

Analysis of candlenut oil as oil adulterant in three functional oils of soybean oil, sunflower oil and grapeseed oil in quaternary mixture systems using FTIR spectroscopy and chemometrics

^{1,2}Riyanta, A.B., ¹Riyanto, S., ¹Lukitaningsih, E. and ^{1,3,*}Rohman, A.

¹Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Universitas Gadjah Mada, Yogyakarta

²Department of Pharmacy, Politeknik Harapan Bersama, Central Java 52147 Indonesia

³Institute of Halal Industry and Systems, Universitas Gadjah Mada, Yogyakarta 55281 Indonesia

Article history:

Received: 15 August 2020

Received in revised form: 15 September 2020

Accepted: 28 October 2020

Available Online: 21 March 2021

Keywords:

FTIR spectroscopy,
Soybean candlenut oil,
Quaternary mixture,
Multivariate calibrations

DOI:

[https://doi.org/10.26656/fr.2017.5\(2\).443](https://doi.org/10.26656/fr.2017.5(2).443)

Abstract

Soybean oil (SBO), sunflower oil (SFO) and grapeseed oil (GPO) contain high levels of unsaturated fats that are good for health and have proximity to candlenut oil. Candlenut oil (CNO) has a lower price and easier to get oil from that seeds than other seed oils, so it is used as adulteration for gains. Therefore, authentication is required to ensure the purity of oils by proper analysis. This research was aimed to highlight the FTIR spectroscopy application with multivariate calibration is a potential analysis for scanning the quaternary mixture of CNO, SBO, SFO and GPO. CNO quantification was performed using multivariate calibrations of principle component (PCR) regression and partial least (PLS) square to predict the model from the optimization FTIR spectra regions. The highest R^2 and the lowest values of root mean square error of calibration (RMSEC) and root mean square error of prediction (RMSEP) were used as the basis for selection of multivariate calibrations created using several wavenumbers region of FTIR spectra. Wavenumbers regions of 4000-650 cm^{-1} from the second derivative FTIR-ATR spectra using PLS was used for quantitative analysis of CNO in quaternary mixture with SBO, SFO and GPO with R^2 calibration = 0.9942 and 0.0239% for RMSEC value and 0.0495%. So, it can be concluded the use of FTIR spectra combination with PLS is accurate to detect quaternary mixtures of CNO, SBO, SFO and GPO with the highest R^2 values and the lowest RMSEC and RMSEP values.

1. Introduction

Candlenut (*Aleurites mollucana* (L.) Willd.) oil is a plant that is able to produce oil up to 3200 kg/ha and it is able to produce 30-50% oil in every kilogram (Riyanta *et al.*, 2020a). The size of candlenut seeds is larger than soybean seeds, sunflower seeds and even grapeseed make candlenut seeds have higher oil potential. However, candlenut oil is more easily produced than other oils and has the same proximity characteristics as soybean oil, sunflower seed oil and grape seed oil. Extraction methods that have been used like pressing, maceration, soxhletation, sonication and using supercritical CO_2 (Siddique *et al.*, 2015; Aryati *et al.*, 2020; Riyanta *et al.*, 2020b)

Fourier transform infrared (FTIR) in combination with chemometrics of classification such as discriminant analysis and multivariate calibration of partial least square (PLS) and principle component regression has

been used for authentication analysis of raw milk (Coitinho *et al.*, 2017), motor oil adulteration (Bassbasi *et al.*, 2013), pumpkin seed oil (Irnawati *et al.*, 2019), olive oil (Nurwahidah *et al.*, 2019), sesame oil (Nurrulhidayah *et al.*, 2014), beef meatball (Rahayu *et al.*, 2018), and fish patin oil (Putri *et al.*, 2019). However, using literature review, there are no reports regarding the analysis candlenut oil (CNO) in the quaternary mixture from other oils using FTIR spectroscopy. However, it is often found that oil products are mixed with other oils to reduce production costs. Therefore, in this study, FTIR spectroscopy combined with chemometrics of multivariate calibrations was performed to analyze of candlenut oil in quaternary oil mixture.

2. Materials and methods

2.1 Materials

The candlenuts (*Aleurites mollucana* (L) Willd) were

*Corresponding author.

Email: abdulrohmanugm@gmail.com

collected from Bumiayu, Brebes, Central Java in February 2019. The candlenuts were collected from seed that has fallen from tree and seeds were opened from the shell by gouging out. The candlenut oils were extracted using apparatus oil press according to Riyanta *et al.* (2020a). The candlenut oil was collected in a brown bottle and stored in the freezer before used for analysis. For soybean oil, sunflower oil, and grapeseed oil were purchased from local markets in Yogyakarta, Indonesia.

2.2 Samples preparation for calibration and validation

Candlenut oils in a quaternary mixture of soybean oil, sunflower oil and grapeseed oil were prepared at concentrations in the range of 0-100.0% v/v. Samples were prepared randomly by Microsoft Office Excel (Inc. USA) and shown in Table 1. Then analyzed using FTIR spectrophotometer.

Table 1. The composition of candlenut oil (CNO) in quaternary mixtures with soybean oil (SBO), sunflower oil (SFO) and grapeseed oil (GPO)

No	Percentage (%) composition of oils			
	SBO	SFO	GSO	CNO
1	29.00	26.00	36.00	9.00
2	13.71	8.13	8.20	69.97
3	22.08	5.78	37.05	35.05
4	15.31	14.66	7.36	62.66
5	27.08	35.07	30.24	7.62
6	2.47	41.70	45.24	10.59
7	17.76	10.80	3.03	68.40
8	25.95	8.54	26.73	38.78
9	8.08	23.45	31.40	37.07
10	43.58	10.51	35.78	10.14
11	30.05	11.64	24.98	33.34
12	26.66	22.66	48.12	2.56
13	35.45	25.48	18.23	20.85
14	11.51	14.05	13.97	60.47
15	6.42	9.55	44.28	39.75
16	6.48	6.48	35.66	51.38
17	8.22	43.26	29.37	19.15
18	16.75	10.16	47.81	25.28
19	6.25	29.48	12.05	52.22
20	13.00	49.00	37.50	0.00

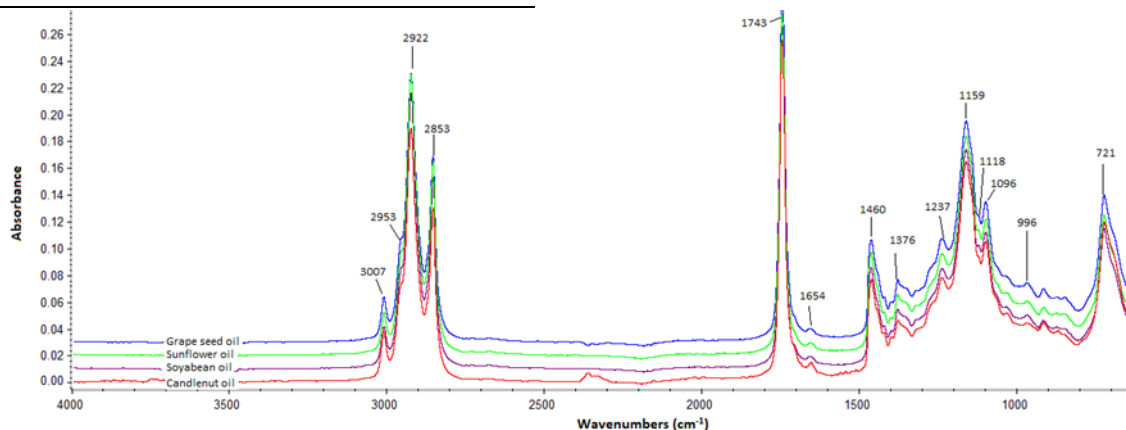


Figure 1. FTIR spectra of candlenut oil, soybean oil, sunflower oil and grape seed oil at mid-infrared region (4000-650 cm^{-1}) measured using attenuated total reflectance mode

2.3 FTIR measurement

Oil samples were scanning with FTIR spectrophotometer (Scientific Nicolet, Madison, WI) with Omnic software for FTIR spectra processing. Scans 4000-650 cm^{-1} of wavenumber with thirty-two scanning and for resolution applied 8 cm^{-1} of Horizontal Attenuated Total Reflectance at wavenumbers. As background were used air for all FTIR spectra and taken every scan for new reference scanning.

2.4 Chemometrics analysis

TQ Analyst software (version 9) by Thermo Fisher Scientific, Inc was used for chemometrics analysis multivariate calibration and discriminant analysis. Partial least square (PLS) regression and principle (PCR) component regression was used for quantification of oils.

3. Results and discussion

FTIR spectroscopy is effectively able to analyze mixed sample like the quaternary mixture of oils. Quantitative analysis can be done by processing FTIR spectra in the form of signals from analytic peak and shoulders to be used as dependent variables using a calibration model. For this research, attenuated total reflectance (ATR) was used as a sampling technique, in where the samples (without any addition of solvent) were directly placed into ATR crystal. The combination of FTIR and multivariate analysis was successfully modelled for the oil adulteration mixture (López-Díez *et al.*, 2003). Principle component (PCR) regression and partial least square (PLS) regression in combination with FTIR-ATR were more powerful technique for quantitative analysis (Rohman and Man, 2010; Irnawati *et al.*, 2020). Figure 1 shows the FTIR-ATR spectra in the regions of wavenumber from 4000-650 cm^{-1} for oil mixture spectra of CNO, SBO, SFO, and GPO.

Wavenumber 3010-2800 cm^{-1} at peak 3007 was stretching vibration of cis C=CH, while the peak at 2953

cm^{-1} from asymmetric stretching vibration of methyl ($-\text{CH}_3$) group. Asymmetric and symmetric stretching vibrations of methylene ($-\text{CH}_2$) showed by the peaks at 2922 and 2853 cm^{-1} . The wavenumber 1800-1600 cm^{-1} showed the stretching vibration from carbonyl ($\text{C}=\text{O}$) was observed at 1744 cm^{-1} , while the peak at 1654 is from $\text{C}=\text{C}$ stretching vibration. The wavenumber 1500-650 were fingerprinting area showed with bending vibrations of methylene and methyl were observed at wavenumbers of 1460 and 1376 cm^{-1} , respectively. The peaks at regions of 1237, 1160, 1118, 1098 were from C-O vibrations. While, peaks at 996 and 850 were due to bending out of plane vibrations of $-\text{HC}=\text{CH}-$ (trans) and $-\text{HC}=\text{CH}-$ (cis), respectively (Inarwati et al., 2020).

The analytical method of FTIR spectra for fingerprint can be used to detect for adulteration and these methods most ideal for detection and quantification of adulterants in mixed samples. Detection of CNO as an adulterant in quaternary mixture with soybean oil (SBO), sunflower oil (SFO), and grapeseed oil (GPO) was performed using optimized spectra regions of FTIR combined with multivariate calibrations. Principle component regression (PCR) and partial least square (PLS) were used as predictor models of principle components of oils mixture (Khudzaifi et al., 2020). Prediction models between actual values of CNO and FTIR predicted values were optimized by PLS and PCR from FTIR spectra. The highest coefficient of determination (R^2) and the lowest root mean square of calibration (RMSEC) and root mean square error of prediction (RMSEP) were used as parameters to be decided as the FTIR spectral area for multivariate calibration.

Table 2 tabulates the prediction of CNO levels as an adulterant in with SBO, SFO and GPO by FTIR

spectroscopy using different wavenumbers and spectral treatments combined with the multivariate calibration of PLS and PCR. Wavenumbers of 4000-650 cm^{-1} using second derivative FTIR spectra combined with PLS was selected for the quantification of CNO in SBO, SFO and GPO and able to provide the highest R^2 values (0.9942) either in calibration or prediction with low RMSEC value (0.0239) and RMSEP value (0.0495). High values of R^2 and low values of RMSEC and RMSEP were capable of developing models revealed the acceptable accuracy and precision. Figure 3A exhibits the linear relationship between actual values of CNO (x-axis) and FTIR predicted values of CNO (y-axis) using FTIR spectroscopy. From Figure 3B, the errors occurring during modelling occurred randomly (not systematic error) around zero difference. This indicated that the systematic errors did not exist and the developed model was reliable to predict CNO in mixture with SBO, SFO and GPO.

Discriminant analysis (DA) is known as the supervised pattern recognition commonly used for discrimination between pure CNO and CNO in the quaternary mixture (Riyanta et al., 2020a). The variables used for quantitative analysis were also used for DA. The Mahalanobis distances using these absorbances were then calculated to create Cooman's plot. Figure 4 shows The Cooman's plot obtained during discriminant analysis for discrimination of pure *Candlenut* Oil (CNO) and CNO mixed with SBO, SFO, and GPO.

4. Conclusion

Combination of Fourier transform infrared (FTIR) spectroscopy and multivariate calibration of partial least square regression (PLS) and discriminant are capable for quantifying the levels of candlenut oil as an adulterant in

Table 2. The results of FTIR spectroscopy using different wavenumbers and spectral treatments coupled with multivariate calibration intended for prediction of soybean oil, sunflower oil, grapeseed oil levels adulterated with candlenut oil (CNO)

	Wavenumber (cm^{-1})	Spectra	Calculation		Validation	
			R^2	RMSEC	R^2	RMSEP
PCR	4000 – 650	Normal	0.9884	0.0337	0.9788	0.0475
		Derivative 1	0.9868	0.0361	0.9809	0.0442
		Derivative 2	0.9878	0.0347	0.9722	0.0547
	3050 - 2800	Normal	0.9885	0.0336	0.9752	0.0522
		Derivative 1	0.9877	0.0348	0.9815	0.0449
		Derivative 2	0.9809	0.0432	0.9776	0.0465
	1800 – 650	Normal	0.9891	0.0327	0.9808	0.0446
		Derivative 1	0.9899	0.0315	0.9853	0.0384
		Derivative 2	0.9884	0.0338	0.9758	0.052
	3000 – 2800 and 1600 - 650	Normal	0.9889	0.033	0.981	0.0431
		Derivative 1	0.9879	0.0345	0.9812	0.0434
		Derivative 2	0.9851	0.0382	0.9749	0.054

The selected variables were marked with bold.

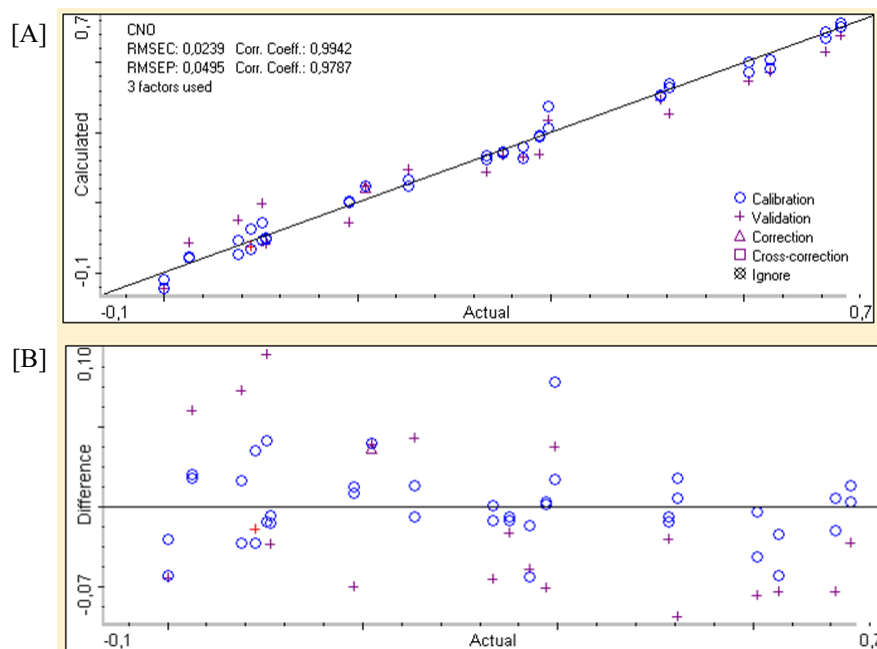


Figure 3. The relationship between actual values of Candlenut Oil (CNO) in x-axis and the predicted values of CNO in y-axis using FTIR spectroscopy [A] along with residual analysis [B]

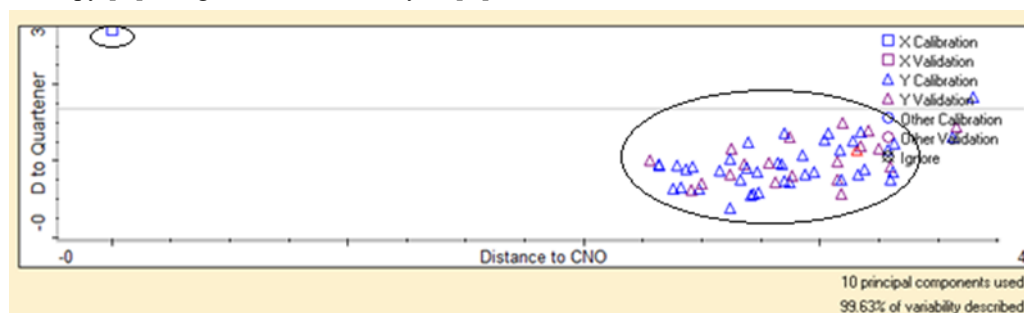


Figure 4. The Cooman's plot obtained during discriminant analysis for discrimination of pure Candlenut Oil (CNO) and CNO mixed with SBO, SFO, and GPO

soybean oil (SBO), sunflower oil (SFO) and grapeseed oil (GPO) in the quaternary mixture at optimized wavenumbers of 4000-650 cm^{-1} . The highest R^2 and lowest RMSEC and RMSEP model as acceptable accuracy and precision. Discriminant analysis can be used to classify CNO as an adulterant in soybean oil (SBO), sunflower oil (SFO), and grapeseed oil (GPO) mixtures.

Conflict of interest

We have no conflict of interest to declare.

Acknowledgement

The authors thank to Universitas Gadjah Mada (UGM), Yogyakarta Indonesia for financial support trough the scheme Rekognisi Tugas Akhir 2020 with contract number of 2129/UN1/DITLIT/DIT-LIT/LT/2020 (awarded to Professor Dr Abdul Rohman).

References

Aryati, W.D., Azka, K.M. and Mun'im, A. (2020). Ultrasonic-assisted extraction using a betaine-based

natural deep eutectic solvent for resveratrol extraction from melinjo (*Gnetum gnemon*) seeds. *International Journal of Applied Pharmaceutics*, 12 (Special Issue 1), 26–31. <https://doi.org/10.22159/ijap.2020.v12s1.FF001>

Bansal, S., Singh, A., Mangal, M., Mangal, A.K. and Kumar, S. (2017). Food adulteration: Sources, health risks, and detection methods. *Critical Reviews in Food Science and Nutrition*, 57(6), 1174–1189. <https://doi.org/10.1080/10408398.2014.967834>

Bassbasi, M., Hafid, A., Platikanov, S., Tauler, R. and Oussama, A. (2013). Study of motor oil adulteration by infrared spectroscopy and chemometrics methods. *Fuel*, 104, 798–804. <https://doi.org/10.1016/j.fuel.2012.05.058>

Baydar, N.G. and Akkurt, M. (2001). Oil content and oil quality properties of some grape seeds. *Turkish Journal of Agriculture and Forestry*, 25(3), 163–168. <https://doi.org/10.3906/tar-9909-25>

Choi, Y.S., Choi, J.H., Han, D.J., Kim, H.Y., Lee, M.A., Kim, H.W., Lee, J.W., Chung, H.J. and Kim, C.J. (2010). Optimization of replacing pork back fat with grape seed oil and rice bran fiber for reduced-fat

- meat emulsion systems. *Meat Science*, 84(1), 212–218. <https://doi.org/10.1016/j.meatsci.2009.08.048>
- Clemente, T.E. and Cahoon, E.B. (2009). Soybean oil: Genetic approaches for modification of functionality and total content. *Plant Physiology*, 151(3), 1030–1040. <https://doi.org/10.1104/pp.109.146282>
- Coitinho, T.B., Cassoli, L.D., Cerqueira, P.H.R., da Silva, H.K., Coitinho, J.B. and Machado, P.F. (2017). Adulteration identification in raw milk using Fourier transform infrared spectroscopy. *Journal of Food Science and Technology*, 54(8), 2394–2402. <https://doi.org/10.1007/s13197-017-2680-y>
- Davies, A.N., McIntyre, P. and Morgan, E. (2000). Study of the use of molecular spectroscopy for the authentication of extra virgin olive oils. Part I: Fourier transform Raman spectroscopy. *Applied Spectroscopy*, 54(12), 1864–1867. <https://doi.org/10.1366/0003702001948998>
- Dulf, F.V., Bele, C., An, M.U.Ş. and Socaciu, C. (2009). Phytosterols as Markers in Identification of the Adulterated Pumpkin Seed Oil with Sunflower Oil. *Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca - Agriculture*, 66 (2), 301–307. <https://doi.org/10.15835/buasvmcn-agr:3746>
- Flores, G., Ruiz Del Castillo, M.L., Herraiz, M. and Blanch, G.P. (2006). Study of the adulteration of olive oil with hazelnut oil by on-line coupled high performance liquid chromatographic and gas chromatographic analysis of filbertone. *Food Chemistry*, 97(4), 742–749. <https://doi.org/10.1016/j.foodchem.2005.06.008>
- González, A., Armenta, S. and de la Guardia, M. (2010). Adulteration detection of argan oil by inductively coupled plasma optical emission spectrometry. *Food Chemistry*, 121(3), 878–886. <https://doi.org/10.1016/j.foodchem.2009.11.091>
- Harris, H.C., McWilliam, J.R. and Mason, W.K. (1978). Influence of temperature on oil content and composition of sunflower seed. *Australian Journal of Agricultural Research*, 29(6), 1203–1212. <https://doi.org/10.1071/AR9781203>
- Irnawati, Riyanto, S., Martono, S. and Rohman, A. (2020). Determination of sesame oil, rice bran oil and pumpkin seed oil in ternary mixtures using FTIR spectroscopy and multivariate calibrations. *Food Research*, 4(1), 135–142. [https://doi.org/10.26656/fr.2017.4\(1\).260](https://doi.org/10.26656/fr.2017.4(1).260)
- Khudzaifi, M., Retno, S. and Rohman, A. (2020). The employment of FTIR spectroscopy and chemometrics for authentication of essential oil of *Curcuma mangga* from candlenut oil. *Food Research*, 4(2), 515–521. [https://doi.org/10.26656/fr.2017.4\(2\).313](https://doi.org/10.26656/fr.2017.4(2).313)
- López-Díez, E.C., Bianchi, G. and Goodacre, R. (2003). Rapid quantitative assessment of the adulteration of virgin olive oils with hazelnut oils using Raman spectroscopy and chemometrics. *Journal of Agricultural and Food Chemistry*, 51(21), 6145–6150. <https://doi.org/10.1021/jf034493d>
- Marina, A.M., Che Man, Y.B., Nazimah, S.A.H. and Amin, I. (2009). Monitoring the adulteration of virgin coconut oil by selected vegetable oils using differential scanning calorimetry. *Journal of Food Lipids*, 16(1), 50–61. <https://doi.org/10.1111/j.1745-4522.2009.01131.x>
- Matthäus, B. (2008). Virgin grape seed oil: Is it really a nutritional highlight? *European Journal of Lipid Science and Technology*, 110(7), 645–650. <https://doi.org/10.1002/ejlt.200700276>
- Mildner-Szkudlarz, S. and Jeleń, H.H. (2010). Detection of olive oil adulteration with rapeseed and sunflower oils using mos electronic nose and smpe-ms. *Journal of Food Quality*, 33(1), 21–41. <https://doi.org/10.1111/j.1745-4557.2009.00286.x>
- Mironeasa, S., Leahu, A. and Codin, G. (2010). Grape Seed: physico-chemical, structural characteristics and oil content. *Journal of Agroalimentary Processes and Technologies*, 16(1), 1–6.
- Riyanta, A.B., Riyanto, S., Lukitaningsih, E. and Rohman, A. (2020a). Analysis of sunflower oil in ternary mixture with grapeseed oil and candlenut oil. *Food Research*, 4(5), 1726–1731.
- Riyanta, A.B., Riyanto, S., Lukitaningsih, E. and Rohman, A. (2020b). The employment of fourier transform infrared spectroscopy (FTIR) and chemometrics for analysis of candlenut oil in binary mixture with grape seed oil. *Food Research*, 4(1), 184–190. [https://doi.org/10.26656/fr.2017.4\(1\).279](https://doi.org/10.26656/fr.2017.4(1).279)
- Roche, J., Bouniols, A., Mouloungui, Z., Barranco, T. and Cerny, M. (2006). Management of environmental crop conditions to produce useful sunflower oil components. *European Journal of Lipid Science and Technology*, 108(4), 287–297. <https://doi.org/10.1002/ejlt.200500310>
- Rohman, A. and Man, Y.B.C. (2010). Fourier transform infrared (FTIR) spectroscopy for analysis of extra virgin olive oil adulterated with palm oil. *Food Research International*, 43(3), 886–892. <https://doi.org/10.1016/j.foodres.2009.12.006>
- Siddique, B.M., Ahmad, A., Alkarkhi, A.F.M., Ibrahim, M.H. and Omar A.K.M. (2011). Chemical Composition and Antioxidant Properties of Candlenut Oil Extracted by Supercritical CO₂.

Journal of Food Science, 76(4), 535–542. <https://doi.org/10.1111/j.1750-3841.2011.02146.x>

Siddique, B.M., Muhamad, I.I., Ahmad, A., Ayob, A., Ibrahim, M.H. and Mohd Omar, A.K. (2015). Effect of frying on the rheological and chemical properties of palm oil and its blends. *Journal of Food Science and Technology*, 52(3), 1444–1452. <https://doi.org/10.1007/s13197-013-1124-6>

Vigli, G., Philippidis, A., Spyros, A. and Dais, P. (2003). Classification of edible oils by employing ³¹P and ¹H NMR spectroscopy in combination with multivariate statistical analysis. A proposal for the detection of seed oil adulteration in virgin olive oils. *Journal of Agricultural and Food Chemistry*, 51(19), 5715–5722. <https://doi.org/10.1021/jf030100z>

Wang, H.J. and Murphy, P.A. (1994). Isoflavone Content in Commercial Soybean Foods. *Journal of Agricultural and Food Chemistry*, 42(8), 1666–1673. <https://doi.org/10.1021/jf00044a016>