

Antioxidant capacity of pitcher (*Nepenthes*) extract on oxidative rancidity of sun-dried squid

*Minh, N.P.

Faculty of Biotechnology, Binh Duong University, Vietnam

Article history:

Received: 6 June 2021

Received in revised form: 23 July 2021

Accepted: 18 September 2021

Available Online: 3 October 2021

Keywords:

Pitcher extract,
Dried squid,
Acid value,
Carbonyl value,
Peroxide value,
Oxidative rancidity

Abstract

Consumers demand new biologically natural-active ingredients with a great capacity to promote a healthier advantage and free from toxicity. Pitcher (*Nepenthes*) is one of the naturally carnivorous climber herbs. Extract from *Nepenthes* plant contains numerous beneficial bioactivity characteristics. This research compared the impact of 0.05% pitcher extract demonstrated with 10 ppm butylated hydroxyanisole on the accumulation of acid value, peroxide value, carbonyl value and thiobarbituric acid during 12 months of storage. Results showed that there was no significant difference between 0.05% pitcher extract and 10 ppm butylated hydroxyanisole in respect of acid value, peroxide value, carbonyl value and thiobarbituric acid during preservation of the dried squid. Pitcher extract would be a promising alternative to replace synthetic antioxidants to overcome oxidative rancidity in the dried seafood.

DOI:

[https://doi.org/10.26656/fr.2017.5\(5\).404](https://doi.org/10.26656/fr.2017.5(5).404)

1. Introduction

Pitcher (*Nepenthes*) is an important tropical carnivorous pitcher plant widely distributed in Vietnam and Southeast Asian countries. Pitcher (*Nepenthes*) has a unique pitcher organ growing from the tip of the leaf (Wang *et al.*, 2009). It is able to trap and digest insects to obtain nutrients (Wan-Nor-Adibah-Wan *et al.*, 2016). *Nepenthes* are commonly utilized as a folk remedy to cure different ailments. It has been reported to be effective to cure gastrointestinal discomfort, regulate the menstrual cycle, ease child-birth, relieve asthma, treat eye inflammation, gastric ulcer, jaundice, high blood pressure and astringent (Aung *et al.*, 2002; Schwallier *et al.*, 2015; Van Thanh *et al.*, 2015; Thao *et al.*, 2016; Sanusi *et al.*, 2017). Its extract contains numerous phytochemical constituents like steroids, flavonoids, alkaloids, terpenoids, tannins (Shuaibu *et al.*, 2017). Extract from this plant is responsible for pharmacological properties like cytotoxicity, anti-inflammatory, antibacterial, antidiabetic, antifungal, antioxidant, antimalarial, antiosteoporotic and hypolipidemic potentials (Likhitwitayawuid *et al.*, 1998; Shil *et al.*, 2010; Ismail *et al.*, 2015; Van *et al.*, 2015).

Lipid oxidation induces decomposition and degradation in three steps: initiation, propagation, and termination (Frankel, 1984; Fraser and Sumar, 1998). Rancidity in seafood could be initiated and propagated

by peroxidase, lipoxygenase, auto-oxidation, photosensitized oxidation (Hsieh and Kinsella, 1989). Squid is one of the most important seafood species. There is increased consumer demand for dried squid in the world. It is rich in unsaturated fatty acids therefore it is susceptible to oxidative rancidity (Losada *et al.*, 2007). Lipid oxidation results in protein denaturation leading to lower quality of the dried product (Lugasi *et al.*, 2007; Beck, 2014). Artificial antioxidants such as butylated hydroxyanisole, butylated hydroxytoluene, tertiary butylated hydroxyl quinone and propyl gallate are commonly used in the seafood industry to control lipid oxidation. However, there are different health concerns when using these substances (Patel *et al.*, 2010; Carocho and Ferreira, 2013; Reddy and Grace, 2016). Natural extracts with bioactive functions are highly preferred to replace synthetic antioxidants (Barros *et al.*, 2007; Thao *et al.*, 2016). Phenol-rich plant extracts such as the summer savoury flavoured powder, beetroot leaf extract and olive polyphenol successfully inhibited lipid and protein oxidation (Stina *et al.*, 2020). Natural antioxidants from fruits, vegetables, herbs, and spices included beneficial essential oils suitable for seafood preservation by preventing oxidative chain reactions and prolonging the stability of these products (Lorenzo *et al.*, 2018; Smaou *et al.*, 2019; Sood *et al.*, 2020; Alirezalu *et al.*, 2020; Munekata *et al.*, 2020; Dominguez *et al.*, 2020; Mirian *et al.*, 2021). Plant extract had antifungal,

*Corresponding author.

Email: npminh@bdu.edu.vn

antioxidant, antimutagenic activities, and retard lipid oxidation in seafood (Cando *et al.*, 2014; Ghayempour *et al.*, 2015; Ashrafi *et al.*, 2018; Kharchoufi *et al.*, 2018). There was not any research that mentioned the utilization of pitcher extract to control lipid oxidation in the dried squid during storage. Therefore, the purpose of our study focused on the influence of pitcher extract demonstrated with butylated hydroxyanisole on the accumulation of acid value, peroxide value, carbonyl value and thiobarbituric acid during 12 months of storage.

2. Materials and methods

2.1 Materials

Raw squids were obtained from Tran De district, Soc Trang province, Vietnam. *Nepenthes* was collected from Da Lat district, Lam Dong province, Vietnam. A 100 g of *Nepenthes* was extracted under an alcoholic-aqueous solution (70% v/v) at 45°C for 3 hrs. The *Nepenthes* extract was obtained via filtration (Whatman paper No. 1). Chemical reagents were all analytical grade purchased from Fluka (Switzerland) and Sigma Aldrich (USA).

2.2 Methods

Squids were preliminarily gutted and washed under tap water to remove the ink. They were soaked in *Nepenthes* extract, butylated hydroxyanisole (BHA) solution (10 ppm) and control (without any additive) for 90 mins. They were dried under sunlight for 3 consecutive days to obtain a final moisture content of around 18%. The dried squids were kept in plastic bags for 12 months. In 3 month-interval, they were sampled to analyse the acid value, peroxide value, carbonyl value, thiobarbituric acid (TBA) value.

Free radical scavenging activity (DPPH, mg Trolox/100 g) of *Nepenthes* extract was evaluated by the method described by Bakar *et al.* (2017). Acid value (mg KOH/g) was determined by titration by 0.1 N KOH with phenolphthalein indicator to pink appearance. Peroxide value (meq/kg) was estimated by weighing 5 g of sample, adding 15 mL acetic acid and 10 mL chloroform, supplementing 1 mL KI solution. This mixture was kept in a dark place for 15 mins and a starch indicator was added. The final titration was defined by adding 0.01 N Na₂S₂O₃ until colourless. Carbonyl value (meq/kg) was determined by high-performance liquid chromatography with ultraviolet (UV) detection (Vanessa and Zenilda, 2013). Thiobarbituric acid (TBA) value (mg malonaldehyde/kg) was measured by the 2-thiobarbituric acid spectrophotometric method (Anna *et al.*, 2017).

2.3 Statistical analysis

The experiments were run in triplicates with different groups of samples. The data were presented as mean±standard deviation. Statistical analysis was performed by the Statgraphics Centurion version XVI.

3. Results and discussion

Free radical scavenging activity or DPPH (mg Trolox/100 g) of *Nepenthes* extract was shown at 75.83±0.03 (mg Trolox/100 g) compared to butylated hydroxyanisole (BHA) solution 77.09±0.01 (mg Trolox/100 g). *Nepenthes* extract has excellent antioxidant capacity ideal for lipid oxidative prevention. Excellent antioxidant activity of pitcher extract might have originated from a synergistic impact of terpenes, terpenoids, essential oils and other available metabolites such as flavonoids (Adhikari *et al.*, 2003; Grassmann, 2005; Ozturk, 2012). Tiewlasubon *et al.* (2014) reported that *Nepenthes* extract showed powerful antioxidant capacity with 23.33±0.441 (mg Trolox/100 g). The leaf *Nepenthes* extract was demonstrated to be more effective than standard butylated hydroxytoluene (Ismail *et al.*, 2015). In another research, the radical scavenging activity of the pitcher extract was noted at 0.148±0.04 mg/mL (Muhammad *et al.*, 2018).

The acid value is commonly applied as an important indicator of fat rancidity. Under the incorporation of *Nepenthes* extract in the soaking solution, the dried squid had a low acid value similar to the sample treated by BHA during 12 months of storage. Meanwhile, the control sample had accumulated acid value every month (Table 1). At the end of 12 months of storage, the dried squid treated with *Nepenthes* extract still had a low acid value (2.70±0.03 mg KOH/g). It is suggested that *Nepenthes* extract significantly retarded the rancidity in dried squid. These results were in accordance with other findings such as the herbal extract that was demonstrated to be effective in the control of lipid oxidation in dried eel (Song, 2019). Furthermore, green tea extract was also suitable to retard fat rancidity in mackerel (Nam *et al.*, 2011). While the combination of spices (turmeric, chilli) greatly improved the quality of dried Taki (Rahman *et al.*, 2017). *Eryngium caucasicum* extracts were also as effective to slow down the acid value of silver carp fillets (Raiesi *et al.*, 2017).

Primary lipid oxidation was followed by the peroxide value (Servet and Hudayi, 2011). Peroxide values (meq/kg) of the dried squid treated by *Nepenthes* extract, butylated hydroxyanisole and control were all presented in Table 2. The supplementation of *Nepenthes* extract was able to maintain the peroxide value of the dried squid at a low amount (2.91±0.01 meq/kg) after 12

Table 1. Acid value (mg KOH/g) of the dried squid

Storage (month)	0	3	6	9	12
<i>Nepenthes</i> extract 0.05%	1.06±0.02 ^{ab}	1.45±0.00 ^{ab}	1.94±0.01 ^b	2.32±0.00 ^b	2.70±0.03 ^b
Butylated hydroxyanisole 10 ppm	0.98±0.01 ^b	1.29±0.03 ^b	1.85±0.02 ^b	2.21±0.01 ^b	2.59±0.02 ^b
Control	1.17±0.00 ^a	1.97±0.02 ^a	4.83±0.03 ^a	7.69±0.02 ^a	10.85±0.01 ^a

Values are presented as mean±SD, n = 3. Values with the same superscript within the same row are not significantly different ($\alpha = 5\%$).

months of storage. Meanwhile, the peroxide value of the control sample increased dramatically from the 3rd to the 12th month. There was no significant difference in the peroxide value between those treated with *Nepenthes* extract and butylated hydroxyanisole. It is suggested that *Nepenthes* extract could safely be used and effectively minimize lipid oxidation. The effectiveness of rancidity retardation by *Nepenthes* extract was similar to other plant extracts such as the green tea extract that strongly retards the oxidative rancidity in eel oil (Song and Kim, 2018). The peroxide value of anchovy oil was kept at low content by green tea extract (Kang *et al.*, 2007). Basil leaf essential oil was also effective to limit peroxide value in Sea bass slices (Arfat *et al.*, 2015). Pomegranate peel extract was also appropriated for Nile tilapia fillets to slow down peroxide value (Alsaggaf *et al.*, 2017). *Eryngium caucasicum* extract was found to be a beneficial substance to decrease peroxide value in Silver carp fillets (Raiesi *et al.*, 2017).

Carbonyl is commonly originated from proteolysis and amino acid oxidation to release malondialdehyde. Carbonyl value (meq/kg) of the dried squid soaked in *Nepenthes* extract, butylated hydroxyanisole and control were all presented in Table 3. It was thoroughly realized that with the addition of *Nepenthes* extract, low traces of

carbonyl values of the dried squid were obtained (2.66±0.00 meq/kg) after 12 months of storage. Meanwhile, the carbonyl value of the control sample increased steeply from the 3rd to the 12th month. There was no significant difference in carbonyl value between the squids that were treated by *Nepenthes* extract and butylated hydroxyanisole. It was recommended that *Nepenthes* extract could be utilized to replace butylated hydroxyanisole to control the carbonyl substance. This result was relevant to another study when eels treated with green tea extract had a low carbonyl value (Choi, 2006).

In the evaluation of oxidative rancidity, the thiobarbituric acid (TBA) value should be mentioned. It is an important quality indicator to measure fat oxidation for dried and semi-dried seafood (Bahar *et al.*, 2006). The basic mechanism of this procedure is the interaction of 1 molecule of malonaldehyde and 2 molecules of TBA to release a purple appearance of the malonaldehyde-TBA complex. Table 4 shows the influence of *Nepenthes* extract demonstrated with butylated hydroxyanisole to control the TBA value. It was found that the *Nepenthes* extract effectively retarded TBA formation in the dried squid during 12 months of storage. These results were relevant to fennel extracts

Table 2. Peroxide value (meq/kg) of the dried squid

Storage (month)	0	3	6	9	12
<i>Nepenthes</i> extract 0.05%	1.34±0.03 ^a	1.75±0.02 ^b	2.03±0.00 ^b	2.48±0.02 ^b	2.91±0.01 ^b
Butylated hydroxyanisole 10 ppm	1.28±0.02 ^a	1.69±0.01 ^b	1.98±0.03 ^b	2.37±0.00 ^b	2.80±0.00 ^b
Control	1.42±0.03 ^a	4.27±0.03 ^a	7.93±0.01 ^a	11.86±0.03 ^a	18.70±0.02 ^a

Values are presented as mean±SD, n = 3. Values with the same superscript within the same row are not significantly different ($\alpha = 5\%$).

Table 3. Carbonyl value (meq/kg) of the dried squid

Storage (month)	0	3	6	9	12
<i>Nepenthes</i> extract 0.05%	0.83±0.01 ^a	1.15±0.03 ^b	1.59±0.01 ^b	1.94±0.03 ^b	2.66±0.00 ^b
Butylated hydroxyanisole 10 ppm	0.79±0.00 ^a	1.04±0.02 ^b	1.48±0.02 ^b	1.87±0.03 ^b	2.59±0.01 ^b
Control	0.84±0.00 ^a	3.68±0.03 ^a	8.53±0.00 ^a	14.73±0.02 ^a	19.93±0.00 ^a

Values are presented as mean±SD, n = 3. Values with the same superscript within the same row are not significantly different ($\alpha = 5\%$).

Table 4. Thiobarbituric acid value (mg malonaldehyde/kg) of the dried squid

Storage (month)	0	3	6	9	12
<i>Nepenthes</i> extract 0.05%	1.07±0.03 ^a	1.70±0.00 ^b	2.02±0.02 ^b	2.68±0.01 ^b	2.95±0.03 ^b
Butylated hydroxyanisole 10 ppm	1.02±0.01 ^a	1.63±0.00 ^b	1.97±0.01 ^b	2.59±0.00 ^b	2.93±0.02 ^b
Control	1.13±0.02 ^a	5.54±0.01 ^a	9.13±0.03 ^a	16.07±0.00 ^a	23.46±0.01 ^a

Values are presented as mean±SD, n = 3. Values with the same superscript within the same row are not significantly different ($\alpha = 5\%$).

that were ideal to increase the stability of silver carp fillets by slowing down TBA value (Mazandrani *et al.*, 2016). *Eryngium caucasicum* extract was also effective to limit the TBA value of silver carp fillets (Raiesi *et al.*, 2017).

4. Conclusion

Pitcher (*Nepenthes*) extract was rich in bioactive components, mainly polyphenols, which are valuable alternatives to replace synthetic antioxidants in seafood processing and preservation. Findings of this research concluded that Pitcher (*Nepenthes*) extract revealed excellent antioxidant potential to retard an accumulation of lipid oxidation in the dried squid. Its incorporation during soaking fresh squid would maintain the acid value, peroxide value, carbonyl value and TBA value at the low amount in the dried squid. It would be considered as an upcoming alternative to reduce the consumption of synthetic additives due to concerns of the possible toxic effects.

Conflict of interest

The author strongly confirms that this research was conducted with no conflict of interest.

Acknowledgement

We acknowledge the financial support for the publication provided by Binh Duong University, Thu Dau Mot city, Binh Duong province, Vietnam.

References

- Adhikari, S., Joshi, R., Patro, B., Ghanty, T., Chintalwar, G., Sharma, A., Chattopadhyay, S. and Mukherjee, T. (2003). Antioxidant activity of bakuchiol: Experimental evidences and theoretical treatments on the possible involvement of the terpenoid chain. *Chemical Research in Toxicology*, 16(9), 1062-1069. <https://doi.org/10.1021/tx034082r>
- Alirezalu, K., Pateiro, M., Yaghoubi, M., Alirezalu, A., Peighambardoust, S.H. and Lorenzo, J.M. (2020). Phytochemical constituents, advanced extraction technologies and techno-functional properties of selected Mediterranean plants for use in meat products. A comprehensive review. *Trends Food Science Technology*, 100(2), 292–306. <https://doi.org/10.1016/j.tifs.2020.04.010>
- Alsaggaf, M.S., Moussa, S.H. and Tayel, A.A. (2017). Application of fungal chitosan incorporated with pomegranate peel extract as edible coating for microbiological, chemical and sensorial quality enhancement of Nile tilapia fillets. *International Journal of Biological Macromolecules*, 99(6), 499–505. <https://doi.org/10.1016/j.ijbiomac.2017.03.017>
- Anna, R., Monika, S., Jozef, N., Slavomir, M., Boris, S., Milan, C. and Tatiana, K. (2017). Lipid peroxidation process in meat and meat products: a comparison study of malondialdehyde determination between modified 2-thiobarbituric acid spectrophotometric method and reverse-phase high-performance liquid chromatography. *Molecules*, 22(11), 1988. <https://doi.org/10.3390/molecules22111988>
- Arfat, Y.A., Benjakul, S., Vongkamjan, K., Sumpavapool, P. and Yarnpakdee, S. (2015). Shelf-life extension of refrigerated sea bass slices wrapped with fish protein isolate/fish skin gelatin-ZnO nanocomposite film incorporated with basil leaf essential oil. *Journal of Food Science and Technology*, 52(1), 6182–6193. <https://doi.org/10.1007/s13197-014-1706-y>
- Ashrafi, A., Jokar, M. and Mohammadi-Nafchi, A. (2018). Preparation and characterization of biocomposite film based on chitosan and kombucha tea as active food packaging. *International Journal of Biological Macromolecules*, 108(1), 444–454. <https://doi.org/10.1016/j.ijbiomac.2017.12.028>
- Aung, H., Chia, L., Goh, N., Chia, T., Ahmed, A., Pare, P. and Mabry, T. (2002). Phenolic constituents from the leaves of the carnivorous plant *Nepenthes gracilis*. *Fitoterapia*, 73(5), 445-447. [https://doi.org/10.1016/s0367-326x\(02\)00113-2](https://doi.org/10.1016/s0367-326x(02)00113-2)
- Bahar, T., Koray, K. and Deniz, A. (2006). Comparison of two thiobarbituric acid (TBA) method for monitoring lipid oxidation in fish. *European Journal of Fisheries and Aquatic Sciences*, 23(3), 331–334.
- Bakar, M.F.A., Sanusi, S.B., Bakar, F.I.A., Cong, O.J. and Mian, Z. (2017). Physicochemical and antioxidant potential of raw unprocessed honey from Malaysian stingless bees. *Pakistan Journal of Nutrition*, 16(11), 888-894. <https://doi.org/10.3923/pjn.2017.888.894>
- Barros, L., Ferreira, M.J., Queiros, B., Ferreira, I.C. and Baptista, P. (2007). Total phenols, ascorbic acid, β -carotene and lycopene in Portuguese wild edible mushrooms and their antioxidant activities. *Food Chemistry*, 103(2), 413-419. <https://doi.org/10.1016/j.foodchem.2006.07.038>
- Beck, C. (2014). Effects of high pressure processing on lipid oxidation: A review. *Innovative Food Science and Emerging Technology*, 22(2), 1–10. <https://doi.org/10.1016/j.ifset.2013.10.012>
- Cando, D., Morcuende, D., Utrera, M. and Estevez, M. (2014). Phenolic-rich extracts from Willowherb (*Epilobium hirsutum* L.) inhibit lipid oxidation but accelerate protein carbonylation and discoloration of beef patties. *European Food Research Technology*,

- 238(1), 741–751. <https://doi.org/10.1007/s00217-014-2152-9>
- Carocho, M. and Ferreira, I.C. (2013). A review on antioxidants, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and Chemical Toxicology*, 51(1), 15-25. <https://doi.org/10.1016/j.fct.2012.09.021>
- Choi, B.D., Kang, S.J., Ha, Y.L., Kim, S.Y. and Lee, J.J. (2006). Oxidative stability of lipids from eel (*Anguilla japonica*) fed conjugated linoleic acid. *Journal of the Korean Society of Food Science and Nutrition*, 35(1), 61-67. <https://doi.org/10.3746/jkfn.2006.35.1.061>
- Dominguez, R., Munekata, P.E.S., Pateiro, M., Maggiolino, A., Bohrer, B. and Lorenzo, J.M. (2020). Red beetroot. A potential source of natural additives for the meat industry. *Applied Sciences*, 10 (23), 8340. <https://doi.org/10.3390/app10238340>
- Frankel, E.N. (1984). Chemistry of free radical and singlet oxidation of lipids. *Progress in Lipid Research*, 23(4), 197–221. [https://doi.org/10.1016/0163-7827\(84\)90011-0](https://doi.org/10.1016/0163-7827(84)90011-0)
- Fraser, O. and Sumar, S. (1998). Compositional changes and spoilage in fish-an introduction. *Nutrition and Food Science*, 98(5), 275–279. <https://doi.org/10.1108/00346659810224208>
- Ghayempour, S., Montazer, M. and Mahmoudi, R.M. (2015). Tragacanth gum as a natural polymeric wall for producing antimicrobial nanocapsules loaded with plant extract. *International Journal of Biological Macromolecules*, 81(4), 514–520. <https://doi.org/10.1016/j.ijbiomac.2015.08.041>
- Grassmann, J. (2005). Terpenoids as plant antioxidants. *Vitamins and Hormones*, 72(1), 505-535. [https://doi.org/10.1016/S0083-6729\(05\)72015-X](https://doi.org/10.1016/S0083-6729(05)72015-X)
- Hsieh, R.J. and Kinsella, J.E. (1989). Oxidation of polyunsaturated fatty acids: Mechanisms, products, and inhibition with emphasis on fish. *Advance Food Nutrition Research*, 33(1), 233–341. [https://doi.org/10.1016/s1043-4526\(08\)60129-1](https://doi.org/10.1016/s1043-4526(08)60129-1)
- Ismail, N.A., Kamariah, A.S., Lim, L.B. and Ahmad, N. (2015). Phytochemical and pharmacological evaluation of methanolic extracts of the leaves of *Nepenthes bicalcarata* Hook. *F. International Journal of Pharmacognosy and Phytochemical Research*, 7(6), 1127-1138.
- Kang, S.T., Yoo, U.H., Nam, K.H., Kang, J.Y. and Oh, K.S. (2007). Antioxidative effects of green tea extract on the oxidation of anchovy oil. *Journal of Agriculture and Life Science*, 41(3), 47-53.
- Kharchoufi, S., Licciardello, F., Siracusa, L., Muratore, G., Hamdi, M. and Restuccia, C. (2018). Antimicrobial and antioxidant features of ‘Gabsi’ pomegranate peel extracts. *Industrial Crops and Products*, 111(1), 345–352. <https://doi.org/10.1016/j.indcrop.2017.10.037>
- Likhitwitayawuid, K., Kaewamatawong, R., Ruangrunsi, N. and Krungkrai, J. (1998). Antimalarial naphthoquinones from *Nepenthes thorelii*. *Planta Medicine*, 64(3), 237-241. <https://doi.org/10.1055/s-2006-957417>
- Lorenzo, J.M., Pateiro, M., Dominguez, R., Barba, F.J., Putnik, P., Kovacevic, D.B., Shpigelman, A., Granato, D. and Franco, D. (2018). Berries extracts as natural antioxidants in meat products: A review. *Food Research International*, 106(2), 1095–1104. <https://doi.org/10.1016/j.foodres.2017.12.005>
- Losada, V., Barros-Velázquez, J. and Aubourg, S.P. (2007). Rancidity development in frozen pelagic fish: Influence of slurry ice as preliminary chilling treatment. *LWT-Food Science and Technology*, 40 (6), 991–999. <https://doi.org/10.1016/j.lwt.2006.05.011>
- Lugasi, A., Losada, V., Hovari, J., Lebovics, V., Jakoczi, I. and Aubourg, S. (2007). Effect of pre-soaking whole pelagic fish in a plant extract on sensory and biochemical changes during subsequent frozen storage. *LWT-Food Science and Technology*, 40(5), 930–936. <https://doi.org/10.1016/j.lwt.2005.09.021>
- Mazandrani, H.A., Javadian, S. and Bahram, S. (2016). The effect of encapsulated fennel extracts on the quality of silver carp fillets during refrigerated storage. *Food Science Nutrition*, 4(2), 298–304. <https://doi.org/10.1002/fsn3.290>
- Muhammad, A.F.R., Kamalrul, A.A. and Hoe-Han, G. (2018). Antioxidant activity of pitcher extracts from three *Nepenthes* species. *Sains Malaysiana*, 47(12), 3069–3075. <https://doi.org/10.17576/jsm-2018-4712-17>
- Munekata, P.E.S., Gullon, B., Pateiro, M., Tomasevic, I., Dominguez, R. and Lorenzo, J.M. (2020). Natural antioxidants from seeds and their application in meat products. *Antioxidants*, 9(9), 815. <https://doi.org/10.3390/antiox9090815>
- Nam, K.H., Jang, M.S., Lee, D.S., Yoon, H.D. and Park, H.Y. (2011). Effect of green tea and lotus leaf boiled water extracts treatment on quality characteristics in salted mackerel during storage. *Korean Journal of Food Preservation*, 18(5), 643-650. <https://doi.org/10.11002/kjfp.2011.18.5.643>
- Ozturk, M. (2012). Anticholinesterase and antioxidant activities of savoury (*Satureja thymbra* L.) with identified major terpenes of the essential oil. *Food*

- Chemistry*, 134(1), 48-54. <https://doi.org/10.1016/j.foodchem.2012.02.054>
- Patel, V.R., Patel, P.R. and Kajal, S.S. (2010). Antioxidant activity of some selected medicinal plants in western region of India. *Advances in Biological Research*, 4(1), 23-26.
- Raiesi, S., Ojagh, S.M., Sharifi-Rad, M., Sharifi-Rad, J. and Quek, S.Y. (2017). Evaluation of *Allium paradoxum* (M.B.) G. Don. and *Eryngium caucasicum* Trautv. Extracts on the shelf-life and quality of silver carp (*Hypophthalmichthys molitrix*) fillets during refrigerated storage. *Journal of Food Safety*, 37(3), 12321. <https://doi.org/10.1111/jfs.12321>
- Rahman, M.S., Rasul, M.G., Hossain, M.M., Uddin, W., Majumdar, B.C., Sarkar, M.S.I. and Bapary, M.A.J. (2017). Impact of spice treatments on the quality and shelf life of sun dried taki (*Channa punctatus*). *Journal of Chemical, Biological and Physical Sciences*, 7(72), 409-420.
- Reddy, A. and Grace, J.R. (2016). In vitro evaluation of antioxidant activity of methanolic extracts of selected mangrove plants. *Medicinal and Aromatic Plants*, 5(3), 250-255. <https://doi.org/10.4172/2167-0412.1000250>
- Sanusi, S.B., Bakar, M.F.A., Mohamed, M., Sabran, S.F. and Mainasara, M.M. (2017). Ethnobotanical, phytochemical, and pharmacological properties of *Nepenthes* species: A review. *Asian Journal of Pharmaceutical and Clinical Research*, 10(11), 16-19. <https://doi.org/10.22159/ajpcr.2017.v10i11.20050>
- Schwallier, R., de Boer, H.J., Visser, N., van Vugt, R.R. and Gravendeel, B. (2015). Traps as treats: A traditional sticky rice snack persisting in rapidly changing Asian kitchens. *Journal of Ethnobiology and Ethnomedicine*, 11, 24. <https://doi.org/10.1186/s13002-015-0010-x>
- Servet, A. and Hudayi, E. (2011). Chemical composition of European squid and effects of different frozen storage temperatures on oxidative stability and fatty acid composition. *Journal of Food Science and Technology*, 48(1), 83-89. <https://doi.org/10.1007/s13197-010-0139-5>
- Shil, D., Mohanty, J.P., Das, T., Bhuyan, N.R. and Uriah, T. and Mohamed-Saleem, T.S. (2010). Protective role of pitcher of *Nepenthes khasiana* Hook against dexamethazone induced hyperlipidemia and insulin resistance in rat. *International Journal Research Pharmaceutical Science*, 1(2), 195-198.
- Shuaibu, B.S., Mohd, F.A.B., Maryati, M., Siti, F.S. and Muhammad, M.M. (2017). Ethnobotanical, phytochemical, and pharmacological properties of *Nepenthes* species: A review. *Asian Journal of Pharmaceutical and Clinical Research*, 10(11), 16-19. <https://doi.org/10.22159/ajpcr.2017.v10i11.20050>
- Smaou, S., Hlima, H.B., Mtibaa, A.C., Fourati, M., Sellem, I., Elhadeif, K., Ennouri, K. and Mellouli, L. (2019). Pomegranate peel as phenolic compounds source: Advanced analytical strategies and practical use in meat products. *Meat Science*, 158(4), 107914.
- Song, H.S. (2019). Antioxidant effects of ethanol extracts from plants on semi-dried eels. *Korean Journal of Food Preservation*, 26(1), 109-114. <https://doi.org/10.11002/kjfp.2019.26.1.109>
- Song, H.S. and Kim, Y.M. (2018). Antioxidant effects of ethanol extracts from plants on peroxide content in semi-dried eels. *The Korean Journal of Food and Nutrition*, 31(5), 647-652. <https://doi.org/10.11002/kjfp.2019.26.1.109>
- Sood, V., Tian, W., Narvaez-Bravo, C., Arntfield, S.D. and Gonzalez, A.R. (2020). Plant extracts effectiveness to extend bison meat shelf life. *Journal of Food Science*, 85(4), 936-946. <https://doi.org/10.1111/1750-3841.15062>
- Stina, C.M.B., Anders, E., Uko, B., Tonu, P., Martin, J., Jarkko, H., Sari, M., Risto, K., Pirjo, H.M., Vitalijs, R., Dalija, S., Asa, H., Kimmo, R. and Eva, T. (2020). Lipid oxidation inhibition capacity of plant extracts and powders in a processed meat model system. *Meat Science*, 162(2), 108033. <https://doi.org/10.1016/j.meatsci.2019.108033>
- Thao, N.P., Luyen, B.T.T., Koo, J.E., Kim, S., Koh, Y.S., Thanh, N.V., Cuong, N.X., Kiem, P.V., Minh, C.V. and Kim, Y.H. (2016). In vitro anti-inflammatory components isolated from the carnivorous plant *Nepenthes mirabilis* (Lour.) Rafarin. *Pharmaceutical Biology*, 54(4), 588-594. <https://doi.org/10.3109/13880209.2015.1067234>
- Tiewlasubon, U., Mrityunjaya, B.P. and Sivaiah, K. (2015). In vitro antioxidant and hepatoprotective potential of *Nepenthes khasiana* Hook. f against ethanol-induced. *Journal of Pharmaceutical Research*, 14(4), 81-89. <https://doi.org/10.18579/jpcrkc/2015/14/4/89467>
- Van Thanh, N., Thao, N.P., Huong, P.T., Lee, S.H., Jang, H.D. and Cuong, N.X. (2015). Naphthoquinone and flavonoid constituents from the carnivorous plant *Nepenthes mirabilis* and their anti-osteoporotic and antioxidant activities. *Phytochemical Letters*, 11(1), 254-259. <https://doi.org/10.1016/j.phytol.2015.01.009>

- Vanessa, M.O. and Zenilda, L.C. (2013). Analytical methods to assess carbonyl compounds in foods and beverages. *Journal of the Brazilian Chemical Society*, 24(11), 1711-1718. <https://doi.org/10.5935/0103-5053.20130236>
- Wan-Nor-Adibah-Wan, Z., Loke, K.K., Zulkapli, M.M., Salleh, I.M., Goh, H.H. and Noor, N.M. (2016). RNA-seq analysis of *Nepenthes ampullaria*. *International Journal of Plant Science*, 164(1), 635-639. <https://doi.org/10.3389/fpls.2015.01229>
- Wang, L., Zhou, Q., Zheng, Y. and Xu, S. (2009). Composite structure and properties of the pitcher surface of the carnivorous plant *Nepenthes* and its influence on the insect attachment system. *Progress in Natural Science*, 19(12), 1657-1664. <https://doi.org/10.1016/j.pnsc.2009.09.005>