (Cao *et al.*, 2017).

Generally, antioxidants from herbal compounds work as scavengers of free radicals. Through scavenging activity, antioxidant indirectly promotes faster healing mechanism by reducing inflammation, the formation of tissue, reepithelization, and differentiation of skin layers (Ktari *et al.* 2017). Song *et al.* (2017) also stated that the phenolic compounds, such as caffeic acid, which possesses antioxidant activity, regulate TGF- β . TGF- β with VEGF accelerates wound healing process by inducing angiogenesis, the formation of granulation tissue, and synthesis of collagen.

Apart from antioxidant activity, anti-inflammatory properties also help in the wound healing process. A defensive approach by an organism or tissue to clear the site of injury is known as inflammation. Many inflammatory promoters were released during this condition. The main targets of this stage are to prevent microbial infestation and to remove cellular debris through phagocytosis mechanism (Hadagali and Chua, 2014). During the inflammatory phase, macrophages released to the wound are supposed to clear the neutrophils and other foreign matter. Afterwards, macrophages undergo apoptosis and stimulate the needed functions for the next proliferative phase.

6. Conclusion

Plant based-polysaccharides have attracted researchers to study their therapeutic value because of the nature of the polysaccharides. Plant polysaccharides are less toxic, biodegradable and biocompatible. Plant-based wound healing may cause minimal adverse effects. Although previously plant polysaccharides have been widely used in pharmaceutical and food industry as a stabilizer, thickener and dissolving agent, it will be an added advantage if polysaccharides from plants have been identified for its biological activity so that they can act as a functional food in both industries.

Conflict of interest

The authors declare no conflict of interest

Acknowledgement

This work was funded by a RU grant (1001/PTEKIND/ 812176) from Universiti Sains Malaysia.

References

- Bae, J.-S., Jang, K.-H. and Jin, H.-K. (2005). Polysaccharides isolated from *Phellinus gilvus* enhances dermal wound healing in streptozotocin-induced diabetic rats. *Journal of Veterinary Science*, 6(2), 161-164. <https://doi.org/10.4142/jvs.2005.6.2.161>
- Barbieri, A., Quagliariello, V., Del Vecchio, V., Falco, M., Luciano A, Amruthraj, N.J. and Arra, C. (2017). Anticancer and Anti-Inflammatory Properties of *Ganoderma lucidum* Extract Effects on Melanoma and Triple-Negative Breast Cancer Treatment. *Nutrients*, 9(3), 210. <https://doi.org/10.3390/nu9030210>
- Benbassat, N., Kostova, B., Nikolova, I. and Rachev, D. (2013). Development and evaluation of novel lozenges containing marshmallow root extract. *Pakistan Journal*

- of *Pharmaceutical Sciences*, 26, 1103-1107.
- Bin, C. (2010). Optimization of extraction of *Tremella fuciformis* polysaccharides and its antioxidant and antitumour activities *in vitro*. *Carbohydrate Polymers*, 81(2), 420-424. <https://doi.org/10.1016/j.carbpol.2010.02.039>
- Boyer, D. (2013). Cultural Considerations in Advanced Wound Care. *Advances in Skin and Wound Care*, 26 (3), 110-111. <https://doi.org/10.1097/01.ASW.0000427923.20121.2d>
- Cao, X.-P., Chen, Y. F., Zhang, J.-L., You, M.-M., Wang, K. and Hu, F.-L. (2017). Mechanisms underlying the wound healing potential of propolis based on its *in vitro* antioxidant activity. *Phytomedicine*, 34, 76-84. <https://doi.org/10.1016/j.phymed.2017.06.001>
- Carvalho, E.G., Soares, C.P., Blau, L., Menegon, R.F. and Joaquim, W.M. (2014). Wound healing properties and mucilage content of *Pereskia aculeata* from different substrates. *Revista Brasileira de Farmacognosia*, 24(6), 677-682. <https://doi.org/10.1016/j.bjp.2014.11.008>
- Chang, Z.Q., Hwang, M.H., Rhee, M.H., Kim, K.S., Kim, J.C., Lee, S.P. and Park, S.C. (2008). The *in vitro* anti-platelet, antioxidant and cellular immunity activity of *Phellinus gilvus* fractional extracts. *World Journal of Microbiology and Biotechnology*, 24(2), 181-187. <https://doi.org/10.1007/s11274-007-9454-z>
- Chen, H., Liu, L.J., Zhu, J.J., Xu, B. and Li, R. (2010). Effect of soybean oligosaccharides on blood lipid, glucose levels and antioxidant enzymes activity in high fat rats. *Food Chemistry*, 119(4), 1633-1636. <https://doi.org/10.1016/j.foodchem.2009.09.056>
- Cheng, P.G., Phan, C.W., Sabaratnam, V., Abdullah, N., Abdulla, M.A. and Kuppusamy, U.R. (2013). Polysaccharides-rich extract of *Ganoderma lucidum* (MA Curtis: Fr.) P. Karst accelerates wound healing in streptozotocin-induced diabetic rats. *Evidence-Based Complementary and Alternative Medicine*, 2013, 671252. <https://doi.org/10.1155/2013/671252>
- Demirci, S., Dogan, A., Demirci, Y. and Sahin, F. (2014). *In vitro* wound healing activity of methanol extract of *Verbascum speciosum*. *International Journal of Applied Research in Natural Products*, 7 (3), 37-44.
- Deters, A., Zippel, J., Hellenbrand, N., Pappai, D., Possemeyer, C. and Hensel, A. (2010). Aqueous extracts and polysaccharides from Marshmallow roots (*Althea officinalis* L.): Cellular internalisation and stimulation of cell physiology of human epithelial cells *in vitro*. *Journal of Ethnopharmacology*, 127(1), 62-69. <https://doi.org/10.1016/j.jep.2009.09.050>
- Galati, E.M., Mondello, M.R., Monforte, M.T., Galluzzo, M., Miceli, N. and Tripodo, M.M. (2003). Effect of *Opuntia ficus-indica* (L.) Mill. cladodes in the wound-healing process. *Journal of the Professional Association for Cactus Development*, 5, 1-16.
- Ghavi, P.P. (2015). The extraction process optimization of antioxidant polysaccharides from Marshmallow (*Althaea officinalis* L.) roots. *International Journal of Biological Macromolecules*, 75, 51-57. <https://doi.org/10.1016/j.ijbiomac.2014.11.047>
- Gomez-Zavaglia, A., Lage, M.A.P., Jimenez-Lopez, C., Mejuto, J.C. and Simal-Gandara, J. (2019). The Potential of Seaweeds as a Source of Functional Ingredients of Prebiotic and Antioxidant Value. *Antioxidants*, 8, 406. <https://doi.org/10.3390/antiox8090406>
- Gothai, S., Arulselvan, P., Tan, W.S. and Fakurazi, S. (2016). Wound healing properties of ethyl acetate fraction of *Moringa oleifera* in normal human dermal fibroblasts. *Journal of Intercultural Ethnopharmacology*, 5(1), 1-6. <https://doi.org/10.5455/jice.20160201055629>
- Guo, S.A. and DiPietro, L.A. (2010). Factors affecting wound healing. *Journal of Dental Research*, 89(3), 219-229. <https://doi.org/10.1177/0022034509359125>
- Hadagali, M.D. and Chua, L.S. (2014). The anti-inflammatory and wound healing properties of honey. *European Food Research and Technology*, 239(6), 1003-1014. <https://doi.org/10.1007/s00217-014-2297-6>
- Hashemi, S.A., Madani, S.A. and Abediankenari, S. (2015). The review on properties of *Aloe vera* in healing of cutaneous wounds. *BioMed Research International*, 2015, 714216. <https://doi.org/10.1155/2015/714216>
- Huang, W. (2013). Anticancer effect of plant-derived polysaccharides on mice. *Journal of Cancer Therapy*, 4, 500-503. <https://doi.org/10.4236/jct.2013.42061>
- International Diabetes Federation. (2017). IDF Diabetes Atlas, 8th. Retrieved from IDF website: <https://www.idf.org/e-library/epidemiology-research/diabetes-atlas/134-idf-diabetes-atlas-8th-edition.html>
- Jeddou, K.B., Chaari, F., Maktouf, S., Nouri-Ellouz, O., Helbert, C.B. and Ghorbel, R.E. (2016). Structural, functional, and antioxidant properties of water-soluble polysaccharides from potatoes peels. *Food Chemistry*, 205, 97-105. <https://doi.org/10.1016/j.foodchem.2016.02.108>
- Kardošová, A. and Machova, E. (2006). Antioxidant

- activity of medicinal plant polysaccharides. *Fitoterapia*, 77(5), 367-373. <https://doi.org/10.1016/j.fitote.2006.05.001>
- Khamlue, R., Naksupan, N., Ounaron, A. and Saelim, N. (2012). Skin wound healing promoting effect of polysaccharides extracts from *Tremella fuciformis* and *Auricularia auricula* on the ex-vivo porcine skin wound healing model. *International Proceedings of Chemical, Biological and Environmental Engineering*, 43, 93-98. <https://doi.org/10.7763/ipcbee>
- Köksal, E., Bursal, E., Gülçin, İ., Korkmaz, M., Çağlayan, C., Gören, A.C. and Alwasel, S.H. (2017). Antioxidant activity and polyphenol content of Turkish thyme (*Thymus vulgaris*) monitored by liquid chromatography and tandem mass spectrometry. *International Journal of Food Properties*, 20(3), 514-525. <https://doi.org/10.1080/10942912.2016.1168438>
- Kolsi, R.B.A., Fakhfakh, J., Krichen, F., Jribi, I., Chiarore, A., Patti, F.P. and Belghith, K. (2016). Structural characterization and functional properties of antihypertensive *Cymodocea nodosa* sulfated polysaccharide. *Carbohydrate Polymers*, 151, 511-522. <https://doi.org/10.1016/j.carbpol.2016.05.098>
- Kim, S.H., Lim, J.H., Moon, C., Park, S.H., Kim, S.H., Shin, D.H. and Kim, J.C. (2011). Antiinflammatory and antioxidant effects of aqueous extract from *Phellinus gilvus* in rats. *Journal of Health Science*, 57(2), 171-176. <https://doi.org/10.1248/jhs.57.171>
- Ktari, N., Trabelsi, I., Bardaa, S., Triki, M., Bkhairia, I., Salem, R.B.S.B. and Salah, R.B. (2017). Antioxidant and hemolytic activities, and effects in rat cutaneous wound healing of a novel polysaccharide from fenugreek (*Trigonella foenum-graecum*) seeds. *International Journal of Biological Macromolecules*, 95, 625-634. <https://doi.org/10.1016/j.ijbiomac.2016.11.091>
- Lakshmi, A., Rao, Y.M., Bhargavi, C. and Seelam, U. (2011). Antidiabetic and wound healing activity of various bark extracts of *Polyalthia longifolia*. *Asian Journal of Pharmaceutical and Clinical Research*, 4 (1), 109-113.
- Li, S.P., Zhang, G.H., Zeng, Q., Huang, Z.G., Wang, Y.T., Dong, T.T.X. and Tsim, K.W.K. (2006). Hypoglycemic activity of polysaccharide, with antioxidation, isolated from cultured *Cordyceps mycelia*. *Phytomedicine*, 13(6), 428-433. <https://doi.org/10.1016/j.phymed.2005.02.002>
- Li, X. and Zhou, A. (2007). Preparation of polysaccharides from *Acanthopanax senticosus* and its inhibition against irradiation-induced injury of rat. *Carbohydrate polymers*, 67(2), 219-226. <https://doi.org/10.1016/j.carbpol.2006.05.008>
- Lin, C.L., Wang, C.C., Chang, S.C., Inbaraj, B.S. and Chen, B.H. (2009). Antioxidative activity of polysaccharide fractions isolated from *Lycium barbarum* Linnaeus. *International Journal of Biological Macromolecules*, 45(2), 146-151.
- Majdoub, H., Roudesli, S. and Deratani, A. (2001). Polysaccharides from prickly pear peel and nopals of *Opuntia ficus-indica*: extraction, characterization and polyelectrolyte behaviour. *Polymer International*, 50 (5), 552-560. <https://doi.org/10.1002/pi.665>
- Matsuhiro, B., Lillo, L.E., Sáenz, C., Urzúa, C.C. and Zárata, O. (2006). Chemical characterization of the mucilage from fruits of *Opuntia ficus indica*. *Carbohydrate Polymers*, 63(2), 263-267. <https://doi.org/10.1016/j.carbpol.2005.08.062>
- Nayak, B.S. and Pereira, L.M.P. (2006). *Catharanthus roseus* flower extract has wound-healing activity in Sprague Dawley rats. *BMC Complementary and Alternative Medicine*, 6(1), 41. <https://doi.org/10.1186/1472-6882-6-41>
- Negahdari, S., Galehdari, H., Kesmati, M., Rezaie, A. and Shariati, G. (2017). Wound healing activity of extracts and formulations of *Aloe vera*, henna, adiantum capillus-veneris, and myrrh on mouse dermal fibroblast cells. *International Journal of Preventive Medicine*, 10(8), 18. https://doi.org/10.4103/ijpvm.IJPVM_338_16
- Ovodov, Y.S. (2010). Bioglycans and natural glycosides as a promising research topic in bioorganic chemistry. *Acta Naturae*, 2(2), 28-36. <https://doi.org/10.32607/20758251-2010-2-2-28-35>
- Park, S.C., Cheon, Y.P., Son, W.Y., Rhee, M.H., Kim, T.W., Song, J.C. and Kim, K.S. (2009). Hepatoprotective Effects of Polysaccharides Isolated from *Phellinus gilvus* Against Carbon Tetrachloride-induced Liver Injury in Rats. *Toxicological Research*, 25(1), 29-33. <https://doi.org/10.5487/TR.2009.25.1.029>
- Permana, D.R. and Purnawan, A. (2016). Characteristics of Jelly Fungus (*Tremella fuciformis*, Berk.) As an Edible Mushroom presented at Seminar Nasional XII Pendidikan Biologi FKIP UNS, November 2015, p. 849-854. Surakarta, Indonesia: Sebelas Maret University.
- Radha, M.H. and Laxmipriya, N.P. (2015). Evaluation of biological properties and clinical effectiveness of *Aloe vera*: A systematic review. *Journal of Traditional and Complementary Medicine*, 5(1), 21-26. <https://doi.org/10.1016/j.jtcme.2014.10.006>
- Sandhya and Gowri. (2015). Effects of *Aloe Vera* on Skin and on Wound Healing - A Review.

- International Journal of Science and Research*, 6(4), 1629-1631.
- Shi, X.D., Nie, S.P., Yin, J.Y., Que, Z.Q., Zhang, L.J. and Huang, X.J. (2017). Polysaccharide from leaf skin of *Aloe barbadensis* Miller: Part I. Extraction, fractionation, physicochemical properties and structural characterization. *Food Hydrocolloids*, 73, 176-183. <https://doi.org/10.1016/j.foodhyd.2017.06.039>
- Song, Y., Zeng, R., Hu, L., Maffucci, K.G., Ren, X. and Qu, Y. (2017). *In vivo* wound healing and *in vitro* antioxidant activities of *Bletilla striata* phenolic extracts. *Biomedicine and Pharmacotherapy*, 93, 451-461. <https://doi.org/10.1016/j.biopha.2017.06.079>
- Trombetta, D., Puglia, C., Perri, D., Licata, A., Pergolizzi, S., Lauriano, E.R. and Bonina, F.P. (2006). Effect of polysaccharides from *Opuntia ficus-indica* (L.) cladodes on the healing of dermal wounds in the rat. *Phytomedicine*, 13(5), 352-358. <https://doi.org/10.1016/j.phymed.2005.06.006>
- Valiei, M., Shafaghat, A. and Salimi, F. (2011). Chemical composition and antimicrobial activity of the flower and root hexane extracts of *Althaea officinalis* in Northwest Iran. *Journal of Medicinal Plants Research*, 5(32), 6972-6976. <https://doi.org/10.5897/JMPR11.963>
- Valizadeh, R., Hemmati, A.A., Houshmand, G., Bayat, S. and Bahadoram, M. (2015). Wound healing potential of *Althaea officinalis* flower mucilage in rabbit full-thickness wounds. *Asian Pacific Journal of Tropical Biomedicine*, 5(11), 937-943. <https://doi.org/10.1016/j.apjtb.2015.07.018>
- Veljović, S., Veljović, M., Nikićević, N., Despotović, S., Radulović, S., Nikšić, M. and Filipović, L. (2017). Chemical composition, antiproliferative and antioxidant activity of differently processed *Ganoderma lucidum* ethanol extracts. *Journal of Food Science and Technology*, 54, 1312-1320. <https://doi.org/10.1007/s13197-017-2559-y>
- Velnar, T., Bailey, T. and Smrkolj, V. (2009). The wound healing process: an overview of the cellular and molecular mechanisms. *Journal of International Medical Research*, 37(5), 1528-1542. <https://doi.org/10.1177/147323000903700531>
- Wen, L., Gao, Q., Ma, C.W., Ge, Y., You, L., Liu, R.H. and Liu, D. (2016). Effect of polysaccharides from *Tremella fuciformis* on UV-induced photoaging. *Journal of Functional Foods*, 20, 400-410. <https://doi.org/10.1016/j.jff.2015.11.014>
- Xie, J.H., Jin, M.L., Morris, G.A., Zha, X.Q., Chen, H.Q., Yi, Y. and Shang, P. (2016). Advances on bioactive polysaccharides from medicinal plants. *Critical Reviews in Food Science and Nutrition*, 56(1), 60-84. <https://doi.org/10.1080/10408398.2015.1069255>
- Xie, J.H., Tang, W., Jin, M.L., Li, J.E. and Xie, M.Y. (2016). Recent advances in bioactive polysaccharides from *Lycium barbarum* L., *Zizyphus jujuba* Mill, *Plantago spp.*, and *Morus spp.*: Structures and functionalities. *Food Hydrocolloids*, 60, 148-160. <https://doi.org/10.1016/j.foodhyd.2016.03.030>
- Xu, C., HaiYan, Z., JianHong, Z. and Jing, G. (2008). The pharmacological effect of polysaccharides from *Lentinus edodes* on the oxidative status and expression of VCAM-1mRNA of thoracic aorta endothelial cell in high-fat-diet rats. *Carbohydrate Polymers*, 74(3), 445-450. <https://doi.org/10.1016/j.carbpol.2008.03.018>
- Zhang, H.N. and Lin, Z.B. (2004). Hypoglycemic effect of *Ganoderma lucidum* polysaccharides. *Acta Pharmacologica Sinica*, 25(2), 191-195. <https://doi.org/10.1007/s12272-012-1012-z>
- Zhong, X.K., Jin, X., Lai, F.Y., Lin, Q.S. and Jiang, J.G. (2010). Chemical analysis and antioxidant activities in vitro of polysaccharide extracted from *Opuntia ficus indica* Mill. cultivated in China. *Carbohydrate Polymers*, 82(3), 722-727. <https://doi.org/10.1016/j.carbpol.2010.05.042>
- Zhonghui, Z., Xiaowei, Z. and Fang, F. (2014). *Ganoderma lucidum* polysaccharides supplementation attenuates exercise-induced oxidative stress in skeletal muscle of mice. *Saudi Journal of Biological Sciences*, 21(2), 119-12. <https://doi.org/10.1016/j.sjbs.2013.04.004>