

Health promoting properties of *Alternanthera brasiliana* leaves and *Hibiscus sabdariffa* calyces used in fortification of maize-bambara groundnut malt and maize-cowpea malt complementary foods

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Abstract

The study evaluated the chemical and antioxidant properties of *Alternanthera brasiliana* leaves and *Hibiscus sabdariffa* calyces used in iron and zinc fortification of maize-bambara groundnut malt and maize-cowpea malt complementary foods. *A. brasiliana* leaves and *H. sabdariffa* calyces were freshly harvested, dried at 50°C for 48 hours and analyzed for the relevant chemical components and antioxidant activities. The vitamin A content was 6996 and 745.6 µgRE/kg while the vitamin C was 238.26 and 294.78 mg/kg respectively. The aqueous extracts of *A. brasiliana* and *H. sabdariffa* calyces contained 509.5 mg/kg and 5234.72 mg/kg of alkaloids, 1545 mg/kg and 384 mg/kg of anthocyanins, 767.3 and 235.83 mg/kg of carotenoids, 14,702.8 and 26,428.3 mg/kg of phenols, 1043.5 and 897.63 mg/kg steroids and 462.0 mg/kg and 1006.5 mg/kg of flavonoids respectively. *A. brasiliana* and *H. sabdariffa* extracts had concentration-dependent DPPH activity with IC₅₀ of 1.76 mg/ml and 5.745 mg/ml, nitric oxide scavenging activity with IC₅₀ of 0.675 mg/ml and 3.976 mg/ml while the ferric reducing power had an absorbance range of 0.5 – 0.982 and 0.959 – 0.986 respectively. The study revealed that *A. brasiliana* leaves and *H. sabdariffa* calyces contain components that will impact positively on the health of the infants when used to formulate complementary foods.

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1. Introduction

The role of diet and nutrition as determinants of chronic disease is well documented (Middleton *et al.*, 2000; Yi-fang, 2002) and there is growing evidence that chemical components of plants and microbial foods may play an integral role in the link between food and long-term health (Ferrari, 2004). Complementary foods are any nutrient containing food given to young children along with breast-milk (Gibbs, 2010) when the breast milk nutrients become inadequate for their energy and growth needs (WHO, 2003) within the 6 - 23 months window. It is required to ensure adequate growth, to prevent malnutrition, stunting, and anaemia (Bhasin *et al.*, 2003). Poor complementary feeding is the immediate direct cause of malnutrition [which manifests as protein energy malnutrition (PEM) and micronutrient

deficiencies] leading to growth faltering and high rates of infections during infancy.

In most traditional cultures of the developing world, cereal-based gruels are the first complementary foods to be introduced to infants, sometime between 4 and 6 months of age. They are followed by vegetables, fruits, fruit juices, and meat products. These plant-based traditional complementary foods do not meet these nutritional requirements because they are deficient in micronutrients such as calcium, iron, zinc and vitamin A (Dewey and Brown, 2003). This is because the level of these micronutrients falls below the Recommended Dietary Allowance (RDA) of 50 g/kg for calcium, 275 mg/kg for iron, 125 mg/kg for zinc and 5000 µgRE/kg for vitamin A. Hence, calcium, iron, zinc and vitamin A deficiencies continue to pose great danger to 6-12 months old infants leading to increased susceptibility to

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infections such as diarrhea, cholera and impaired immunity (Lutter and Dewey, 2003).

In infants, iron and zinc requirements are difficult to meet from non-fortified complementary foods (WHO/UNICEF, 2003). These can be prevented by fortification of infant foods. Fortification of plant-based complementary foods with vitamin and mineral pre-mix or animal supplements such as milk makes the foods expensive for low-income earners who earn less than 300 naira (1.5 USD) per day (Anon., 2012). The alternative is to adopt food-to-food fortification. This has fueled the interest in plant foods as sources of micronutrients for combating micronutrient deficiency of chemicals that may have useful roles in the health of infants. The suitability of a complementary food depends on a number of factors including the nature of raw materials and methods of processing and fortification practices adopted.

Alternanthera brasiliana (L) O. Kuntze belongs to the Family Amaranthaceae commonly known in Brazil as joy weed or Josephs coat is widely used as a medicinal agent to cure different disease, such as inflammation, wound healing, analgesic, antitumor activity, immune modulator and lymphocyte proliferation (Duarte and Debur, 2004; Saawan et al., 2011). Roselle (*Hibiscus sabdariffa* L.) on the other hand belongs to the Family Malvaceae and is known in Nigeria as “Zobo”. It is used effectively in folk medicines for the treatment of hypertension, inflammatory diseases and cancer (Kong et al., 2003; Lin et al., 2007), decrease blood viscosity and reduce hypertension (Christian et al., 2006).

Apart from these properties, there is evidence that the calyces of *Hibiscus sabdariffa* can be used in food-to-food fortification to boost the iron and zinc contents in maize-bambara groundnut malt and maize-cowpea malt complementary foods (Attaugwu, 2015). It is against this background that the health promoting properties of *Hibiscus sabdariffa* and *Alternanthera brasiliana* which is known to have good iron and zinc contents were investigated.

2. Materials and methods

2.1 Materials

Fresh leaves of Brazilian joy weed [*Alternanthera brasiliana* (L.) O. Kuntze] and calyces of Roselle [*Hibiscus sabdariffa* (L.) malvaceae] was obtained from

a farm in the University of Nigeria Nsukka.

2.2 Methods

One thousand grams (1000 g) of freshly harvested *A. brasiliana* leaves and *H. sabdariffa* calyces were chopped into 2 mm slices and dried at 50°C in a convection Gallenkamp oven (Model IH-150, Gallenkamp, London, England). The extracts for the free radical scavenging activities were prepared according to the method described by Oyedemi et al. (2010) with a slight modification of using ethanol for extraction instead of methanol.

2.3 Analysis

The steroid content, total carotenoid, ascorbic acid, thiamine, riboflavin, niacinamide, pyridoxine hydrochloride and cobalamin contents were determined using the method as described in AOAC (2010). The vitamin E content of the samples was determined by the α,α -dipyridyl method of Pearson (1976). The vitamin K content of the samples was determined by the 2, 4-dinitrophenylhydrazine method of Snell and Snell (1953). The flavonoid and alkaloid contents were determined using the method of Harborne (1973). The total phenolic content in the aqueous extracts (1:4 w/v) of dried *A. brasiliana* leaves and dried *H. sabdariffa* calyces were determined using the method of Wolfe et al. (2003). Total flavonoids were determined by the method of Ordoñez et al. (2006). Total flavonol content was determined by the spectrophotometric procedure described by Kumaran and Karunakaran (2007). Total proanthocyanidin determination was based on the spectrophotometric procedure of Sun et al. (1998). The DPPH and nitric oxide free radical scavenging activities were determined by the methods of Liyana-Pathiranan and Shahidi (2005) and Oyedemi et al. (2010) respectively. The ferric oxide reducing power of the ethanolic extract was evaluated according to the spectrophotometric method of Yen and Chen (1995).

3. Results and discussion

3.1 Vitamin composition of *A. brasiliana* leaves and *H. sabdariffa* calyces

The vitamin composition of *A. brasiliana* leaves and *H. sabdariffa* calyces are presented in Table 1. The vitamin A content of *A. brasiliana* and *H. sabdariffa* calyces were 6996 $\mu\text{gRE/kg}$ and 745.6 $\mu\text{gRE/kg}$ respectively. The vitamin A content of *A. brasiliana* leaves and *H. sabdariffa* calyces are above the RDA

requirement of 500 $\mu\text{gRE/kg}$ in complementary foods (Lutter and Dewey, 2003) implying that they could be a good source of vitamin A which is needed for good vision in infants. Hence the use of *A. brasiliana* leaves and *H. sabdariffa* calyces in complementary foods could improve the vision of infants taking the food. Saawan *et al.* (2011) reported 19260 $\mu\text{g/kg}$ pro-vitamin A carotenoids in *A. brasiliana*.

Table 1. Vitamin composition of *A. brasiliana* leaves and *H. sabdariffa* calyces

Micronutrient	<i>A. brasiliana</i>	<i>H. sabdariffa</i>
Vitamin A ($\mu\text{gRE/kg}$)	6996.00 \pm 0.0167 ^b	745.60 \pm 0.0042 ^a
B ₁ (mg/kg)	11.12 \pm 0.0551 ^a	12.37 \pm 0.0045 ^b
B ₂ (mg/kg)	654.10 \pm 0.0318 ^a	4094.50 \pm 0.0271 ^b
B ₃ (mg/kg)	4184.40 \pm 0.0242 ^b	2163.10 \pm .0324 ^a
B ₆ (mg/kg)	21.01 \pm .0009 ^a	8.32 \pm 0.0042 ^b
B ₁₂ (mg/kg)	1.44 \pm .0015 ^a	43.80 \pm 0.0057 ^b
C (mg/kg)	238.36 \pm 0.0159 ^a	294.78 \pm 0.0411 ^b

Results are the means of three replications. Values carrying different superscript in the same row are significantly different ($p < 0.05$)

The vitamin B₁ (thiamin) content of *A. brasiliana* and *H. sabdariffa* were 11.12 mg/kg and 12.37 mg/kg respectively. The vitamin B₁ content of *H. sabdariffa* of this report is higher than the 1.23 and 1.77 mg/kg reported by Luvonga *et al.* (2012) for fresh and dried *H. sabdariffa* respectively. The differences may be attributed to genetic variety and type of soil. These results implied that infants consuming these foods will have an improved appetite, a healthier nervous system and a higher release of energy from the complementary foods as these are the major functions of vitamin B₁ (thiamin) in the body.

The vitamin B₂ (riboflavin) content of *A. brasiliana* and *H. sabdariffa* were 654 mg/kg and 4094.50 mg/kg respectively. The vitamin B₂ content of *A. brasiliana* was higher than the 140 mg/kg content reported by Saawan *et al.* (2011). The differences may be attributed to genetic variety and type of soil. Vitamin B₂ (riboflavin) is responsible for maintaining healthy blood cells, plays an important role in the conversion of food into energy and in fat and protein metabolism. Hence, infants consuming complementary foods containing *A. brasiliana* and *H. sabdariffa* will have healthy blood

cells, healthier vision and skin as well as improved fat and protein metabolism.

The vitamin B₃ (niacin) content of *A. brasiliana* leaves and *H. sabdariffa* calyces were 4184.40 mg/kg and 1216.31 mg/kg respectively. The vitamin B₃ content of *A. brasiliana* in this report was lower than the 12,000 mg/kg reported by Saawan *et al.* (2011) for *A. brasiliana*. The differences may be attributed to genetic variety and type of soil. However, the vitamin B₃ content of *H. sabdariffa* (1216.31 mg/kg) of this report was higher than the 37.7 mg/kg reported by Wong *et al.* (2002) for *H. sabdariffa*. The difference in the vitamin B₃ content of the present report could be attributed to genetic variety and geographic differences. Vitamin B₃ helps in lowering the level of bad cholesterol and elevation of good (HDL) cholesterol level leading to significant decrease in heart disease. It also offers protection for certain skin cancers.

The vitamin B₆ content of *A. brasiliana* leaves and *H. sabdariffa* calyces were 21.01 mg/kg and 8.32 mg/kg respectively. The high vitamin B₆ content of *A. brasiliana* and *H. sabdariffa* implies that the use of *A. brasiliana* leaves and *H. sabdariffa* calyces in complementary foods will help in fighting anemia in children if consumed since the health benefits of vitamin B₆ includes stimulating co-enzymatic activities, positive effect on hormone control, cardiac diseases, kidney disorder and anemia.

The vitamin C content of *A. brasiliana* leaves and *H. sabdariffa* calyces were 238.36 mg/kg and 294.78 mg/kg respectively. The vitamin C content of *A. brasiliana* (238.36mg/kg) was higher than the 170 mg/kg content reported by Saawan *et al.* (2011) while that of *H. sabdariffa* (294.78 mg/kg) was higher than 67.01 and 49.90 mg/kg reported by Luvonga *et al.* (2012) for fresh and dried *H. sabdariffa*. The difference could be attributed to genetic variety and type of soil. The high vitamin C content of the samples could enhance non-heme iron absorption from the fortified complementary foods hence reduce the prevalence of anemia in the infants consuming it. It will also help in fighting some cancers as vitamin C is a known antioxidant.

3.2 Phytochemical composition of *A. brasiliana* and *H. sabdariffa* calyces

The phytochemical composition of aqueous extracts of *A. brasiliana* leaves and *H. sabdariffa* calyces are presented in Table 2. The alkaloid content of aqueous extracts of *A. brasiliana* leaves and *H. sabdariffa*

calyces were 509.5 mg/kg and 5234.72 mg/kg respectively. The anthocyanin content of *A. brasiliana* leaves and *H. sabdariffa* calyces aqueous extracts were 1545 mg/kg and 384 mg/kg respectively. The phenolic contents of aqueous extracts of *A. brasiliana* leaves and *H. sabdariffa* calyces were 14,702.80 mg/kg and 26,428.30 mg/kg respectively. The flavonoid content of extracts of *A. brasiliana* leaves and *H. sabdariffa* calyces were 462.00 mg/kg and 1006.50 mg/kg respectively. The carotenoid contents of the plant extracts were 767.30 mg/kg and 235.83 mg/kg for *A. brasiliana* leaves and *H. sabdariffa* calyces respectively. Saawan *et al.* (2011) reported 19.26 mg/kg carotenoids in *A. brasiliana* leaves while Wong *et al.* (2002) reported 0.3 mg/kg carotenoid content in *H. sabdariffa* calyces. The steroid content of *A. brasiliana* leaves and *H. sabdariffa* calyces were 1043.50 mg/kg and 897.63 mg/kg respectively. *H. sabdariffa* calyces had 4725.2 mg/kg higher alkaloid content than *A. brasiliana* leaves. The anthocyanin content of *A. brasiliana* was 1161 mg/kg higher than that of *H. sabdariffa*. *A. brasiliana* leaves would have more antioxidant/ health promoting benefit than *H. sabdariffa* calyces. The phenolic content of *H. sabdariffa* was 11,725.50 mg/kg higher than that of *A. brasiliana*. This suggests that the *H. sabdariffa* extract will possess higher antioxidant activity than *A. brasiliana*. The flavonoid content of *H. sabdariffa* calyces was 544.50 mg/kg higher than that of *A. brasiliana* leaves. This could be attributed to the fact that flowers generally have more flavonoids which are the most important pigments for flower and petal coloration and are used mainly for the attraction of pollinator animals. The result of this study is similar to that of Elija *et al.* (2010) who reported higher flavonoid content in flowers of *Ipomoea carnea* than in the stem and leaves antioxidant effects of which are attributed to their redox properties. They are therefore reducing agents, hydrogen donors, singlet oxygen quenchers and metal- chelators (Vladimir-Knezevic *et al.*, 2011), properties which could contribute to their potential role in the prevention of cancer and heart disease.

3.3 Antioxidant properties

The antioxidant activities of ethanolic extracts of dried *Alternanthera brasiliana* leaves and *Hibiscus sabdariffa* calyces are illustrated in Figures 1, 2 and 3.

Proanthocyanidins, vitamin C and phenols content have been reported to have high antioxidant activity (Oyademi *et al.*, 2010; Azza *et al.*, 2011). The DPPH activity of *A. brasiliana* increased with increasing concentration of extract while that of *H. sabdariffa*

decreased with increasing concentration of extract. The latter could be due to exhaustion of DPPH reacting species at a lower concentration of extract or, due to the acidic pH of the extract (Luvonga *et al.*, 2012). A similar result of decrease has been reported by Luvonga *et al.* (2012).

Table 2. Phytochemical composition of *A. brasiliana* leaves and *H. sabdariffa* calyces

Results are the means of three replications. Values carrying

Phytochemical composition	<i>A. brasiliana</i> (mg/kg)	<i>H. sabdariffa</i> (mg/kg)
Alkaloids	509.50 ± 0.0531 ^a	52,347.20 ± 0.1089 ^b
Anthocyanin	1545.00 ± 0.0473 ^b	384.00 ± 0.0300 ^a
Carotenoids	767.30 ± 0.0410 ^b	235.803 ± 0.0111 ^a
Steroids	1043.50 ± 0.0234 ^b	897.63 ± 0.0762 ^a
Phenols	14,702.40 ± 0.0212 ^a	26,428.33 ± 0.0122 ^b
Flavonoids	462.00 ± 0.0891 ^a	1006.50 ± 0.0140 ^b
Total phenol (mg/kg gallic acid)	67,320.00 ± 0.0222 ^a	67,440.00 ± 0.0341 ^b
Total flavonoids (mg/kg quercetin equivalent)	69,030.00 ± 0.0541 ^b	57,810.00 ± 0.0751 ^a
Total flavonols (mg/kg quercetin equivalent)	50,430.00 ± 0.0323 ^b	8710.00 ± 0.0211 ^a
Proanthocyanidins (mg/kg catechin equivalent)	461,340.00 ± 0.0412 ^a	729,290.00 ± 0.0122 ^b

different superscripts in the same row are significantly different (p<0.05)

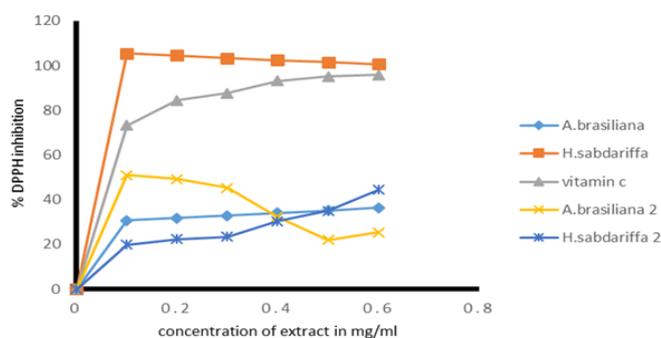


Figure 1. DPPH scavenging activity of *A. brasiliana* and *H. sabdariffa* extracts. *A. brasiliana* = *Alternanthera brasiliana* extract, *H. sabdariffa* = *Hibiscus sabdariffa* extract, Vitamin C = Vitamin C, *A. brasiliana* 2 = *Alternanthera brasiliana* extract adjusted with NaOH, *H. sabdariffa* 2 = *Hibiscus sabdariffa* extract adjusted with NaOH.

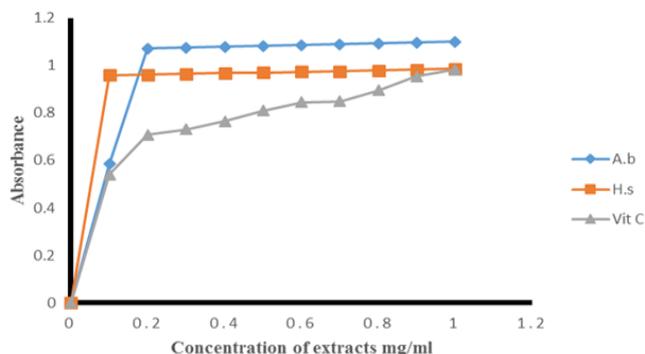


Figure 2. Ferric oxide reducing power of *A. brasiliana* and *H. sabdariffa* extracts. A.b = *Alternanthera brasiliana* extract, H.s = *Hibiscus sabdariffa* extract, Vit C = Vitamin C.

The abilities of the extracts to scavenge DPPH radicals (Figure 1) showed that *A. brasiliana* and *H. sabdariffa* extracts had DPPH radical scavenging activity with IC_{50} of 1.76 mg/ml and 5.745 mg/ml respectively. The DPPH activity of *A. brasiliana* and *H. sabdariffa* extracts were lower than the IC_{50} of 0.088 mg/ml for vitamin C, the reference antioxidant. Although the IC_{50} of *A. brasiliana* was higher than the IC_{50} of *H. sabdariffa*, which at a low concentration of 0.1 mg/ml inhibited 105.54% of DPPH radicals. This decreased gradually to 1.5 % at 10.05 mg/ml implying that *H. sabdariffa* possesses higher DPPH activity than *A. brasiliana* extract. The higher DPPH activity of *H. sabdariffa* could be attributed to its higher content of total phenol (67,440 mg/kg gallic acid equivalent), proanthocyanidins (729,290 mg/kg catechin equivalent) and vitamin C (294.78mg/kg) compared to the total phenol content of 67,340 mg/kg gallic acid, proanthocyanidins (461,340 mg/kg catechin equivalent) and vitamin C (238.26 mg/kg) in *A. brasiliana*.

The effect of pH of *H. sabdariffa* extract was evident when the different concentrations of extracts (0.1 -1.0 mg/ml) used for the assay were treated with 5M NaOH to neutralize the acidity of the extracts. The DPPH activity of the neutralized extract of *H. sabdariffa* increased with increasing concentration of the extract; the IC_{50} was 16.75 mg/ml, implying that the DPPH scavenging activity of *H. sabdariffa* decreased as pH increased from 2 to 7. In *A. brasiliana*, however, the DPPH scavenging activity of the extract adjusted with 5 M NaOH decreased with increasing concentration and had an IC_{50} of 0.229 mg/ml. *A. brasiliana* leaves extract showed a concentration-dependent DPPH scavenging activity while *H. sabdariffa* showed a pH dependent DPPH scavenging activity.

The ability of ethanolic extracts of *A. brasiliana* and *H. sabdariffa* to reduce Fe^{3+} to Fe^{2+} was high (Figure 2).

A. brasiliana extract had a wider absorbance range of 0.5841-1.1012 at 0.1 – 1 mg/ml than *H. sabdariffa* extract (0.959 - 0.986) and vitamin C (0.5 – 0.982). *A. brasiliana* extract with an absorbance range of 0.5841-1.1012 had higher ferric ion reducing power than *H. sabdariffa* (0.959 – 0.986) and vitamin c. This could be attributed to the higher total flavonoids (69,034 mg/kg quercetin equivalent) and flavonols (50,430 mg/kg quercetin equivalent) in *A. brasiliana* compared to the 57,810 mg/kg and 8,710 mg/kg quercetin equivalent for flavonoids and flavonols in *H. sabdariffa*. This indicated a higher ability of the *A. brasiliana* extract to reduce ferric ion than vitamin C which at 0.1 – 1 mg/ml had an absorbance range of 0.5 – 0.982. The ferric reducing power of the extracts was concentration dependent implying that they increased with increasing concentration of extracts.

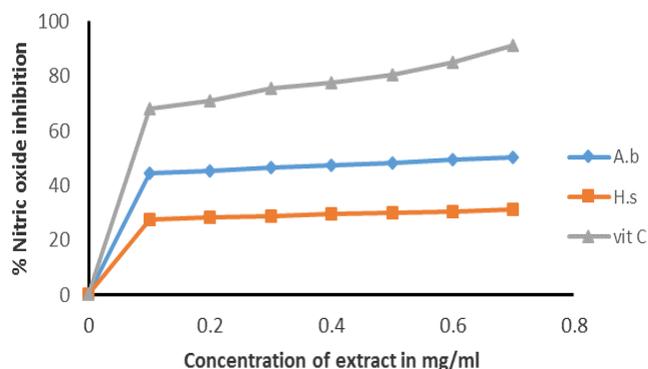


Figure 3. Nitric oxide scavenging activity of *A. brasiliana* and *H. sabdariffa* extracts. A. b = *Alternanthera brasiliana* extract, H. s = *Hibiscus sabdariffa* extract, Vit C = Vitamin C

The nitric oxide radical scavenging activity of *A. brasiliana* and *H. sabdariffa* extracts (Figure 3) were high and dose-dependent, with IC_{50} of 0.675 mg/ml and 3.976 mg/ml respectively. The nitric oxide scavenging activity of *A. brasiliana* and *H. sabdariffa* extract were lower than the IC_{50} (0.322 mg/ml) of vitamin C. *A. brasiliana* extract with IC_{50} of 0.675 mg/ml had a higher nitric oxide scavenging activity than *H. sabdariffa* calyces extract with IC_{50} of 3.976 mg/ml. This could be attributed to its higher total flavonoid content (69,034 mg/kg quercetin equivalent); higher flavonols (50,430 mg/kg quercetin equivalent) content compared to the total flavonoid (57,810 mg/kg quercetin equivalent) and total flavonols (8,710 mg/kg quercetin equivalent) of *H. sabdariffa*. Brachado *et al.* (2003) identified six flavonoids in *A. brasiliana* leaf extract, out of which 3 inhibited lymphocyte proliferations. The high nitric oxide scavenging activity of the extracts suggests that the extracts be used in preventing inflammation, carcinomas

and other diseases caused by nitric oxide radicals derived from cellular by-products as nitric oxide radical is known to play an important role in various inflammatory processes such as carcinomas, juvenile diabetes, multiple sclerosis, arthritis and ulcerative colitis (Hazra *et al.*, 2008).

4. Conclusion

The study has revealed that *A. brasiliana* leaves and *H. sabdariffa* calyces are good sources of vitamin B₁, B₂, B₃, and vitamin C as well as other health-promoting components such as flavonoids and phenols. The results implied that infants consuming these foods may have an improved appetite, a healthier nervous system and a higher release of energy from the complementary foods as well as be able to fight cancer better.

References

- Anon. (2012). Omega-3 fatty acids. Retrieved on August 15, 2012 from EMUMAGIC website: <http://emumagic.com/omega>.
- Association of Official Analytical Chemists (AOAC). (2010). Official Methods of Analysis. 18th ed. Washington, D.C.: AOAC.
- Attaugwu, R.N. (2015). Improving the chemical and functional properties of maize-bambara groundnut malt and maize-cowpea malt complementary foods by food-to-food fortification. Nsukka, Nigeria: University of Nigeria, PhD. Dissertation.
- Azza, A.A., Ferial, M.A. and Esmat, A.A. (2011). Physicochemical properties of natural pigments (anthocyanin) extracted from Roselle calyces (*Hibiscus sabdariffa*). *Journal of American Science*, 7(7), 445-456.
- Bhasin, R., Chatterjee, K. and Ramalingam, V. (2003). Blood transfusion transmitted diseases. In Saran, R. (Ed.). Transfusion medicine-technical manual (2), p. 143 – 174. New Delhi, India.
- Brachado, C., Almeida, A., Barreto, B. O., Costa, L. P., Ribeiro, L. S., Pereira, R. L., Gonclaves Koaz., V. L. and Costa, S. S. (2003). Flavonolrobinobiosides and rutosides from *Alternanthera brasiliana* (Amaranthaceae) and their effects on lymphocyte proliferation *in vitro*. *Journal of Brazilian Chemical Society*, 14(3), 1 - 5.
- Christian, K.R., Nair, M.G. and Jackson, J.C. (2006). Antioxidant and cyclooxygenase inhibitory activity of sorrel (*Hibiscus sabdariffa*). *Journal of Food Composition and Analysis*, 19, 778-783.
- Dewey, K.G. and Brown, K.H. (2003). Update on technical issues concerning complementary feeding of young children in developing countries and implications for intervention programs. *Food and Nutrition Bulletin*, 24(1), 5 - 28.
- Duarte, M.R. and Debur, M.C. (2004). Characteristics of the leaf and stem morpho-anatomy of *Alternanthera brasiliana* (L) O. Kuntze, Amaranthaceae. *Brazilian Journal of Pharmaceutical Sciences*, 40(1), 3-9.
- Elija K., Vaishali, B., Manik, M.K., Deshpande, N.R. and Kashaikar, R.V. (2010). Spectroscopic determination of total phenol and flavonoid contents of *Ipomoea carnea*. *International Journal of Chemical Technology Research*, 2(3), 1698-1701
- Ferrari, C. K. B. 2004. Functional Foods, Herbs and Nutraceutical towards Biochemical Mechanisms of Healthy aging. *Biogerontology*, 5, 275-289.
- Gibbs, M.M. (2010). Manufactured complementary foods for infant and young child feeding in Asia: micronutrient adequacy and improvement. Dunedin, New Zealand: University of Otago, MSc. Thesis.
- Harborne, J.B. (1973). Phytochemical Methods a Guide to Modern Techniques of plant Analysis, New York: Chapman and Hall.
- Hazra, B., Santana, B. and Nripendranath, M. (2008). Antioxidant and free radical scavenging activity of *Spondias pinnata*. *Journal of BMC Complementary and Alternative Medicine*, 8, 63.
- Kong, J. M., Chia, L. S., Goh, N. K., Chia, T. F. and Brouillard, R. (2003). Analysis and biological activities of anthocyanin. *Phytochemistry*, 64, 923-933.
- Kumaran, A. and Karunakaran, F.J. (2007). *In vitro* antioxidant activities of methanol extracts of phyllanthus species from India. *Life Technology*, 40, 344 - 352.
- Lin, T-L., Lin, H.H., Chen, C.C., Lin, M.C., Chou, M.C. and Wang, C.J. (2007). *Hibiscus sabdariffa* extract reduces serum cholesterol in men and women. *Nutrition Research*, 27, 140-145.
- Liyana-Pathiranan, C.M. and Shahidi, F. (2005). Antioxidant activity of commercial soft and hard wheat (*Triticumaestivum*.L.) as affected by gastric pH conditions. *Journal of Agriculture and Food Chemistry*, 53, 2433 – 2440.
- Lutter, C.K. and Dewey, K.G. (2003). Proposed nutrient composition for fortified complementary foods. *The Journal of Nutrition* 133(9), 3011s - 3020s.
- Luvonga, W.A., Njoroge, M.S., Makokha. A. and Ngunjiri, P.W. (2012). Chemical characterization of *Hibiscus sabdariffa* (Roselle) calyces and evaluation of its functional potential in the food industry.

- Retrieved on September 12, 2016 from JKUAT Website: [Journal. Jkuat.ac.ke/index.php/jscp/article/download/745/687](http://Journal.Jkuat.ac.ke/index.php/jscp/article/download/745/687).
- Middleton, E., Kandaswami, C. and Theonarides, T.C. (2000). The effects of Plant Flavonoids on Mammalian Cells: Implications for Inflammation, Heart Disease, and Cancer. *Pharmacological Reviews*, 52, 673 – 751.
- Ordoñez, A.A.L., Gomez, J.D., Vattuone, M.A. and Isla, M.L. (2006). Antioxidant activities of *sechiumedule*. *Food Chemistry*, 97, 452 – 458.
- Oyedemi, S.O., Bradley, G. and Afolayan, A.J. (2010). In-vitro and vivo antioxidant activities of aqueous extract of *strychnos henningsii* Gilg. Retrieved on June 4, 2014 from ACADEMICJOURNALS website: <http://www.academicjournals.org/ajpp>.
- Pearson, D. (1976). Laboratory techniques in food and analysis. London: Butterworth and Company Publishing Ltd.
- Saawan, K., Pradeep, S., Garima, M., Saurabb, S., Jha, K.K. and Khosa, R.L. (2011). Phytopharmacological review of *Alternanthera brasiliiana* (Amaranthaceae). *Asian Journal of Plant Science and Research*, 1(1), 41-47
- Snell, F.D. and Snell, C.T. (1953). Colorimetric methods of analysis. Vol. 3, p. 165 – 166. New York: D. Van Nostrand Company Incorporated.
- Sun, J.S., Tsuang, Y.W., Chen, J.J., Huang, Y.S. and Lu, F.J. (1998). An ultra-weak chemiluminescence study on oxidative stress in rabbits following acute thermal injury. *Burns*, 24, 225 - 231.
- Vladimir-Knezevic, S., Blazekovic, B., Stefan, M.B., Alegro, A., Koszegi, T. and Petrik, J. (2011). Antioxidant activities and polyphenolic contents of three selected *micromeria* species from Croatia. *Molecules*, 16 (2), 1454 - 1470
- WHO. (2003). Feeding and Commercialization of infants and young child guidelines for the WHO European region with emphasis on the former Soviet Union. European Series, No. 87, p. 1-296. Denmark: WHO Regional Publications
- WHO/UNICEF (2003). Global strategy for infant and young child feeding. WHO/UNICEF. Retrieved on March 3, 2012 from WHO Website: http://www.who.int/child_adolescent_health/documents/9241562218/en/index.html.
- Wolfe, K., Wu, X. and Liu, R.H. (2003). Antioxidant activity of apple peels. *Journal of Agriculture and Food Chemistry*, 51, 609 - 614
- Wong, P.K., Yusuf, S., Ghazali, H.M. and Man, Y.B.C. (2002). Physio-chemical characteristics of Roselle (*Hibiscus sabdariffa* L.). *Journal of Nutrition and Food Science*, 32, 68-73.
- Yen, G. and Chen, H. (1995). Antioxidant activity of various tea extract in relation to their antimutagenicity. *Journal of Agriculture and Food Chemistry*, 43, 7-32.
- Yi-Fang, C, Jie S., Xian-Hong, W.W and Rui-Hai, L. (2002). Antioxidant and Antiproliferative activities of common vegetables. *Review Journal of Agriculture and Food Chemistry*, 50, 6910-6916.