

Formulation and evaluation of herbal antioxidant cream from selected plants collected from the Nepalese Himalayan region

^{1,3}Paudel, P.N., ^{1,2}Acharya, A., ^{1,2}Paudel, A., ^{1,2}Shrestha, A., ⁴Satyral, P., ⁵Setzer, W.N. and ^{1,2,*}Gyawali, R.

¹Medicinal Plant Research Laboratory, Kathmandu University, Dhulikhel, Kavre, Nepal

²Department of Pharmacy, School of Science, Kathmandu University, Dhulikhel, Kavre, Nepal

³Department of Chemical Science and Engineering, School of Engineering, Kathmandu University, Dhulikhel, Kavre, Nepal

⁴Aromatic Plant Research Center, 230 N 1200 E, Suite 100, Lehi, UT 84043, USA

⁵Department of Chemistry, The University of Alabama in Huntsville, Huntsville, AL 35899, USA

Article history:

Received: 1 March 2024

Received in revised form: 17 June 2024

Accepted: 21 January 2025

Available Online: 5 May 2025

Keywords:

Herbals,
Crude extracts,
Cream,
Antioxidants,
Tyrosinase inhibition

DOI:

[https://doi.org/10.26656/fr.2017.9\(3\).054](https://doi.org/10.26656/fr.2017.9(3).054)

Abstract

Due to the strong tyrosinase inhibition and antioxidant effects, green tea and licorice are valuable herbal sources in the cosmetic industry for their beneficial effects on the skin. However, data on the addition of essential oils, green tea and licorice, to cream formulations to examine antioxidant activities is limited. Herbal antioxidants fight against radicals, UV rays, and unstable oxygen molecules that affect skin cells and produce wrinkles. The purpose of this study was to develop and assess herbal creams' antioxidant and tyrosinase-inhibitory characteristics using crude aqueous extracts of green tea and licorice loaded with essential oils. To load the best concentration on cream formulations, plant aqueous extracts were designed, evaluated, and correlated in terms of total phenolic content (TPC), total flavonoid content (TFC), and 2,2-diphenyl-1-picrylhydrazyl (DPPH) scavenging activity. Moreover, *Ocimum tenuiflorum* and *O. basilicum* essential oils were extracted and added to a cream formulation. The spreadability profile, water washability, centrifugation test, and organoleptic characteristics of the formulated oil-in-water cream were all satisfactory. The cream exhibited a non-Newtonian rheological profile and a pH range of 6.353 ± 0.065 to 6.467 ± 0.050 over successive 0, 1, 2, and 3 months at normal room temperature. The 50% inhibition concentrations shown by the herbal cream were 13.764 ± 0.153 $\mu\text{g/mL}$, 301.445 ± 1.709 $\mu\text{g/mL}$ and 8.082 ± 0.055 $\mu\text{g/mL}$ respectively, for 2,2-diphenyl-1-picrylhydrazyl (DPPH) scavenging activity, ferric (Fe^{3+}) reducing antioxidant power (FRAP) and 2,2'-azinobis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS) radical scavenging activity, and that of standard ascorbic acid were 6.716 ± 0.077 $\mu\text{g/mL}$, 171.604 ± 1.551 $\mu\text{g/mL}$ and 5.645 ± 0.034 $\mu\text{g/mL}$, which showed the formulated cream had strong antioxidant characteristics. The formulated herbal cream with a 50% tyrosinase inhibition concentration of 22.254 ± 0.369 $\mu\text{g/mL}$ compared to standard Kojic acid 12.535 ± 0.098 $\mu\text{g/mL}$ demonstrated a satisfactory tyrosinase inhibition profile for skin whitening properties. Herbal cream was reportedly stable in physical and chemical parameters for successive 0, 1, 2, and 3 months at both real and accelerated time study zones, according to the results of the stability study. The results provided evidence that formulated creams using plant-based natural ingredients are antioxidant-enriched products for skin protection.

1. Introduction

Skin, a large complex barrier organ of the body, structurally and functionally, is important for body immunologic and homeostasis regulation (Khavkin and Ellis, 2011). Being the most voluminous organ of the

body, skin's multilayer composition is responsible for several major bodily functions like temperature regulation, nervous system polarization, water system regulation, and being involved in body defensive roles, for instance, mechanical concussions and body protection against foreign chemicals, radiations, and

*Corresponding author.

Email: gyawali@ku.edu.np

microorganisms (Dąbrowska *et al.*, 2018; Zhang and Duan, 2018). Skin permeability, a key aspect of maintaining good skin hygiene, varies depending on individual age, ethnicity, food, gender, and body mass index (BMI), which mostly influence cosmetic beauty (Khavkin and Ellis, 2011; Dąbrowska *et al.*, 2018). However, physiological and environmental factors like inherited skin conditions, metabolic and hormonal disorders (MHD), prolonged sun exposure, oxidative stress, indoor pollution, an unbalanced lifestyle, and synthetic cosmeceutical heavy metals worsen skin hygiene, hastening the onset of premature skin aging (Bonte *et al.*, 2019). Reactive oxygen species (ROS) in the body interfere with the skin turnover cycle, causing dry, sagging skin, degradation of cutaneous and epidermal processes, a decline in skin elasticity and defensive function, and dermal atrophy, which is a sign of aging (Yeh *et al.*, 2021). Most commonly, ultraviolet (UV) radiation accumulates free radicals on skin cells, accelerating both chronological and photo-aging phenomena and producing matrix metalloproteinases (MMPs), which inhibit hyaluronic acid (HA) synthesis, depigment skin, disarrange skin collagen and elastin, and cause the dermal extracellular matrix to degenerate over time (Cao *et al.*, 2020; Yeh *et al.*, 2021). Aging mostly results from bodily oxidative stress exceeding antioxidant properties, so in order to prevent early aging, skin care routines today include cosmetic antioxidant supplements, either herbal or synthetic. Antioxidants strengthen the skin's natural defense system, counteracting oxidative free radicals and reducing skin aging. In addition to antioxidant theory, important ongoing study areas for anti-aging techniques include microRNA, cellular senescence, and advanced glycation end products (AGEs) (Eckhart *et al.*, 2019; Cao *et al.*, 2020). Daily modern cosmetics like gentle surfactant cleansers or anti-pore clogging lotions, skin toners, moisturizer, and cosmeceuticals unblock have significantly slowed down the signs of aging; however, synthetic cosmetics containing parabens preservatives, formaldehyde, diethylhexylphthalates, phenylenediamine, mercury-like heavy metals, sodium lauryl sulphate (SLS) surfactant, and methylene glycol are equally harmful to skin as they cause skin irritation, dryness, hormonal imbalances, and are carcinogenic (Bonte *et al.*, 2019; Draeos, 2019). Due to negative effects like redness, stinging, and itching, hydroquinone, the gold standard for skin whitening, is a contentious cosmetic chemical in the Food and Drug Administration (FDA) realm (Draeos, 2019). Additionally, loading broad-spectrum UV filter nano-metal oxides into cosmetics and personal care items like sunscreen, soap, and shampoo demonstrates both dose-dependent toxicity and eco-unfriendliness (Subramaniam *et al.*, 2019).

Synthetic antioxidants like retinoid derivatives, α -hydroxy acids (AHA), β -hydroxyl-butanoic acid (BHB), idebenone, or polyfluoroalkyl substances (PFAS) are cytotoxic, carcinogenic, and skin irritants, inducing premature senescence. Also, long-term usage of cosmeceutical fragrances has been linked to allergic contact dermatitis (ACD), but combining them with biological sources like crude plant extracts, essential oils, and probiotic cosmetics has demonstrated synergistic benefit, limiting any potential possible skin side effects (Alani *et al.*, 2013; Carocho and Ferreira, 2013; Bonte *et al.*, 2019; Subramaniam *et al.*, 2019). For example, adding complex natural plant extracts or essential oils to synthetic cosmetics can stimulate beneficial skin effects (Jadoon *et al.*, 2015).

For healthy skin, phyto-oxidants like terpenes (carotenoids and essential oils) and polyphenols (flavonoids and stilbenes) are excellent green cosmeceutical substitutes and are considered nowadays as innovative beauty products (Jadoon *et al.*, 2015; Bonte *et al.*, 2019). Botanical creams of arbutin, aleosin, kojic acids, and licorice extract (liquirtin, glabridin, glabrene, and isoliquirtin) are effective natural tyrosinase inhibitors for skin lightening with no adverse effects (Jadoon *et al.*, 2015; Draeos, 2019; Ahuja *et al.*, 2021). Around 400 isolated flavonoids from licorice, such as glycyrrhizin, glycyrrhetic acid, glabridin, quercetin, formononetin, hispaglabridin B, and methylglabridin, show potential antioxidant, anticancer, antimicrobial, and anti-inflammatory effects; however, licorice polyphenolic extracts, such as liquiritigenin, isoliquiritigenin, chalcones, and licochalcone C, are also strong skin-lightening agents (Ahuja *et al.*, 2021; Wahab *et al.*, 2021). Green tea (*Camellia sinensis*) aqueous or ethanolic extraction contains highly enriched green tea polyphenols (GTP) such as gallic acid (GA), galocatechin (GC), catechin (C), epicatechingallate (ECG), and epigallocatechingallate (EGCG) that improve cell turnover by activating phase II antioxidant enzymes in the body and keep skin hydrated (Forester and Lambert, 2011; Jadoon *et al.*, 2015). When green tea (*Camellia sinensis*) extract is combined with other extracts of naturally occurring antioxidant plants like *Ruscus aculeatus*, *Nelumbo nucifera*, *Calendula officinalis*, *Embllica officinalis*, *Capparis decidua*, *Castanea sativa*, *Coffea arabica*, *Malus domestica*, *Moringa oleifera*, *Morus alba*, and *Polygonum minus*, it demonstrates incredible skin anti-aging potential due to their multi-phytochemical synergistic interactions, which are considered more beneficial for skin than loading specific synthetic antioxidants into cosmetic creams (Jadoon *et al.*, 2015; Ahuja *et al.*, 2021). Topical delivery of natural phenolic acids, flavonoids, or essential oils encapsulated on synthetic lipid cream

vesicles may potentially rejuvenate aging skin and guarantee effective green antioxidant skin care formulations with safer, eco-friendly ingredients (Ahuja et al., 2021; Lohani et al., 2021). Research has been done on probiotics found in cosmetic products like *Lactobacillus acidophilus* and *Vitreoscilla filiformis* that naturally rebalance skin microbiota, restoring skin signal components, making them more beneficial to skin hygiene in addition to phyto-antioxidant therapies for treating skin aging (Bonte et al., 2019; Draelos, 2019; Puebla-Barragan and Reid, 2021). Cosmetic interventions on marine flora like *Sargassum myriocystum* and phytomoisturizers, supplemented with neurotransmitter peptides, are two examples of new replacement tactics in synthetic cosmetic fields that are emerging, but research study fields are still limited to this date (Bonte et al., 2019; Draelos, 2019; Subramaniam et al., 2019; Siahaan et al., 2022). Direct use of crude plant extracts is now a beneficial, quick approach for the commercialization of cosmetic care that provides the finest potential skincare results. Without a doubt, phytocosmetic creams are today's greatest cosmetic skin care products over synthetic ones because they are eco-friendly and give positive skin benefits with little to no side effects. Thus, this study aimed to formulate a novel phytocosmetic cream with essential oils and crude aqueous plant extracts from *Camellia sinensis* leaves and *Glycyrrhiza glabra* roots that show good tyrosinase-inhibitory and antioxidant properties. This study also aimed to provide new research and development (R and D) and secure safer alternatives to cosmeceutical skin-care formulations.

2. Materials and methods

2.1 Plant material collection

Ocimum tenuiflorum and *O. basilicum* fresh plant samples were collected from Kapilvastu with latitude 27°40'7.9''N and longitude 83°10'12.9''E, and Bardiya with latitude 28°15'14.3''N and longitude 81°30'7.0''E, Nepal, in March/April 2022, respectively (as shown in Figure 1), then authorized by the National Herbarium

and Plant Laboratories, Lalitpur, Nepal. Freshly verified *Glycyrrhiza glabra* roots and *Camellia sinensis* leaves were purchased from Herbs Production and Processing Co. Ltd. (HPPCL), Kathmandu, Nepal.

2.2 Extraction of essential oils

Each plant (100 g) was subjected to hydro distillation for 3 hrs in 500 mL of distilled water on a Clevenger apparatus (Jain Scientific Glass Work, JSGW, India) after shade-drying and chopping the aerial parts of *O. tenuiflorum* and *O. basilicum*. The extracted essential oils were dried with activated sodium sulphate, filtered, and stored at 4°C until further analysis and cream formulation.

2.3 Plant extraction

Each plant of 5 g sun-dried powder of *Glycyrrhiza glabra* roots and *Camellia sinensis* leaves was extracted separately in 50 mL of distilled water and heated in an enclosed 100 mL beaker at 70-75°C in a heating mantle (Biobase 1000 mL capacity, Germany) for 45 mins. Such crude aqueous extracts were collected, stored, and labelled in a clean 100 mL centrifuge tube separately, after being filtered by filter paper (Whatman GE Healthcare, Cat No. 1001 125, UK) (Whangsomnuek et al., 2019; Kurzawa et al., 2022).

Table 1. Optimal concentration design for each aqueous plant extracts.

Design	<i>Camellia sinensis</i> extraction (V1)	<i>Glycyrrhiza glabra</i> extraction (V2)
D1	-1	-1
D2	-1	0
D3	-1	+1
D4	0	-1
D5	0	0
D6	0	+1
D7	+1	-1
D8	+1	0
D9	+1	+1

2.4 Design for aqueous extracts

The optimal aqueous extract load on cream formulation was designed using the Statgraphics 19 Centurion programme with three level designs, -1, 0, and +1, which corresponded to 0.5 g, 1.5 g, and 3 g of each aqueous extract, as shown in Table 1. For instance, D3 signified 0.5 g of V1 and 3 g of V2 to load on the cream formulation.

2.5 Total phenolic content and total flavonoid content of aqueous extract determination

With a small modification to the previously

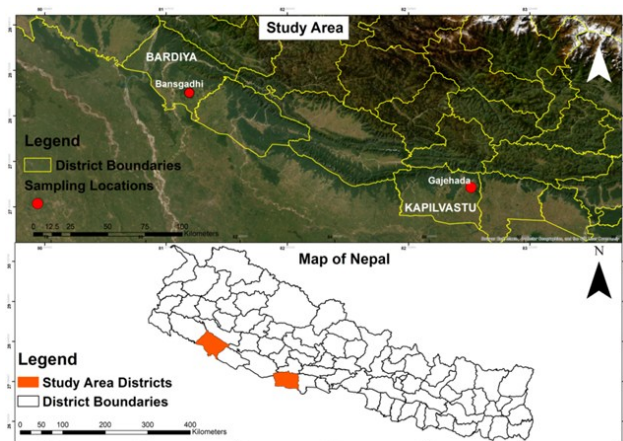


Figure 1. Geographical location of the sample collection sites.

described methods, Srisuksomwong *et al.* (2023) and Mungmai *et al.* (2019) were used to calculate the total phenolic content (TPC). Each 250 μL of design extract was mixed with 1250 μL of deionized water and 250 μL of ethanol (95%). Then, each mixture was subjected to 125 μL of 50% phenol reagent (Fisher Scientific, Prod. No. 35953, India), shaken for 3 mins, and 250 μL of 5% Na_2CO_3 added at room temperature. The absorbance of each mixture was measured at 725 nm in a UV spectrophotometer (UV-1800 Shimadzu, Japan) after only 1 hr of sample incubation at a dark room temperature. Standard gallic acid (Sigma-Aldrich, Phytolab GmbH and Co.KG, Germany) was produced in methanol at varied concentrations of 6.25 $\mu\text{g}/\text{mL}$, 12.5 $\mu\text{g}/\text{mL}$, 25 $\mu\text{g}/\text{mL}$, 50 $\mu\text{g}/\text{mL}$, and 100 $\mu\text{g}/\text{mL}$, and the absorbance was measured using the same technique as for samples for developing a calibration curve and calculating TPC for samples represented as gallic acid equivalent GAE (mg/g) (Genwali *et al.*, 2013; Agbor *et al.*, 2014). Triplicate data were taken.

Total Flavonoid Content (TFC) for an aqueous extract was determined using the aluminium chloride colorimetric method with a small modification to the standard procedure previously described by Madaan *et al.* (2011) and Phosri *et al.* (2022). A mixture of 50 μL of 10% AlCl_3 and 750 μL of methanol was added to each 250 μL aqueous extract. The mixture was shaken and incubated for 30 mins at dark room temperature after the addition of 50 μL potassium acetate (1M) and 1400 μL deionized water. Absorbance was measured in triplicate for each design at 415 nm via a UV spectrophotometer (UV-1800 Shimadzu, Japan) (Srisuksomwong *et al.*, 2023). Standard quercetin (Sigma-Aldrich, USA) was produced in absolute ethanol at varied concentrations of 6.25 $\mu\text{g}/\text{mL}$ to 100 $\mu\text{g}/\text{mL}$ by serial dilution, and the absorbance was measured three times using the same technique as for samples for developing a calibration curve and calculating TFC for samples represented as quercetin equivalent QE (mg/g).

2.6 Screening for DPPH activity in aqueous extract designs and essential oils

The DPPH colorimetric method, which was slightly modified from the previously described standard method (Paudel *et al.*, 2022), was used to determine antioxidant properties. Through serial dilutions, different aqueous extracts and essential oils were developed in ethanol with concentrations ranging from 15.625 $\mu\text{g}/\text{mL}$ to 1000 $\mu\text{g}/\text{mL}$. Approximately 3.9 mg DPPH (Sigma-Aldrich, MO, USA) was diluted in 100 mL of ethanol for 0.1 mM DPPH stock solution preparation. Each 2 mL of different sample concentrations was mixed with 2 mL of DPPH (0.1 mM) solution, and the mixtures were allowed to stand in a dark room at room temperature for 30 mins

before being measured in triplicate at 517 nm on a UV spectrophotometer (UV-1800 Shimadzu, Japan). The absorbance of control solution (2 mL DPPH stock solution and 2 mL ethanol) and blank solution (2 mL ethanol) was also measured at 517 nm, and the IC_{50} value was calculated using the linearity of the DPPH scavenging effect (%) given by the authors (Hasan *et al.*, 2009; Kurzawa *et al.*, 2022).

$$\text{DPPH scavenging effect (\%)} = \left[\frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of control}} \right] \times 100$$

2.7 Pre-formulation physical compatibility test

Each excipient from the cream formulation was mixed in a 1:1 ratio with plant extracts and synthetic actives for 10 mins before being placed in separate vials at room temperature with direct sunlight exposure for 1 month to maximize damage effects. The physical changes observed included texture, homogeneity, odor, and any color development (Liao *et al.*, 2022).

2.8 Antioxidant-enriched phytocosmetic cream formulation

The oil phases were heated at 70-80°C and the water phases were heated at 50-55°C with continuous stirring. The heated oil phase was added slowly in the water phase at a maintained temperature of 50°C with continuous stirring at 2000 rpm for 25 mins, and the cold phases were added with increasing speed to 2500 rpm for 10 mins. During the mixing process of the cold phase, the optimal aqueous extract and essential oil were loaded, and the cream was cooled to room temperature (Tan *et al.*, 2022). *Ocimum basilicum* essential oil was added via syringe drop as a fragrance. Carbopol-940 was soaked one night before addition to the formulation. The different compositions of formulated cream are presented in Table 2.

The design of aqueous plant extracts was loaded onto cream by best demonstrating TFC, TPC, and DPPH activity. In addition, *O. tenuiflorum* essential oil was loaded with an IC_{50} value calculated from DPPH. As a fragrance, a syringe drop of *O. basilicum* essential oil was added.

2.9 Organoleptic characteristics

Physical texture, homogeneity, and smoothness were evaluated by pressing a 1 g cream between the thumb and index finger. Odor and color were also accessed (Tan *et al.*, 2022). Cream was inverted at 180° to observe any flow properties.

Table 2. Herbal cream composition.

Ingredient	Quantity for 100 g (%)
Water phase	
Glycerin	9.0
Sodium benzoate	0.3
Hyaluronic acid	0.2
Allantoin	0.5
Distilled water	q.s
Oil phase	
Cetyl alcohol	3.0
Arlacel	2.0
Liquid paraffin	5.0
Beeswax	1.0
Glycerylmonostearate	2.0
Cold phase	
Dimethicone	3.0
Triethanol amine	0.3
Cyclopentasiloxane	5.0
Potassium sorbate	0.3
Retinol	0.2
Niacinamide	0.5
Carbopol-940	0.2
Magnesium aluminum silicate	0.8
Guar hydroxyl propyltrimonium chloride	0.2
Zinc oxide	0.1
<i>Ocimum basilicum</i>	0.1

2.10 Cream type determination and water washability test

A dilution test was carried out with a 1:10 ratio of cream to distilled water for the miscibility test with water. Oil-in-water (o/w) type cream shows good miscibility with water and vice versa (Ahmed and Nath, 2017). Also, the microscopic view (40×) for base cream and herbal loads was observed separately on the microscope (OptiCx, LABOMED, India) (Glusac et al., 2018).

On the upper hand's skin section, 0.2 g of cream was applied, and the area was gently washed under running water. A good washability cream can be easily removed from the application site and leaves the skin's surface without being greasy (Yadav et al., 2014; Tan et al., 2022).

2.11 Cream spreadability, pH determination, and centrifugation test

A glass slide, into which 200 g of weight was placed, was used to press 0.5 g of cream placed in center of another glass slide for 2 mins at room temperature. The diameter of the cream was measured in triplicate for three months. Spreadability of 5-7 cm demonstrates that cream can be easily distributed with little effort on the site of application and shows uniform particle distribution (Yadav et al., 2014; Tan et al., 2022).

The pH of a 10% w/v cream suspension in distilled water was measured three times at 25°C for three months using a pH meter (HI 2210 pH meter, Italy), which was previously calibrated using standard buffers with pH 4, 7, and 10, respectively. An effective cream has a skin physiological pH range of 6-8 (Pinto et al., 2021; Tan et al., 2022).

About 2 g of cream was centrifuged at 5000 rpm for 10 mins at 25°C and emulsion instability (%) was calculated using the following equation (Restu et al., 2015) for three months;

$$\text{Emulsion instability (\%)} = \left[\frac{\text{Emulsion separation (cm)}}{\text{total emulsion height (cm)}} \right] \times 100\%$$

An ideal unstable cream emulsion has 100% emulsion instability (Restu et al., 2015; Tan et al., 2022). This experiment was repeated three times.

2.12 Rheology profile study

The formulated cream was directly immersed into the Brookfield viscometer (DV III Ultra, Italy) with spindle no. 64, initially with an adjusted rpm of 1 to 10 at room temperature and later with a shear rate of 17-102 s⁻¹ (Gyawali et al., 2020), where the shear rate was 1.7 times the adjusted spindle rpm. The final results were represented as a graph of viscosity (Pascal-second) vs. shear rate (s⁻¹). The thixotropic profile was also studied as a function of shear stress (Pascal) vs. shear rate (s⁻¹), which demonstrates that a good thixotropic formulation quickly restores to its initial position and is therefore thought to be stable (Almeida et al., 2015; Gyawali et al., 2020; Pinto et al., 2021). The measured dial readings were multiplied by a factor of 10 to give viscosities in centipoise (cP) or milipascal-second (mPa.s), where 1cP = 1mPa.s.

2.13 Antioxidant assays

The 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging method, ferric (Fe³⁺) reducing antioxidant power (FRAP), and 2,2'-azinobis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS) radical scavenging assay were performed to determine cream antioxidant activity.

The cream was produced on ethanol from 1.25 µg/mL to 40 µg/mL on serial dilution after filtration using the DPPH method as mentioned above for aqueous extract designs. A standard ascorbic acid (Fischer Scientific, India) sample was also prepared in ethanol to various concentrations ranging from 1.25 µg/mL to 40 µg/mL by serial dilutions, and absorbance was measured as described above. This experiment was carried out three times (Paudel et al., 2022; Phosri et al., 2022).

A slight modification of the previously established

protocol was used for the FRAP assay (Park *et al.*, 2008). All chemicals for this assay were purchased from Fischer Scientific, India. By serial dilutions, standard ascorbic acid and cream formulations were developed in ethanol to various concentrations ranging from 31.25 µg/mL to 500 µg/mL. Freshly prepared solutions of 2.5 mL phosphate buffer (0.2 M, pH 6.6) and 2.5 mL potassium ferricyanide (1%) were added to each 1 mL sample preparation and incubated for 20 mins at 50°C before adding 2.5 mL trichloroacetic acid (10%) and centrifuging for 10 mins at 1000 rpm. 2.5 mL of each collected supernatant was diluted to 5 mL with distilled water before adding 0.5 mL of FeCl₃ (0.1%) by vigorous shaking at room temperature. After 30 mins, absorbance at 700 nm was measured three times on a 96-microplate reader (BioTek, EPOCH).

A graph of mean absorbance vs. sample concentration was plotted to compare effective concentrations (EC₅₀-value, µg/mL) with standard ascorbic acid, where the EC₅₀ value represents the effective reducing concentration of the sample with a mean absorbance value of 0.5 (Paudel *et al.*, 2022; Phosri *et al.*, 2022). The lower the EC₅₀ value, the greater the antioxidant capacity of the sample to reduce ferric (Fe³⁺) ions to ferrous (Fe²⁺) ions.

For the ABTS radical scavenging assay, a slightly modified version of the previously described method was used (Re *et al.*, 1999). Standard ascorbic acid (Fischer Scientific, India) and cream formulations were developed in methanol using serial dilutions to concentrations ranging from 1.25 µg/mL to 40 µg/mL. A 7 mM ABTS (38.402 mg of ABTS in 10 mL distilled water) (Glenthams Life Sciences, UK) was mixed with potassium persulfate (6.600 mg) to give the final ABTS stock (10 mL, 2.45 mM) and allowed to stand at room temperature for 14-16 hrs to form a stable oxidized ABTS radical. This working ABTS stock solution was diluted with methanol (1:90 v/v) until the absorbance at 734 nm was 0.70±0.02. Now 140 µL of ABTS stock solutions were added to each 70 µL of sample preparation in a 96-well plate, incubated for 10 mins at dark room temperature, and triplicate absorbance was measured at 734 nm using a 96-microplate reader (Bio Tek, EPOCH). Using the given equation (Shah and Modi, 2015), the linear percentage inhibition concentration (IC%) was calculated, and an IC₅₀% value was compared with the standard.

$$IC \% = [(mean\ absorbance\ of\ control - mean\ absorbance\ of\ sample) / mean\ absorbance\ of\ control] \times 100$$

As a control sample, 70 µL of methanol was mixed with 140 µL of ABTS stock solutions.

2.14 Mushroom tyrosinase inhibition study

The inhibition activity of the tyrosinase in cream formulation was studied using the standard procedure described by Pandey *et al.* with minor modifications (Pandey *et al.*, 2022). All chemicals were obtained from Sigma Chemical Co., USA. The substrate for this reaction was 1-4-dihydroxyphenylalanine (L-DOPA). The positive control was Kojic acid. Cream and Kojic acid were developed using serial dilutions of ethanol concentrations ranging from 1.25 µg/mL to 80 µg/mL. 685 µL of freshly prepared phosphate buffer (0.05 M, pH 6.5), 15 µL of mushroom tyrosinase (2500 units/mL), 200 µL of cream concentrations, and 100 µL of L-DOPA were allowed to react before immediately measuring the mixture reaction as dopachrome formation at 492 nm in a 96-Microplate reader (Bio Tek, EPOCH). This experiment was repeated three times. The effective concentration to inhibit 50% of mushroom tyrosinase (IC₅₀%) was calculated using a linearity graph of % inhibition vs. concentration, as given by equations (Di Petrillo *et al.*, 2016; Vichit and Saewan, 2022):

$$\text{Tyrosinase inhibition (\%)} = [(mean\ absorbance\ of\ control - mean\ absorbance\ of\ sample) / mean\ absorbance\ of\ control] \times 100$$

In place of cream concentrations, 200 µL of ethanol was added as a control sample, and the absorbance was measured at 492 nm using the same method. The best tyrosinase inhibition property favorable for whitening impact is shown by a lower value of IC₅₀ (%).

2.15 Stability study

The formulated cream was subjected to ICH climatic zone 30±2°C and relative humidity (RH) 75±5% for 3 months. Also, the freeze-thaw cycling test for accelerated stability was carried out for 48 hrs at 5-8°C and 48 hrs at 40-42°C for three successive months. The cream's spreadability, pH, centrifugation, and antioxidant activities were determined in triplicate as previously described methods (Pinto *et al.*, 2021; Plyduang *et al.*, 2022).

2.16 Statistical data analysis

All independent experiments were run three times (n = 3), and mean ± standard deviation (SD) was used to express the data. The figures for IC₅₀ (%) values were computed using Microsoft Excel OriginPro 2016 64Bit (Origin version 9.3, USA). The sampling site map was made using GIS software (ArcGIS software, USA). To design the optimal aqueous extract loading concentration for analysis, Statgraphics19 Centurion (Statgraphics Technologies, Inc., USA) software was used.

3. Results and discussion

3.1 Screening for total phenolic content, total flavonoid content, and DPPH activity in aqueous extract designs, and DPPH activity in essential oil

The TPC, TFC, and DPPH activities for various aqueous designs and essential oils are shown in Figure 2. The aqueous design D3 had the highest TPC value of 294.029 ± 0.227 mg GAE/g, whereas D5 had a lower value of 174.059 ± 0.325 mg GAE/g compared to standard gallic acid, which has a linear correlation coefficient $R^2 = 0.991$, $y = 0.011x + 0.059$. Also, D3 showed the highest TFC value of 88.504 ± 0.315 mg QE/g, while D4 showed a lower TFC value of 10.289 ± 0.253 mg QE/g compared to standard quercetin, which has a linear correlation coefficient $R^2 = 0.991$, $y = 0.013x + 0.173$. Since phenolics and flavonoids are potent antioxidants, the higher the TPC and TFC values in an aqueous design, the more skin-beneficial properties are present (Kurzawa et al., 2022). This data showed that there were more hydrophilic polyphenolic compounds like gallic acid (GA), gallic acid (GA), gallic acid (GA), catechin (C), epicatechin (EC), epigallocatechin (EGC), epicatechingallate (ECG), epigallocatechingallate (EGCG), and coumaroylquinic acid (CA) in crude aqueous extract than hydrophobic flavonoids (glycyrrhizin, glycyrrhetic acid, and glabridin), as comparatively all designs had more TPC value than TFC value, since most of Licorice flavonoids are hydrophobic (Forester and Lambert, 2011; Tungmunnithum et al., 2018; Wahab et al., 2021). Secondary plant metabolites, such as phenolic and flavonoid chemicals like EGCG, EC, GA, isoliquiritigenin, and chalcones, are primarily hydrophilic and have significant UV protective benefits for sun-damaged skin (Forester and Lambert, 2011; Tungmunnithum et al., 2018; Wahab et al., 2021).

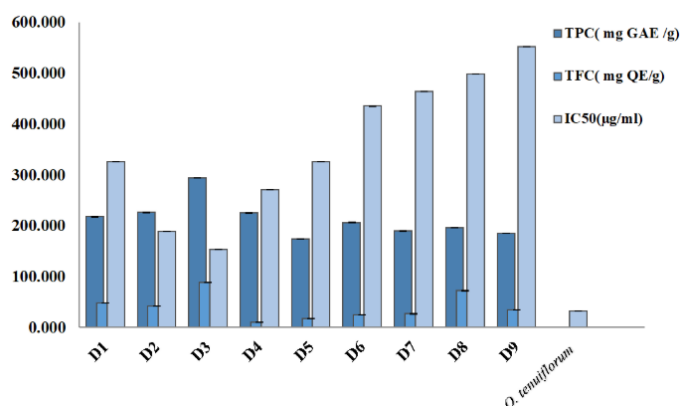


Figure 2. TPC, TFC and DPPH activities (IC_{50}) in aqueous extract designs and essential oil. Data presented are in mean \pm SD ($n = 3$).

The designs D2 and D3 demonstrated stronger DPPH antioxidant activities (low IC_{50} values) than other crude aqueous extracts since low IC_{50} values indicate that lower concentrations ($\mu\text{g/mL}$) of extracts are

required to scavenge free radicals of DPPH. The D3 demonstrated strong antioxidant activity with an IC_{50} value of 153.481 ± 0.002 $\mu\text{g/mL}$, and the D9 demonstrated weaker antioxidant activity with an IC_{50} value of 552.047 ± 0.002 $\mu\text{g/mL}$, however, *O. tenuiflorum* essential oil demonstrated stronger antioxidant activity than all other crude aqueous designs with an IC_{50} value of 32.073 ± 0.020 $\mu\text{g/mL}$ since *Ocimum* species essential oils show potential antioxidant properties (Awah and Verla, 2010; Chaudhary et al., 2020). Therefore, essential oil of *O. tenuiflorum* 33 $\mu\text{g/mL}$ was loaded by syringe drop into the cream with aqueous design D3, as it showed stronger antioxidant activity than other herbal extracts during the screening process. As an initial step in the skin-anti-aging strategy, it is believed that the synergistic addition of this chosen concentration of plant extracts, when incorporated with cream bases, can strongly reduce oxidative stress on the skin by demonstrating high antioxidant capacity (Awah and Verla, 2010).

However, the correlation of DPPH activity (IC_{50} , $\mu\text{g/mL}$) with TPC (mg GAE/g) and TFC (mg QE/g) for aqueous designs (D1-D9) is shown in Figure 3 (a, b). The total phenolic content in aqueous extracts like EGCG, EG, GA, and chalcones contributed weakly to antioxidant activity, as indicated by the 0.562 correlation coefficient (R^2) between DPPH activity (IC_{50} , $\mu\text{g/mL}$)

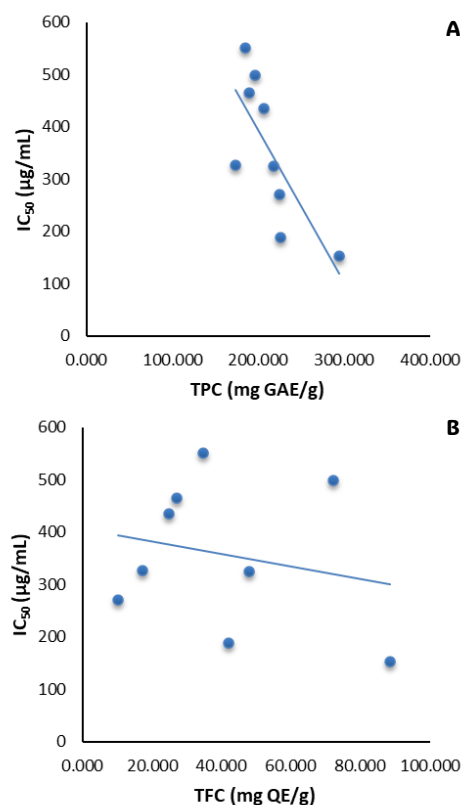


Figure 3. Correlation between DPPH activity (IC_{50} , $\mu\text{g/mL}$): (A) with TPC (mg GAE/g), $R^2=0.562$ demonstrated lesser correlation, (B) with TFC (mg QE/g), $R^2=0.048$ demonstrated no significant correlation. Data presented are in mean \pm SD ($n = 3$).

and TPC (mg GAE/g), which also indicated a lower value of IC₅₀ (µg/mL) on a greater TPC (mg GAE/g) value. Moreover, no significant correlation ($R^2 = 0.048$) was found between total hydrophilic flavonoids and antioxidant properties in this present study, yet phenolic and flavonoid contents are both crucial for skin-beneficial properties (Kumar *et al.*, 2014).

3.2 Physical compatibility test

The plant extracts and synthetic actives showed no characteristic change (NCC) in color, texture, homogeneity, or odor when mixed with cream-based excipients for a month in separate vials. Physical compatibility testing of excipients is required because it is important in product development and commercialization, allowing for the selection of the best excipients and maximizing shelf life (Tan *et al.*, 2022).

3.3 Organoleptic characteristics

The organoleptic characteristics of the cream were evaluated physically. The prepared cream had a uniform texture that was smooth and calming, with no grit or greasiness at all. It had a light yellowish-brown color and a fresh *O. basilicum* aroma because it was loaded with essential oils and aqueous plant extracts. At a 180° inversion, the cream did not flow, an optimal requirement for topical application (Salehi *et al.*, 2022).

3.4 Cream-type determination and water washability test

The cream showed good miscibility with water and easy water-washing properties. A good oil-in-water (o/w) type cream is less greasy than a water-in-oil (w/o) type cream and is mostly acceptable cosmetically (Mishra *et al.*, 2014). The formulated herbal cream enriched with added phyto-antioxidants is shown in Figure 4. The microscopic view of herbal cream is also shown in Figure 5.



Figure 4. Formulated herbal cream enriched with added phyto-antioxidants.

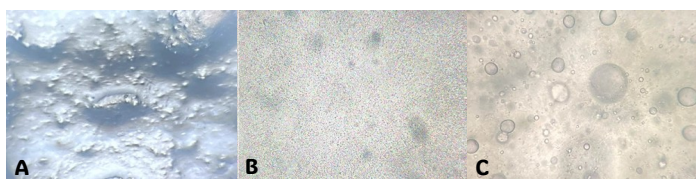


Figure 5. Microscopic profile (40×) representation, A: homogeneous distribution of cream base only, B: loaded cream base with emulsified aqueous extract design (D3), C: visible cream base with aqueous extract design (D3) and essential oil droplets seen (final herbal cream).

3.5 Cream spreadability, pH determination, and centrifugation test

The results for cream spreadability, pH determination, and centrifugation tests for both real-time and accelerated studies (freeze-thaw cycling) are presented in Table 3. The spreadabilities and cream pH over the course of all studies were within acceptable limits (Yadav *et al.*, 2014; Pinto *et al.*, 2021; Tan *et al.*, 2022). The emulsion instability (%) ranged from 8.222 ± 1.678 to 13.333 ± 0.667 , which were comparatively low and showed that the formulated cream was a good stable emulsion in both real and accelerated time studies (Restu *et al.*, 2015). While pH showed no significant change, freeze-thaw cycling had little effect on cream spreadability and centrifugation parameters. This is because cream viscosity changes under accelerated conditions (Tan *et al.*, 2022), whereas no significant pH change demonstrated that loaded active herbal extracts were in good condition for both long-term real and accelerated conditions. Creams with a pH close to that of the skin (pH values 5-7) irritate the application site less. Any topical cosmeceuticals having a good spreadability index (5-7 cm) show that they can be administered on the skin surface with little effort (Yadav *et al.*, 2014; Tan *et al.*, 2022).

3.6 Rheology profile study

A non-Newtonian rheology profile was shown by the herbal cream since, with increasing shear rate (s^{-1}), the viscosity (Pa.s) had decreased but not linearly. When the viscosity (Pa.s) of cream decreased as shear stress (Pa) was raised, this behavior was more accurately characterized as shear thinning (pseudoplastic) behavior. When this cream was applied to the skin surface, the viscosity would increase because the shear stress was initially low, but when gently rubbed, the viscosity would decrease because it behaved more like a shear thinning profile than a shear thickening profile, as shown in Figure 6. The initial apparent viscosity was highest at 101.667 ± 2.082 Pa.s at a shear rate of $1.7 s^{-1}$, as shear stress was low initially (56.000 ± 2.000 Pa), but apparent viscosity decreased with increasing shear rate (s^{-1}). Figure 6 illustrates the thixotropic profile of the cream. The pseudoplastic profile of cosmetic formulations is advantageous because it improves cream spreadability, which in turn improves skin permeability as rubbing time increases shear stress (Korhonen *et al.*, 2001). The stability of the product is also guaranteed by the rheological properties of the cream; for instance, a thermodynamically unstable cream exhibits phase separation as a result of low interfacial viscous forces between two different phases (oil/water) (Korhonen *et al.*, 2001; Glusac *et al.*, 2018).

Table 3. Cream spreadability (cm), pH and centrifugation test (%) data at room temperature and accelerated stability study (freeze-thaw cycling).

Duration (months)	Room temperature			Freeze-thaw cycling		
	Spreadability (cm)	pH	Centrifugation (%)	Spreadability (cm)	pH	Centrifugation (%)
0	6.133±0.058	6.467±0.050	8.222±1.678	5.633±0.153	6.357±0.055	10.000±2.000
1	6.000±0.173	6.420±0.035	10.667±1.155	5.533±0.115	6.260±0.017	11.556±0.385
2	5.833±0.208	6.420±0.092	8.889±0.770	5.467±0.115	6.327±0.025	13.333±0.667
3	5.567±0.208	6.353±0.065	10.000±1.333	5.433±0.153	6.230±0.062	12.444±0.770

Values are presented as mean±SD of independent triplicate (n = 3).

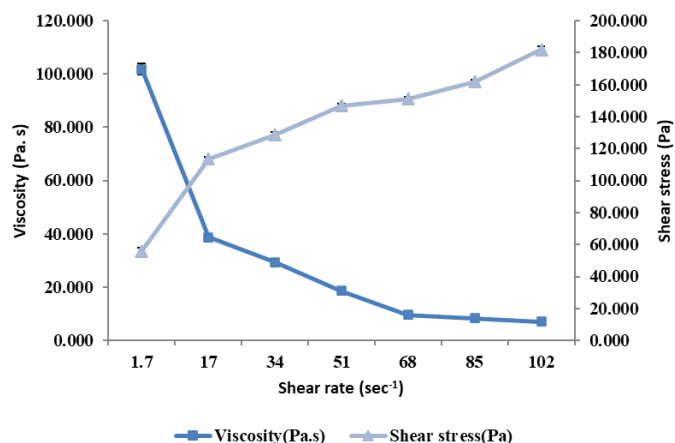


Figure 6. Rheology profile for formulated herbal cream represented graphically. At each shear rate (s^{-1}), the viscosity (Pa.s) and shear stress (Pa) were varied (pseudo plastic behavior). Data presented are in mean±SD (n = 3).

3.7 Antioxidant assays and mushroom tyrosinase inhibition study

The antioxidant and tryosinase inhibition activities of the formulated phytocosmetic cream are given in Figure 7. Antioxidant supplements, an anti-aging marker, can help to improve skin aging caused by ROS stress (Jadoon *et al.*, 2015; Bonte *et al.*, 2019). A complex group of botanical extract-loaded creams have skin-protective properties; for instance, they act against skin wrinkle formation and lesions due to their antioxidant phytochemicals (Jadoon *et al.*, 2015). Skin aging causes hyperpigmentation of the skin due to excessive Tryosinase, a key enzyme for melanin production that, when inhibited, reduces melanogenesis, is a key strategy for skin depigmentation and the leading anti-aging strategy. For example, synthetic tryosinase inhibitors such as hydroquinone and kojic acid are cosmetic anti-pigmenting agents (Di Petrillo *et al.*, 2016; Vichit and Saewan, 2022). In this study, the DPPH activity IC_{50} value shown by the herbal cream was $13.764 \pm 0.153 \mu\text{g/mL}$, whereas the standard ascorbic acid showed $6.716 \pm 0.077 \mu\text{g/mL}$, indicating that this cream had good antioxidant activity for skin beneficial properties. Also, the EC_{50} values for cream and ascorbic acid in the FRAP assay were $301.445 \pm 1.709 \mu\text{g/mL}$ and $171.604 \pm 1.551 \mu\text{g/mL}$, respectively. This FRAP assay also indicated that herbal cream had higher antioxidant properties than

standard ascorbic acid. Furthermore, the ABTS scavenging activity of herbal cream was (IC_{50} value) $8.082 \pm 0.055 \mu\text{g/mL}$ and that of standard ascorbic acid (IC_{50} value) was $5.645 \pm 0.034 \mu\text{g/mL}$, indicating cream had a potent antioxidant property. All of the antioxidant activities demonstrated by herbal creams indicated that the formulation was enriched with antioxidant chemicals such as polyphenol compounds or flavonoids, which are required for any phytocosmetic cream for skin anti-aging purposes because antioxidant properties maintain skin stress and turnover rate by interfering with any ROS in the body. The hydrophilic polyphenolic compounds from green tea, like GA, GC, C, EC, EGC, ECG, and EGCG, are more responsible for antioxidant activities in this formulated cream from other synthetically loaded actives like LMWHA (low molecular weight hyaluronic acid), Retinol, and Niacinamide (Forester and Lambert, 2011; Wahab *et al.*, 2021), since the aqueous extract design D3 TPC (mg GAE/g) was significantly correlated with DPPH activity (R^2 value 0.562) than TFC (mg QE/g). However, loading essential oil of *O. tenuiflorum* (IC_{50} value $32.073 \pm 0.020 \mu\text{g/mL}$) in this cream demonstrated a good synergistic antioxidant effect with herbal aqueous extracts because the cream showed a low antioxidant effective concentration value as compared to loaded essential oil and aqueous designs, as it is believed that loading complex plant extracts with essential oil shows synergistic antioxidant effect in any cosmetic formulation (Jadoon *et al.*, 2015).

Kojic acid (IC_{50} value 12.535 ± 0.098) inhibited the tryosinase enzyme more strongly than herbal cream (IC_{50} value 22.254 ± 0.369). However, because both kojic acid and herbal cream showed comparable inhibition concentrations, this formulated cream also strongly showed potent tyrosinase inhibition. This high tryosinase activity was attributed to the combination of hyaluronic acid with licorice plant extracts (liquirtin, glabridin, glabrene, and isoliquirtin) (Chaiyana *et al.*, 2020; Ahuja *et al.*, 2021). Other actives, such as retinol, niacinamide, green tea extracts, and essential oils, also contributed to the inhibition of mushroom tyrosinase activity. This tryosinase inhibition strategy will be an excellent starting point for the skin anti-aging process because it will cure skin hyperpigmentation, which is the first sign of aging

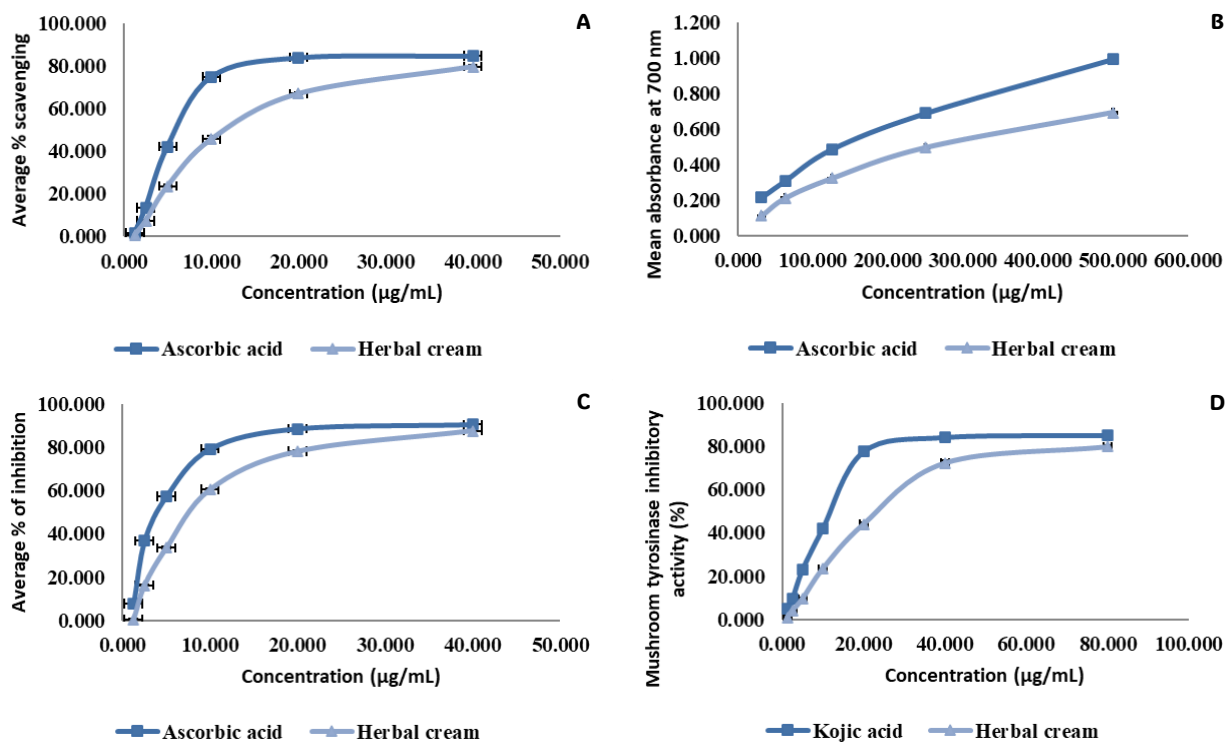


Figure 7. Antioxidant activities and mushroom tyrosinase inhibition shown by herbal cream at various concentrations, A: DPPH radical scavenging activity, B: FRAP assay, C: ABTS radical scavenging activity and D: mushroom tyrosinase inhibition activity. Data presented are in mean \pm SD (n = 3).

Table 4. Herbal cream antioxidant activities (DPPH scavenging activity, FRAP assay and ABTS scavenging activity) data at room temperature and accelerated stability study (freeze-thaw cycling).

Duration (months)	Room temperature			Freeze-thaw cycling		
	DPPH activity (IC ₅₀ , μ g/mL)	FRAP assay (EC ₅₀ , μ g/mL)	ABTS activity (IC ₅₀ , μ g/mL)	DPPH activity (IC ₅₀ , μ g/mL)	FRAP assay (EC ₅₀ , μ g/mL)	ABTS activity (IC ₅₀ , μ g/mL)
0	13.764 \pm 0.153	301.445 \pm 1.709	8.082 \pm 0.055	12.894 \pm 0.103	300.871 \pm 2.001	7.985 \pm 0.225
1	13.284 \pm 0.255	298.542 \pm 0.564	8.075 \pm 1.254	12.845 \pm 0.158	300.744 \pm 0.471	7.994 \pm 0.364
2	13.294 \pm 0.345	301.024 \pm 0.366	8.014 \pm 1.244	12.864 \pm 0.361	299.687 \pm 0.334	7.962 \pm 1.104
3	13.505 \pm 0.399	300.014 \pm 0.214	8.039 \pm 0.874	12.860 \pm 0.247	300.101 \pm 0.124	7.914 \pm 0.941

Values are presented as mean \pm SD of independent triplicate (n = 3).

(Di Petrillo *et al.*, 2016; Vichit and Saewan, 2022).

3.8 Stability study

The results of cream spreadability, pH, and centrifugation tests for three months (0, 1, 2, and 3) under both normal and accelerated time conditions are shown in Table 3. The antioxidant activities of the cream in both real-time and accelerated time were not considerably impacted (Table 4), this means that all loaded herbal or synthetic actives remained stable during the analysis period without noticeably degrading (Tan *et al.*, 2022). This evidenced that the formulation of phytocosmetic cream was good in terms of stability for long-term storage, which is a necessary quality for any stable cosmetic cream.

4. Conclusion

The crude aqueous extracts of *C. sinensis* leaves and *G. glabraroots* had good TPC and TFC properties, while

GTPs like GA, GC, C, EC, EGC, ECG, and EGCG were more correlated with antioxidant properties than TFC; however, the essential oil of *O. tenuiflorum* showed stronger antioxidant properties than all. When these phytoextracts were loaded on cream emulsions containing synthetic actives like retinol, hyaluronic acid, and niacinamide, the cream demonstrated a synergistic antioxidant effect with significant mushroom tyrosinase inhibition. Because the cream was made with a natural fragrance from *O. basilicum* essential oil and superiorcosmetic-grade chemicals, the physical and chemical characteristics of the cream demonstrated acceptance results in both real and accelerated time zones. This finding suggests the successful commercialization of phytocosmetic creams that contain local crude plant aqueous extracts rich with essential oils that are extremely concentrated in antioxidants and mushroom tyrosinase inhibition properties, a first-step natural remedy for skin aging.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgements

The Medicinal Plant Research (MPR) laboratory, where work was done and supervised, is acknowledged by the authors. We appreciate the Department of Pharmacy at Kathmandu University for the assistance and support with this project. We would like to thank the Korean International Cooperation Agency (KOICA), Nepal, for Research and Business Development (R&BD) programmed in the Kathmandu University Integrated Rural Development Project (KU-IRDP) under the topic "HERBAL HEALTH PROJECT" at Kathmandu University, Nepal.

References

- Agbor, G.A., Vinson, J.A. and Donnelly, P.E. (2014). Folin-ciocalteau reagent for polyphenolic assay. *International Journal of Food Science*, 3(8), 147-156. <https://doi.org/10.19070/2326-3350-1400028>
- Ahmed, A.A. and Nath, B.I.P.U.L. (2017). Formulation and in vitro evaluation of polyherbal anti-aging face cream of *Coriandrum sativum* and *Rose hip* oil. *International Journal of Current Pharmaceutical Research*, 9(4), 75-78. <https://doi.org/10.22159/ijcpr.2017v9i4.20961>
- Ahuja, A., Gupta, J. and Gupta, R. (2021). Miracles of herbal phytomedicines in treatment of skin disorders: natural healthcare perspective. *Infectious Disorders-Drug Targets*, 21(3), 328-338. <https://doi.org/10.2174/1871526520666200622142710>
- Alani, J.I., Davis, M.D. and Yiannias, J.A. (2013). Allergy to cosmetics: a literature review. *Dermatitis*, 24(6), 283-290. <https://doi.org/10.1097/DER.0b013e3182a5d8bc>
- Almeida, I.F., Maleckova, J., Saffi, R., Monteiro, H., Goios, F., Amaral, M.H. and Bahia, M.F. (2015). Characterization of an antioxidant surfactant-free topical formulation containing *Castanea sativa* leaf extract. *Drug Development and Industrial Pharmacy*, 41(1), 148-155. <https://doi.org/10.3109/03639045.2013.850712>
- Awah, F.M. and Verla, A.W. (2010). Antioxidant activity, nitric oxide scavenging activity and phenolic contents of *Ocimum gratissimum* leaf extract. *Journal of Medicinal Plants and Research*, 4 (24), 2479-2487. <https://doi.org/10.5897/JMPR10.262>
- Bonte, F., Girard, D., Archambault, J.C. and Desmouliere, A. (2019). Skin changes during aging. In Harris, J. and Korolchuk, V. (Eds.) *Biochemistry and Cell Biology of Aging: Part II Clinical Science*. Subcellular Biochemistry, 91, 249-280. Singapore: Springer. https://doi.org/10.1007/978-981-13-3681-2_10
- Cao, C., Xiao, Z., Wu, Y. and Ge, C. (2020). Diet and skin aging-from the perspective of food nutrition. *Nutrients*, 12(3), 870. <https://doi.org/10.3390/nu12030870>
- Carocho, M. and Ferreira, I.C. (2013). A review on antioxidants, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and Chemical Toxicology*, 51, 15-25. <https://doi.org/10.1016/j.fct.2012.09.021>
- Chaiyana, W., Sirithunyalug, J., Somwongin, S., Punyoyai, C., Laothaweerungsawat, N., Marsup, P., Neimkhum, W. and Yawootti, A. (2020). Enhancement of the antioxidant, anti-tyrosinase, and anti-hyaluronidase activity of *Morus alba* l. leaf extract by pulsed electric field extraction. *Molecules*, 25(9), 2212. <https://doi.org/10.3390/molecules25092212>
- Chaudhary, A., Sharma, S., Mittal, A., Gupta, S. and Dua, A. (2020). Phytochemical and antioxidant profiling of *Ocimum sanctum*. *Journal of Food Science and Technology*, 57(10), 3852-3863. <https://doi.org/10.1007/s13197-020-04417-2>
- Dąbrowska, A.K., Spano, F., Derler, S., Adlhar, C., Spencer, N.D. and Rossi, R.M. (2018). The relationship between skin function, barrier properties, and body-dependent factors. *Skin Research and Technology*, 24(2), 165-174. <https://doi.org/10.1111/srt.12424>
- Di Petrillo, A., Gonzalez-Paramas, A.M., Era, B., Medda, R., Pintus, F., Santos-Buelga, C. and Fais, A. (2016). Tyrosinase inhibition and antioxidant properties of *Asphodelus microcarpus* extracts. *BMC Complementary and Alternative Medicine*, 16(1), 453. <https://doi.org/10.1186/s12906-016-1442-0>
- Draelos, Z.D. (2019). Cosmeceuticals: What's real, what's not. *Dermatologic Clinics*, 37(1), 107-115. <https://doi.org/10.1016/j.det.2018.07.001>
- Eckhart, L., Tschachler, E. and Gruber, F. (2019). Autophagic control of skin aging. *Frontiers in Cell and Developmental Biology*, 7, 143. <https://doi.org/10.3389/fcell.2019.00143>
- Forester, S.C. and Lambert, J.D. (2011). The role of antioxidant versus pro-oxidant effects of green tea polyphenols in cancer prevention. *Molecular Nutrition and Food Research*, 55(6), 844-854. <https://doi.org/10.1002/mnfr.201000641>
- Genwali, G.R., Acharya, P.P. and Rajbhandari, M. (2013). Isolation of gallic acid and estimation of total

- phenolic content in some medicinal plants and their antioxidant activity. *Nepal Journal of Science and Technology*, 14(1), 95-102. <https://doi.org/10.3126/njst.v14i1.8928>
- Glusac, J., Davidesko-Vardi, I., Isaschar-Ovdat, S., Kukavica, B. and Fishman, A. (2018). Gel-like emulsions stabilized by tyrosinase-crosslinked potato and zein proteins. *Food Hydrocolloids*, 82, 53-63. <https://doi.org/10.1016/j.foodhyd.2018.03.046>
- Gyawali, R., Gupta, R.K., Shrestha, S., Joshi, R. and Paudel, P.N. (2020). Formulation and evaluation of polyherbal cream containing *Cinnamomum zeylanicum* Blume, *Glycyrrhizaglabra* L and *Azadirachta indica* A. Juss. extracts to topical use. *Journal of Institute of Science and Technology*, 25 (2), 61-71. <https://doi.org/10.3126/jist.v25i2.33738>
- Hasan, S.R., Hossain, M.M., Akter, R., Jamila, M., Mazumder, M.E.H. and Rahman, S. (2009). DPPH free radical scavenging activity of some Bangladeshi medicinal plants. *Journal of Medicinal Plants Research*, 3(11), 875-879.
- Jadoon, S, Karim, S., Bin Asad, M.H., Akram, M.R., Khan, A.K., Malik, A., Chen, C. and Murtaza, G. (2015). Anti-aging potential of phytoextract loaded-pharmaceutical creams for human skin cell longevity. *Oxidative Medicine and Cellular Longevity*, 2015, 709628. <https://doi.org/10.1155/2015/709628>
- Khavkin, J. and Ellis, D.A. (2011). Aging skin: Histology, physiology, and pathology. *Facial Plastic Surgery Clininics*, 19(2), 229-234. <https://doi.org/10.1016/j.fsc.2011.04.003>
- Korhonen, M., Hellen, L., Hirvonen, J. and Yliruusi, J. (2001). Rheological properties of creams with four different surfactant combinations-effect of storage time and conditions. *International Journal of Pharmaceutics*, 221(1-2), 187-196. [https://doi.org/10.1016/S0378-5173\(01\)00675-5](https://doi.org/10.1016/S0378-5173(01)00675-5)
- Kumar, S., Sandhir, R. and Ojha, S. (2014). Evaluation of antioxidant activity and total phenol in different varieties of *Lantana camara* leaves. *BMC Research Notes*, 7, 560. <https://doi.org/10.1186/1756-0500-7-560>
- Kurzawa, M., Wilczynska, E., Brudzynska, P. and Sionkowska, A. (2022). Total phenolic content, antioxidant capacity and UV radiation protection properties of marigold (*Calendula officinalis*), carrot (*Daucus carota*), tomato (*Solanum lycopersicum*) and hop (*Humulus lupulus*) extracts. *Cosmetics*, 9(6), 134. <https://doi.org/10.3390/cosmetics9060134>
- Liao, R., Parker, T., Bellerose, K., Vollmer, D. and Han, X. (2022). A green tea containing skincare system improves skin health and beauty in adults: An exploratory controlled clinical study. *Cosmetics*, 9 (5), 86. <https://doi.org/10.3390/cosmetics9050096>
- Lohani, A., Verma, A., Hema, G. and Pathak, K. (2021). Topical delivery of geranium/calendula essential oil-entrapped ethanolic lipid vesicular cream to combat skin aging. *Biomed Research International*, 2021, 4593759. <https://doi.org/10.1155/2021/4593759>
- Madaan, R., Bansal, G., Kumar, S. and Sharma, A. (2011). Estimation of total phenols and flavonoids in extracts of *Actaeaspicata* roots and antioxidant activity studies. *Indian Journal of Pharmaceutical Sciences*, 73(6), 666-669. <https://doi.org/10.4103/0250-474X.100242>
- Mishra, A.P., Saklani, S., Milella, L. and Tiwari, P. (2014). Formulation and evaluation of herbal antioxidant face cream of *Nardostachys jatamansi* collected from Indian Himalayan region. *Asian Pacific Journal of Tropical Biomedicine*, 4 (Supplement 2), S679-S682. <https://doi.org/10.12980/APJTB.4.2014APJTB-2014-0223>
- Mungmai, L., Preedalikit, W., Aunsri, N. and Peerakam, N. (2019). Bioactivity test and GC-MS analysis of different solvent extracts from *Perilla frutescens* (Linn.) Britton and cosmetic product application for sensitive skin. *Progress in Applied Science and Technology*, 9(2), 78-93.
- Pandey, B.P., Pradhan, S.P., Joshi, P. and Adhikari, K. (2022). Antioxidant and enzymes inhibitory activities of leaf extracts of plant species traditionally used for medicinal and spiritual purposes in Nepal. *Journal of Herbs, Spices and Medicinal Plants*, 28(3), 265-280. <https://doi.org/10.1080/10496475.2022.2053026>
- Park, Y.S., Jung, S.T., Kang, S.G., Heo, B.G., Arancibia-Avila, P., Toledo, F. and Gorinstein, S. (2008). Antioxidants and proteins in ethylene-treated kiwifruits. *Food Chemistry*, 107(2), 640-648. <https://doi.org/10.1016/j.foodchem.2007.08.070>
- Paudel, P.N., Satyal, P., Satyal, R., Setzer, W.N. and Gyawali, R. (2022). Chemical composition, enantiomeric distribution, antimicrobial and antioxidant activities of *Origanum majorana* L. essential oil from Nepal. *Molecules*, 27(18), 6136. <https://doi.org/10.3390/molecules27186136>
- Phosri, S., Kiattisin, K., Intharuksa, A., Janon, R., Na Nongkhai, T. and Theansungnoen, T. (2022). Anti-aging, anti-acne, and cytotoxic activities of *Houttuynia cordata* extracts and phytochemicals analysis by LC-MS/MS. *Cosmetics*, 9(6), 136. <https://doi.org/10.3390/cosmetics9060136>
- Pinto, D., Lameirao, F., Delerue-Matos, C., Rodrigues,

- F. and Costa, P. (2021). Characterization and stability of a formulation containing antioxidants-enriched *Castanea sativa* shells extract. *Cosmetics*, 8 (2), 49. <https://doi.org/10.3390/cosmetics8020049>
- Plyduang, T., Atipairin, A., Sae Yoon, A., Sermkaew, N., Sakdiset, P. and Sawatdee, S. (2022). Formula development of red palm (*Elaeis guineensis*) fruit extract loaded with solid lipid nanoparticles containing creams and its anti-aging efficacy in healthy volunteers. *Cosmetics*, 9(1), 3. <https://doi.org/10.3390/cosmetics9010003>
- Puebla-Barragan, S. and Reid, G. (2021). Probiotics in cosmetic and personal care products: Trends and challenges. *Molecules*, 26(5), 1249. <https://doi.org/10.3390/molecules26051249>
- Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M. and Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine*, 26(9-10), 1231-1237. [https://doi.org/10.1016/S0891-5849\(98\)00315-3](https://doi.org/10.1016/S0891-5849(98)00315-3)
- Restu, W.K., Sampora, Y., Meliana, Y. and Haryono, A. (2015). Effect of accelerated stability test on characteristics of emulsion systems with chitosan as a stabilizer. *Procedia Chemistry*, 16, 171-176. <https://doi.org/10.1016/j.proche.2015.12.031>
- Salehi, N., Mortazavi, S.M. and Moghimi, H. (2022). Investigating the changes in cream properties following topical application and their influence on the product efficiency. *Iranian Journal of Pharmaceutical Research*, 21(1), e123946. <https://doi.org/10.5812/ijpr.123946>
- Shah, P. and Modi, H.A. (2015). Comparative study of DPPH, ABTS and FRAP assays for determination of antioxidant activity. *International Journal for Research in Applied Science and Engineering Technology*, 3(6), 636-641.
- Siahaan, E.A., Agusman, Pangestuti, R., Shin, K.H. and Kim, S.K. (2022). Potential cosmetic active ingredients derived from marine by-products. *Marine Drugs*, 20(12), 734. <https://doi.org/10.3390/md20120734>
- Srisuksomwong, P., Kaehin, L. and Mungmai, L. (2023). Collagenase and tyrosinase inhibitory activities and stability of facial cream formulation containing cashew leaf extract. *Cosmetics*, 10(1), 17. <https://doi.org/10.3390/cosmetics10010017>
- Subramaniam, V.D., Prasad, S.V., Banerjee, A., Gopinath, M., Murugesan, R., Marotta, F., Sun, X.F. and Pathak, S. (2019). Health hazards of nanoparticles: Understanding the toxicity mechanism of nanosized ZnO in cosmetic products. *Drug and Chemical Toxicology*, 42(1), 84-93. <https://doi.org/10.1080/01480545.2018.1491987>
- Tan, P.L., Rajagopal, M., Chinnappan, S., Selvaraja, M., Leong, M.Y., Tan, L.F. and Yap, V.L. (2022). Formulation and physicochemical evaluation of green cosmeceutical herbal face cream containing standardized mangosteen peel extract. *Cosmetics*, 9 (3), 46. <https://doi.org/10.3390/cosmetics9030046>
- Tungmunnithum, D., Thongboonyou, A., Pholboon, A. and Yangsabai, A. (2018). Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: An overview. *Medicines*, 5(3), 93. <https://doi.org/10.3390/medicines5030093>
- Vichit, W. and Saewan, N. (2022). Anti-oxidant and anti-aging activities of callus culture from three rice varieties. *Cosmetics*, 9(4), 79. <https://doi.org/10.3390/cosmetics9040079>
- Wahab, S., Annadurai, S., Abullais, S.S., Das, G., Ahmad, W., Ahmad, M.F., Kandasamy, G., Vasudevan, R., Ali, M.S. and Amir, M. (2021). Glycyrrhizaglabra (Licorice): A comprehensive review on its phytochemistry biological activities, clinical evidence and toxicology. *Plants*, 10(12), 2751. <https://doi.org/10.3390/plants10122751>
- Whangsomnuek, N., Mungmai, L., Mengamphan, K. and Amornlerdpison, D. (2019). Efficiency of skin whitening cream containing *Etilingera elatior* flower and leaf extracts in volunteers. *Cosmetics*, 6(3), 39. <https://doi.org/10.3390/cosmetics6030039>
- Yadav, N.P., Rai, V.K., Mishra, N., Sinha, P., Bawankule, D.U., Pal, A. and Chanotiya, C.S. (2014). A novel approach for development and characterization of effective mosquito repellent cream formulation containing citronella oil. *BioMed Research International*, 2014, 786084. <https://doi.org/10.1155/2014/786084>
- Yeh, S.J., Lin, J.F. and Chen, B.S. (2021). Multiple-molecule drug design based on systems biology approaches and deep neural network to mitigate human skin aging. *Molecules*, 26(11), 3178. <https://doi.org/10.3390/molecules26113178>
- Zhang, S. and Duan, E. (2018). Fighting against skin aging: The way from bench to bedside. *Cell Transplant*, 27(5), 729-738. <https://doi.org/10.1177/0963689717725755>