Effect of chemical agents, metallic salts on the stability of α-amylase, protease and comparative analyses of enzyme activity of selected salad vegetables

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Abstract

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The present investigation was conducted to analyze the activity of enzymes (protease, α amylase, cellulase and urease) of selected salad vegetables (white radish, red radish, beet, carrot, papaya, cucumber and tomato) as well as to determine the effect of chemical agents and metallic salts on the stability of enzymes. Salad vegetables are one of the cheap sources of adequate vitamins, minerals and enzymes. The protease, a-amylase and cellulase activity were highest in papaya (4.11 ± 0.21 U/g, 1.68 ± 0.21 U/g and 0.26 ± 0.13 U/ g) whereas the urease activity was not detected in papaya, cucumber and tomato. The protease, α -amylase and cellulase activity of papaya were increased 24.83%, 42.26% and 57.69% than cucumber while the cellulase activity of carrot was increased 44.45% than tomato. The activity of protease and α -amylase of beet were decreased 52.44%, 33.63% while the cellulase activity of beet was increased 36.36% from radish. The toxicity indicating urease enzyme activity was not detected in papaya, tomato and cucumber but negligible in radish, beet and carrot which can be nullified. The activities of enzymes were increased in the presence of metallic salts such as Ca²⁺, Mg²⁺ and Mn²⁺ while Fe²⁺, Zn²⁺ and Cu²⁺ inhibited the enzyme's activity moderately. Results revealed that in the presence of higher concentrations of urea, EDTA and acetic acid, the activities of all the enzymes were completely inhibited.

1. Introduction

Salad vegetables are an essential part of people's diet all around the world. They are usually consumed as raw or cooking. Vegetable salad is a very common food accompaniment. The vegetables that usually make up these salad items include carrot, cucumber, tomato, papaya, beet and radish (Itohan et al., 2011). Carrot, cucumber, tomato, papaya, beet and radish are a versatile vegetable that may be consumed raw, cooked or in combination with other vegetables and meats in salads, freshly prepared foods and frozen prepared foods. Fruits and vegetables have health benefits for consumers, due to their content of fiber, vitamins and antioxidant compounds. However, for the antioxidant compounds, many changes occur during harvesting, preparation (fresh-cut fruits) and storage of these fruits. These changes induce a pronounced loss of microbiological and antioxidant qualities (Lindley, 1998). Fresh fruits and vegetables provide mankind with an abundance of benefits. These play an important role in health and have the ability to prevent many diseases such as heart

disease, cancers and diabetes (Liu, 2004; Pandey, *et al.*, 2006). The highly soluble minerals like calcium, phosphorus, iron, magnesium, copper, and potassium contained in vegetables maintain the acid/base balance of hydrogen concentration of body tissues. These help to completely absorb the vitamins, proteins, fats and carbohydrates of the food (Odoh and Okoro, 2012).

The World Health Organization (WHO) (1989) recommends the consumption of at least five 100 g portions of fruits and vegetables daily to stay healthy. Health nutrition specialists recommend adding to everyday diet as much as possible fruits and vegetables although the general public has a limited understanding of what the real benefits are, or how much should be eaten. Current recommendations are that adults should consume at least 400 g (or five servings a day) of fruits and vegetables (WHO, 1989). The nutritional analysis of edible fruit and vegetables plays a crucial role in the assessment of nutritional significance. A comprehensive survey was conducted by an extensive tour throughout the selected locations during both winter and summer FULL PAPER

seasons from April 2010 to December 2012. In the selected locations two or more vegetables are grown every year (Khan *et al.*, 2010). Vegetables are richer in mineral substances as compared with fruits. Enzymes enter into a vast number of biochemical reactions in fruits and vegetables, they may be responsible for changes in flavor, color, texture and nutritional properties. In fruit and vegetable storage and processing, the most important roles are played by the enzymes classes of hydrolyses (invertase, protease, amylase, cellulase) (Enacheshu, 1995). Salad vegetables are one of the cheap sources of adequate vitamins, minerals and enzymes.

Data available on the biochemical parameters of radish produced in Bangladesh are quite scanty. Only limited work has been done on the physiochemical properties of tomato, papaya, cucumber and carrot. Some works have been going on at Bangladesh Agricultural Research Institute (BARI) with emphasis on the higher production and development of new varieties and control of the diseases. No detail work has been done on the chemical composition and enzyme activities in tomato, papaya, cucumber and carrot. Enzymes have become a powerful catalytic tool in a wide variety of chemical processes. Higher plants play an important role as a source of many useful enzymes. Extensive use of enzyme activity determination as an aid to the diagnosis in hospital and laboratories all over the world is one of the dramatic developments in modern medicine (Jahan et al., 2011). The present study is, therefore, designed to study the activities of different enzymes and also to determine the chemical agents and metallic salts on the stability of the enzymes of selected salad vegetables.

2. Materials and methods

2.1 Collection of samples

Samples were collected directly from the cultivated field of the local area in the winter season.

2.2 Enzyme activity assay

Fresh healthy samples were washed thoroughly with distilled water, cut into small pieces and peste in a mortar at 4°C with cold 0.1M phosphate buffer of respective pH, for amylase 6.7, for protease citrate buffer, 5.5, for urease 0.2M phosphate buffer, 7.0 for cellulase 0.2M sodium acetate buffer, 5.2. The extracts were filtered with few layers of cheesecloth and further clarified by centrifugation at 10,000 rpm for 15 mins at 4°C. The clear supernatant was collected and used as a crude enzyme extract.

2.3 Assay of α -amylase activity

Amylase activity was assayed following the method as described by Jayaraman (1981). A total of 1% starch solution was used as the substrate (1.0 g in 100 mL of 0.1M phosphate buffer, pH 6.7). The amylase activity was measured by estimating the release of maltose. The amount of maltose released was calculated from the standard curve prepared with maltose. One unit of amylase activity was defined as the amount required for liberating 1 mg of maltose in 15 mins at 37°C.

2.4 Assay of protease activity

Protease activity was assayed following the modified method as described by Mahadevan and Sridhar (1982). Haemoglobin was used as substrate. The protease activity was measured by estimating the amount of leucine released from haemoglobin. The amount of leucine released was calculated from the standard curve prepared with leucine. One unit of protease activity was defined as the amount required for liberating 1 mg of leucine in 30 mins at 37°C.

2.5 Assay of cellulase activity

Cellulase activity was measured following the method described by Mahadevan and Sridhar (1982). Carboxymethyl cellulose (CMC) was used as substrate. Cellulase activity was measured by estimating the reducing sugar released from the substrate using the DNS method. One unit of cellulase activity was defined as the amount of enzyme required for liberating 1 mg of reducing sugar from CMC per minute at 37°C.

2.6 Assay of urease activity

Urease activity was assayed following the method as described by Jayaraman (1981). A total of 3% urea solution was used as substrate (1 g in 100 mL of 0.2M phosphate buffer, pH 7.0). The Urease activity was measured by estimating the release of ammonium. The amount of ammonium released was calculated from the standard curve prepared with ammonium. One unit of Urease activity was defined as the amount required for liberating 1 μ g of ammonium in 15 mins at 55°C.

2.7 Effect of chemical agents and metallic salts

Treatment with $CaCl_2$, urea, acetic acid, EDTA and various metallic salts at different concentrations were added to 5.0 mL of amylase, protease, cellulase and urease enzymes extract solutions from salad vegetables and incubated for 10 mins at 20°C. The mixtures were again incubated with the substrate for 15 mins at 37°C and the enzyme activities were assayed (Rahman, 2001).

2.8 Statistical analysis

Each experiment was replicated three times.

Reported data represented as mean values \pm SD obtained from three individual experiments. Data were subjected to analysis of variance using the Microsoft Excel Program (Redmond, Washington DC, USA).

3. Results and discussion

3.1 Enzymes activity

Vegetables are very rich sources of essential biochemicals and nutrients such as vitamins, minerals and enzymes which are the major portion of the human diet. The results of protease and α -amylase activity are shown in Table 1. The protease activity of white radish, red radish, beet, carrot, papaya, cucumber and tomato were 1.99±0.20 (U/g), 2.44±1.00 (U/g), 1.16±0.16 (U/g), 2.55±0.19 (U/g), 4.11±0.20 (U/g), 1.18±0.21 (U/g) and 2.85±0.22 (U/g) respectively (Table 1). The maximum protease activity was found in papaya 4.11±0.20 (U/g) and minimum in beet 1.16±0.16 (U/g). The protease activity of papaya 0.72-3.11 (U/g) which was slightly lower than the investigation (Xiang-dong *et al.*, 2006).

The papaya showed maximum α -amylase activity than beet. Results of the present study are in accordance with the findings with some researchers in the case of vegetables (Ajayi *et al.*, 2011). The cellulase activity of white radish, red radish, beet, carrot, papaya, cucumber and tomato were 0.14±0.07 (U/g), 0.072±0.012 (U/g), 0.22±0.11 (U/g), 0.18±0.07 (U/g), 0.26±0.06 (U/g), 0.11±0.03 (U/g), and 0.10±0.05 (U/g) respectively (Table 1). Salad vegetables have a poor amount of protein and carbohydrate contents. That's why the enzyme protease and α -amylase show fewer assay results.

The highest cellulase activity was found in papaya $(0.26\pm0.06 \text{ U/g})$ and the lowest was in red radish $(0.072\pm0.012 \text{ U/g})$. Tomato showed lowest (9.09%) cellulase activity than cucumber among the salad variety. These findings are in close agreement with the researchers who revealed the low cellulase activity in tomato (Wilson, 2012). Besides, the highest cellulase activity was 57.69\% in papaya than cucumber. The

urease activity of white radish, red radish, beet, carrot, papaya, cucumber and tomato were 0.067 ± 0.10 (U/g), 0.061 ± 0.25 (U/g), 0.081 ± 0.10 (U/g), 0.018 ± 0.09 (U/g) respectively (Table 1). The urease activity of papaya, cucumber and tomato were not detected. The α -amylase activity of carrot was 0.18 ± 0.15 (U/g). The cause may be the presence of urease activity in carrot the utilization of nitrogen fertilizer in cultivating soil. Besides this, the results showed no presence of urease activity in carrot in his research work (Richard and Michael, 1973). Urea was found naturally in the environment and is also artificially introduced, comprising more than half of all synthetic nitrogen fertilizers used globally. For this reason, the negligible amount of urease activity was found in radish, beet and carrot (Nowak *et al.*, 2003).

3.2 Effect of calcium on the activities of α -amylase and protease

The effect of calcium as metallic salt on the activities of α -amylase and protease is presented in Table 2. The activities of enzymes were gradually increased with the increasing concentration of calcium and in the presence of 0.50 M Ca²⁺, the activities of α -amylase and protease became 135.24% and 129.46% respectively.

Table 2. Effect of calcium on the activities of α -amylase and protease

Concentration of CaCl ₂ (Molar)	Relative activities (%)	
	α-amylase	Protease
0.000	100.00	100.00
0.001	104.36	105.66
0.005	107.18	108.34
0.010	113.35	112.68
0.050	117.38	116.38
0.300	130.24	123.65
0.500	135.34	129.46

3.3 Activities of the enzymes after treatment with different concentration of urea

Table 3 shows the relatives activities of the enzymes after treatment with different concentration of urea. The activities of the α -amylase and protease were found to be decreased remarkably in the presence of urea

Table 1. Protease, α -amylase, cellulase and urease activities of white radish, red radish, beet, carrot, papaya, cucumber and tomato.

Name of samples	Enzymes activity			
Name of samples	Protease (U/g)	α-amylase (U/g)	Cellulase (U/g)	Urease (U/g)
White Radish	$1.99{\pm}0.20$	1.47 ± 0.20	$0.14{\pm}0.07$	0.067 ± 0.10
Red Radish	$2.44{\pm}1.00$	1.10 ± 0.21	0.072 ± 0.012	0.061 ± 0.25
Beet	1.16 ± 0.16	0.73 ± 0.10	0.22 ± 0.11	0.081 ± 0.10
Carrot	2.55±0.19	1.56 ± 0.18	0.18 ± 0.07	0.018 ± 0.09
Papaya	4.11 ± 0.20	1.68 ± 0.23	0.26 ± 0.06	Not detected
Cucumber	1.18 ± 0.21	0.97 ± 0.20	0.11±0.03	Not detected
Tomato	2.85 ± 0.22	1.52 ± 0.22	$0.10{\pm}0.05$	Not detected

The values are mean \pm SD of determinations made in triplicate

and at 6M urea concentration. Protease and α -amylase decreased by about 88% and 79% respectively.

Table 3. Effect of urea on the activities of α -amylase and protease

Concentration of urea (Molar)	Relative activities (%)	
	α-amylase	Protease
0	100.00	100.00
1	95.17	91.62
2	82.41	75.78
4	46.82	41.76
6	22.64	13.35
8	0.00	0.00

3.4 Activities of α -amylase and protease after treatment with different concentration of EDTA

The relative activities of α -amylase and protease in the presence of different concentrations of EDTA are presented in Table 4. The activities of α -amylase and protease are abolished gradually with the increase of EDTA concentrations and the enzymes lost their activities completely in the presence of 0.50M EDTA.

Table 4. Effect of EDTA on the activities of α -amylase and protease

Relative activities (%)	
α-amylase	Protease
100.00	100.00
85.34	76.69
67.29	59.82
53.22	47.34
34.46	27.64
15.60	13.36
0.00	0.00
	α-amylase 100.00 85.34 67.29 53.22 34.46 15.60

3.5 Activities of a-amylase and protease after treatment – with different concentration of acetic acid

As shown in Table 5, the activities of the enzymes were gradually decreased with the treatment of higher percentage of acetic acid. It is found that α -amylase and protease retained 22.43% and 19.17% after treatment with 20% acetic acid. Overall, the effect of calcium, urea, EDTA and acetic decrease the enzyme's activity which is partially in agreement with Akand (2003) and Haque *et al.* (2017)

Table 5. Effect of acetic acid on the activities of α -amylase and protease

Concentration of acetic acid (%)	Relative activities (%)	
	α-amylase	Protease
0.00	100.00	100.00
2.50	90.16	89.53
5.00	75.33	75.89
10.00	46.65	47.20
20.00	22.43	19.17
30.00	0.00	0.00

3.6 Effect of metallic salts on the activities of α -amylase and protease

Table 6 represents the effect of various metallic salts on the activities of α -amylase and protease. The activities of enzymes were increased remarkably in the presence of Mn^{2+} salts while that was increased slightly in the presence of Mg^{2+} salts. Other metallic salts such as K^{2+} and Na^{2+} produce little or no inhibitory on the activities of the enzymes but the activities of all the enzymes reduce significantly in the presence of Zn^{2+} , Cu^{2+} and Fe^{2+} .

Table 6. Effect of various metallic salts on the activities of α -amylase and protease.

T (C 1)	Concentration (Molar)	Relative activities (%)	
Test Salts		α-amylase	Protease
None	-	100.00	100.00
	0.001	105.72	106.01
MgCl ₂	0.002	108.32	109.12
	0.001	85.24	93.26
$ZnCl_2$	0.002	73.21	86.55
	0.001	86.71	71.84
CuCl ₂	0.002	76.36	53.64
	0.001	117.73	113.42
MnCl ₂	0.002	127.63	121.66
	0.001	100.00	99.72
NaCl ₂	0.002	99.78	99.98
	0.001	100.00	100.00
KCl ₂	0.002	100.00	100.00
	0.001	73.16	67.28
FeCl ₂	0.002	589.44	53.20

The activity of α -amylase and protease were abolished completely by denaturing agents 8.0M urea. These denaturing agents may destroy the secondary or tertiary structure of the enzymes this results in complete destruction of the active enzyme at higher concentration (Rahman, 2001; Haque *et al.*, 2017).

The activities of the enzymes increased significantly in the presence of divalent cation Ca^{2+} and Mn^{2+} suggesting the involvement of these divalent ions in maintaining the active conformation of the enzymes. Other metallic salts such as K^{2+} and Na^{2+} produce little or no inhibitory on the activities of the enzymes but the activities of all the enzymes reduce significantly in the presence of Zn^{2+} , Cu^{2+} and Fe^{2+} (Akand, 2003; Haque *et al.*, 2017).

4. Conclusion

It was aimed to evaluate the enzymatic activity (α -amylase, urease, cellulase and protease) of white radish,

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red radish, beet, carrot, papaya, cucumber and tomato with each other. It was found that the enzymes' activity was different in all the selected salad vegetables its play an important role in the metabolic function of the human body. Through this research work, the enzymatic activity of salad vegetables was determined. Moreover, the observations were shown that higher concentrations of chemical agents and metallic salts inhibit enzymes activity in a maximum potential level.

Conflict of Interest

The authors have no conflict of interest.

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